

# Targeting the Endocannabinoid System: From the Need for New Therapies to the Development of a Promising Strategy. What About Pancreatic Cancer?

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**Abstract.** Pancreatic cancer is one of the most fatal malignancies, and therefore, new strategies, which aim at the improvement of the prognosis of this lethal disease, are needed. Many clinical trials have failed to improve overall survival. Nowadays, research is focused on advances provided by novel potential targets to efficiently enhance life expectancy. Cannabinoids, the active components of *Cannabis sativa L.*, and their derivatives, have been reported as palliative adjuvants to conventional chemotherapeutic regimens. Cannabinoid effects are known to be mediated through the activation of cannabinoid receptors. To date, two cannabinoid receptors, cannabinoid receptor 1 and 2, have been cloned and identified from mammalian tissues. Cannabinoids exert a remarkable antitumoral effect on

pancreatic cancer cells, due to their ability to selectively induce apoptosis of these cells. This review strengthens the perception that cannabinoid receptors might be useful in clinical testing to prognose and treat pancreatic cancer. Many studies have tried to describe the mechanism of cell death induced by cannabinoids. The aim of this review is to discuss the effects of cannabinoid receptors in pancreatic cancer in order to provide a brief insight into cannabinoids and their receptors as pancreatic cancer biomarkers and in therapeutic strategies.

The plant *Cannabis sativa* L. produces approximately 60 unique compounds of bioactive lipids that have been categorized as cannabinoids. At the early stages of their discovery, it was speculated that cannabinoids induced physiological and behavioral actions *via* nonspecific interaction with cell membranes (1-3). Cannabinoids offer potential applications, such as being incorporated into chemotherapeutic regimens, in order to prevent nausea, pain and to stimulate appetite (4). Interestingly, cannabinoids seem to be specific, given that cancer cells can be targeted while normal cells can simultaneously be spared. Cannabinoids include compounds that have either structural, or pharmacological similarities to  $\Delta^9$ -tetrahydrocannabinol or bind to cannabinoid receptors (3, 5).

Although the knowledge concerning the pharmacology of a vast majority of the cannabinoids is poor, it is widely accepted that  $\Delta^9$ -tetrahydrocannabinol is the most important cannabinoid, as this compound is abundant in cannabis (6). Other relevant plant-derived cannabinoids include  $\Delta^8$ -tetrahydrocannabinol, which is almost as potent as  $\Delta^9$ -tetrahydrocannabinol, but it is found in smaller quantities. Tetrahydrocannabinol affects several biological functions by mimicking endogenous substances that activate cannabinoid receptors (5).

Cannabinoid receptors is a term to describe receptors which respond to cannabinoid drugs derived from *Cannabis sativa* and its biologically active synthetic analogs (7, 8). In particular, synthetic agonists that bind to cannabinoid receptors include  $\Delta^9$ -tetrahydrocannabinol-like analogs and aminoalkylindole compounds typified by R-WIN55212. Concerning synthetic cannabinoids which have been developed in scientific laboratories, they have structural similarities to both natural and endogenous cannabinoids, such as WIN-55, 212-2, JWH-133, and (*R*)-methanandamide (9). Many years of research led to the identification of several endogenous ligands of cannabinoid receptors, such as arachidonylethanolamide (anandamide), 2-arachidonoylglycerol and 2-arachidonoylglycerol ether (noladin ether). It is possible that there are also endogenous agonists other than these eicosanoid molecules. Exogenous agonists include synthetic agonists, such as CP55940, and plant-derived natural products, such as tetrahydrocannabinol (5).

At present, research has determined the existence of two cannabinoid receptor types, cannabinoid receptor 1 (CB1)

and cannabinoid receptor 2 (CB2), numbered in order of their discovery. CB1 and CB2 receptors differ in their predicted amino acid sequence, their signaling pathways and tissue distribution. Studies have shown that potent agonists and antagonists with marked selectivity for CB1 or CB2 receptors can be constructed. Likewise, CB1, CB2 and CB1/CB2 knockout mice can be developed (5, 8).

CB1 has been found in rat, mouse and human tissues and is identical in 97% to 99% of its amino acid sequence across species. Interestingly, *CB1* mRNA and protein are detected predominately in brain and neuronal tissue. In 1991, CB1 was first discovered in humans, and it was shown to be abundant in the brain. Moreover, it is expressed in some peripheral presynaptic nerve terminals and in a small number of extra-neural locations. CB1 receptor consists of a seven-transmembrane domain receptor. Its function is associated with biological responses and information about the structure-activity relationships of ligands for interaction with this receptor. Stimulation of the CB1 receptors suppresses the release of neurotransmitters, such as gamma-aminobutyric acid allowing the endocannabinoid system to reduce the activity of the gamma-aminobutyric acid terminal in a retrograde manner back across the synapse (11).

There is a 48% homology between CB1 and CB2. Mouse CB2 has been cloned and its sequence is 82% identical to that of human CB2. The mRNA of CB2 was first detected in immune tissue. Thus, it was thought that CB2 only existed in the immune system, but it has been described as being expressed in many cell types, such as natural killer cells, macrophages, monocytes,  $\beta$ -lymphocytes, microglial cells and inflammatory cells, in various organs of the human body, such as the brain and the gastrointestinal tract (5, 12). Notably, CB2 mRNA is missing from normal nervous tissue.  $\Delta^9$ -Tetrahydrocannabinol-like, aminoalkylindole and eicosanoid ligands are the basic ligands of expressed CB2 protein. CB2 triggers a sustained activation of ceramide biosynthesis (12, 13).

Both CB1 and CB2 are coupled to toxin-sensitive Gi/o proteins. Gi proteins are heterotrimeric GTP-binding proteins that inhibit plasma membrane adenylyl cyclase. Binding to an agonist with CB1 or CB2 leads to a signaling cascade in which the activated receptor activates Gi, which in turn inhibits adenylyl cyclase, resulting in a reduction of intracellular cyclic adenosine monophosphate (cAMP). Essentially, the classical theory of receptor occupancy and activation is a conformational induction theory that fits equivalently with Koshland's induced-fit model for enzymes. In particular, when agonist A binds to the resting-state receptor R, conformational changes are provoked, leading to the activation of receptor R\* (5, 14).

Moreover, CB1 is coupled through Gi/o proteins to a variety of potassium and calcium channels. As far as novel types of cannabinoid receptors are concerned, there is some

Table I. Studies focused on cannabinoids as potential agents against pancreatic cancer as classified by different experimental systems, cannabinoid receptors, and their effects.

Study (Ref)	Experimental system	Effect	Receptor
Carracedo <i>et al.</i> , 2006 (18)	<i>In vitro</i> (MiaPaCa-2, Panc1 cells)	<ul style="list-style-type: none"> <li>• Ceramide-dependent up-regulation of stress-related genes</li> <li>• Apoptosis</li> </ul>	CB2
Carracedo <i>et al.</i> , 2006 (19)	<i>In vitro</i> (MiaPaCa-2, Panc1 cells)	<ul style="list-style-type: none"> <li>• Overexpression in these cell lines compared with normal pancreatic tissue</li> </ul>	CB1, CB2
Michalski <i>et al.</i> , 2006 (61)	<i>In vitro</i> (human tissue)	<ul style="list-style-type: none"> <li>• Inverse association with survival rate of pancreatic ductal adenocarcinoma)</li> </ul>	CB1
Brandi <i>et al.</i> , 2009 (71)	<i>In vivo</i> (human) <i>In vitro</i> (Panc1 cells)	<ul style="list-style-type: none"> <li>• Inverse correlation with cancer pain symptoms</li> <li>• Collapse of the keratin cytoskeleton</li> <li>• Cell death</li> </ul>	CB1, CB2 CB2
Donadelli <i>et al.</i> , 2011 (66)	<i>In vitro</i> (Panc1 cells) <i>In vitro</i> (PaCa44, PaCa3, Panc1, CFPAC1, T3M4 and MiaPaCa2 cell lines)	<ul style="list-style-type: none"> <li>• Antiproliferative effects</li> <li>• Inhibition of pancreatic adenocarcinoma cell growth</li> <li>• Increase of ROS</li> <li>• Inhibition of apoptosis induced by gemcitabine</li> <li>• Cell death</li> <li>• Cytotoxicity</li> </ul>	CB1, CB2 CB1, CB2
Fogli <i>et al.</i> , 2013 (60)	<i>In vitro</i> (MiaPaCa-2 cells) <i>In vitro</i> (MiaPaCa-2 cells)	<ul style="list-style-type: none"> <li>• Increase of AMPK phosphorylation</li> <li>• Autophagy</li> <li>• Increase of ROS production</li> </ul>	CB1 CB1, CB2
Dando <i>et al.</i> , 2013 (80)	<i>In vitro</i> (Panc1 cells)	<ul style="list-style-type: none"> <li>• Increase of AMPK phosphorylation</li> <li>• Autophagy</li> <li>• Increase of ROS production</li> <li>• Increase of tumor-suppressor PTEN</li> </ul>	CB1, CB2
Guo <i>et al.</i> 2018 (62)	<i>In vivo</i> (mouse) <i>In vitro</i> (Panc1 cells)	<ul style="list-style-type: none"> <li>• Binding of fluorescent substance NIR760-XLP6 to CB2 and earlier imaging of the cancer</li> <li>• Binding of fluorescent substance NIR760-XLP6 to CB2 and earlier imaging of the cancer</li> </ul>	CB2 CB2
Yasmin-Karim <i>et al.</i> , 2018 (88)	<i>In vitro</i> (Panc-02 cells) <i>In vivo</i> (mouse) (simultaneous use of smart radiotherapy materials and cannabinoids)	<ul style="list-style-type: none"> <li>• Apoptosis</li> <li>• Increase of ROS production</li> <li>• DNA damage through radiation</li> <li>• Inhibition of tumor growth</li> <li>• Induction of apoptosis</li> <li>• Improvement of survival</li> <li>• Induction of apoptosis</li> <li>• Reduction of cell viability</li> <li>• Reduction of cell migration</li> <li>• Increased chemosensitivity</li> <li>• Induction of chemotoxicity</li> <li>• Induction of cytotoxicity, especially with ALAM108</li> <li>• Weight gain</li> </ul>	NR NR
Luongo <i>et al.</i> , 2020 (63)	<i>In vitro</i> (Panc1 cells and MiaPaCa-2), co-administration of O <sub>2</sub> /O <sub>3</sub>	<ul style="list-style-type: none"> <li>• Induction of apoptosis</li> <li>• Reduction of cell viability</li> <li>• Reduction of cell migration</li> <li>• Increased chemosensitivity</li> <li>• Induction of chemotoxicity</li> <li>• Induction of cytotoxicity, especially with ALAM108</li> </ul>	CB1, CB2
Aizikovitch, 2020 (68)	<i>In vitro</i> (PANC-1 and AsPC-1 cell lines) <i>In vivo</i> (human pancreatic tumor cells xenografted in nude female mice)	<ul style="list-style-type: none"> <li>• Induction of cytotoxicity, especially with ALAM108</li> <li>• Weight gain</li> <li>• Reduction of tumor volume and size</li> <li>• Reduction of interaction between pancreatic cells and stromal cells</li> <li>• Increase of anticancer immunity</li> <li>• Inhibition of cell proliferation</li> <li>• Suppression of pancreatic tumor</li> </ul>	NR NR
Yang <i>et al.</i> , 2020 (83)	<i>In vitro</i> (pancreatic cell lines) <i>In vivo</i> (mice)	<ul style="list-style-type: none"> <li>• Reduction of tumor volume and size</li> <li>• Reduction of interaction between pancreatic cells and stromal cells</li> <li>• Increase of anticancer immunity</li> <li>• Inhibition of cell proliferation</li> <li>• Suppression of pancreatic tumor</li> </ul>	NR NR

AMPK: AMP-activated protein kinase; CB1: cannabinoid receptor 1; CB2: cannabinoid receptor 2; NR: not reported; PTEN: phosphatase and tensin homologue; ROS: reactive oxygen species.

preliminary evidence for their existence based on multiple criteria such as primary structure homology, signal transduction mechanisms and biological functions (5). Additionally, in 2007, Ryberg *et al.* reported a new cannabinoid receptor called G protein-coupled receptor 55 (GPR55) which binds to and is activated by CP55940 with

potential action on the brain (15). Shi *et al.* confirmed this action, explaining how GPR55 mediates anxiolytic-like effects in the medial orbital cortex of mice (16). This effect in conjunction with analgesia, appetite regulation and antiemetic actions are referred to as palliative methods for symptoms of malignancies, with potential use as a new

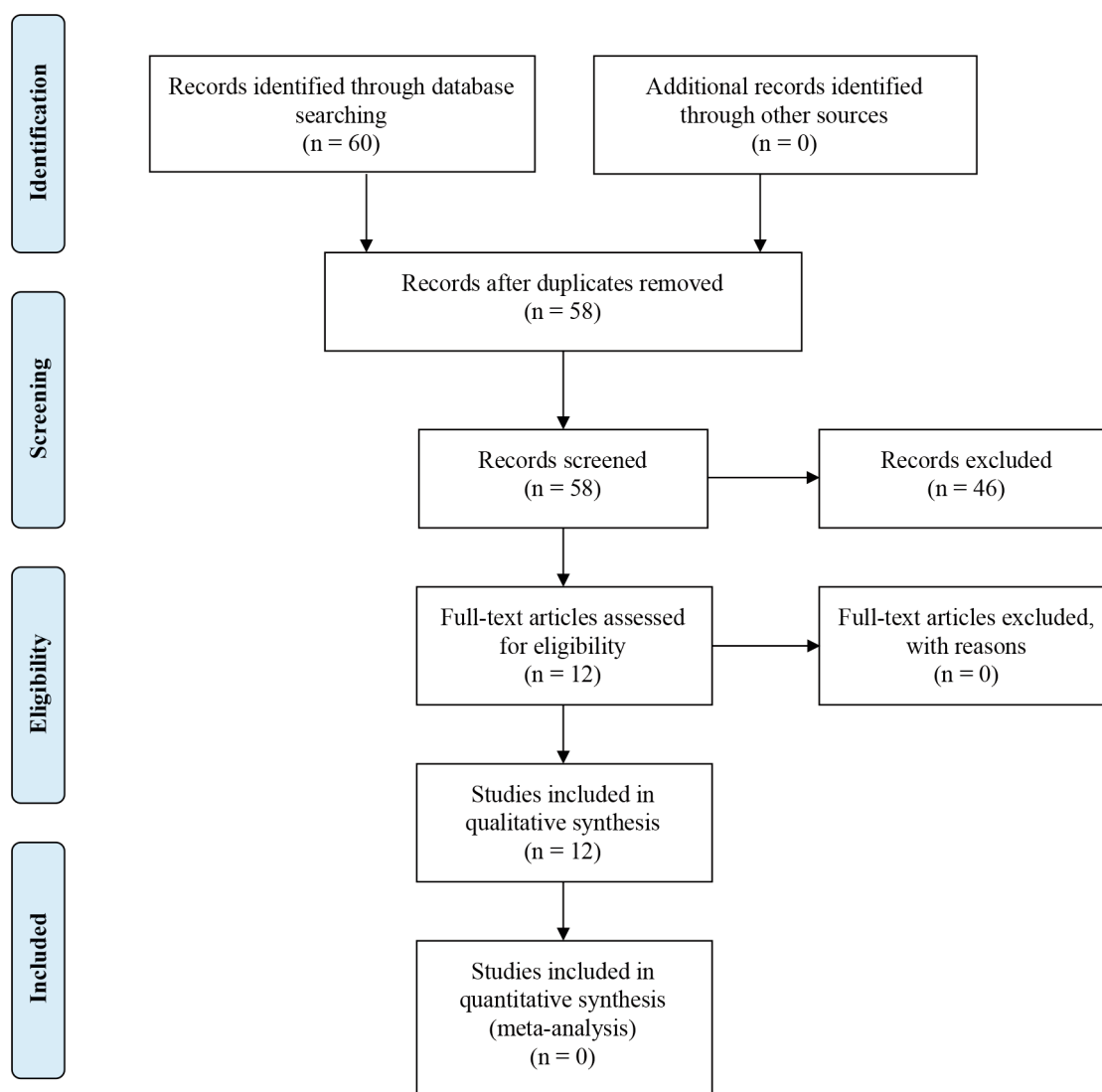


Figure 1. PRISMA flow chart for the current study.

treatment of pancreatic cancer and that is discussed extensively below.

## Materials and Methods

A review of the literature was conducted through the use of the PubMed database and Cochrane library in order to identify articles regarding *in vitro* and *in vivo* studies of cannabinoid use in pancreatic cancer. Specifically, our research was performed by using key words, separately and in various combinations, such as cannabinoid, receptor, cancer, pancreatic and treatment. Furthermore, we checked the references from all articles found aiming to include any other eligible studies. The initial research identified 60 articles. After removal of duplicates, 58 remained. The remaining articles were screened and 46 were excluded for various reasons, *e.g.*, some of them were only abstracts, whereas others were not completely

relevant to the topic. Publications in non-English language were also excluded. Finally, 12 full-text articles were included (Table I). The inclusion process is shown in Figure 1.

## Results

**Cannabinoids against cancer.** Cancer is a group of diseases characterized by abnormal cell growth and their potential to invade or spread. This uncontrolled division is caused by DNA mutations and damage, cell-cycle perturbations, and changes in apoptotic mechanisms. Thus, cancer treatment decisions rely on genetics and clinical pharmacology. In particular, molecules that modulate apoptosis in order to maintain steady-state cell growth can be beneficial agents for targeted cancer treatment. These modifications affect

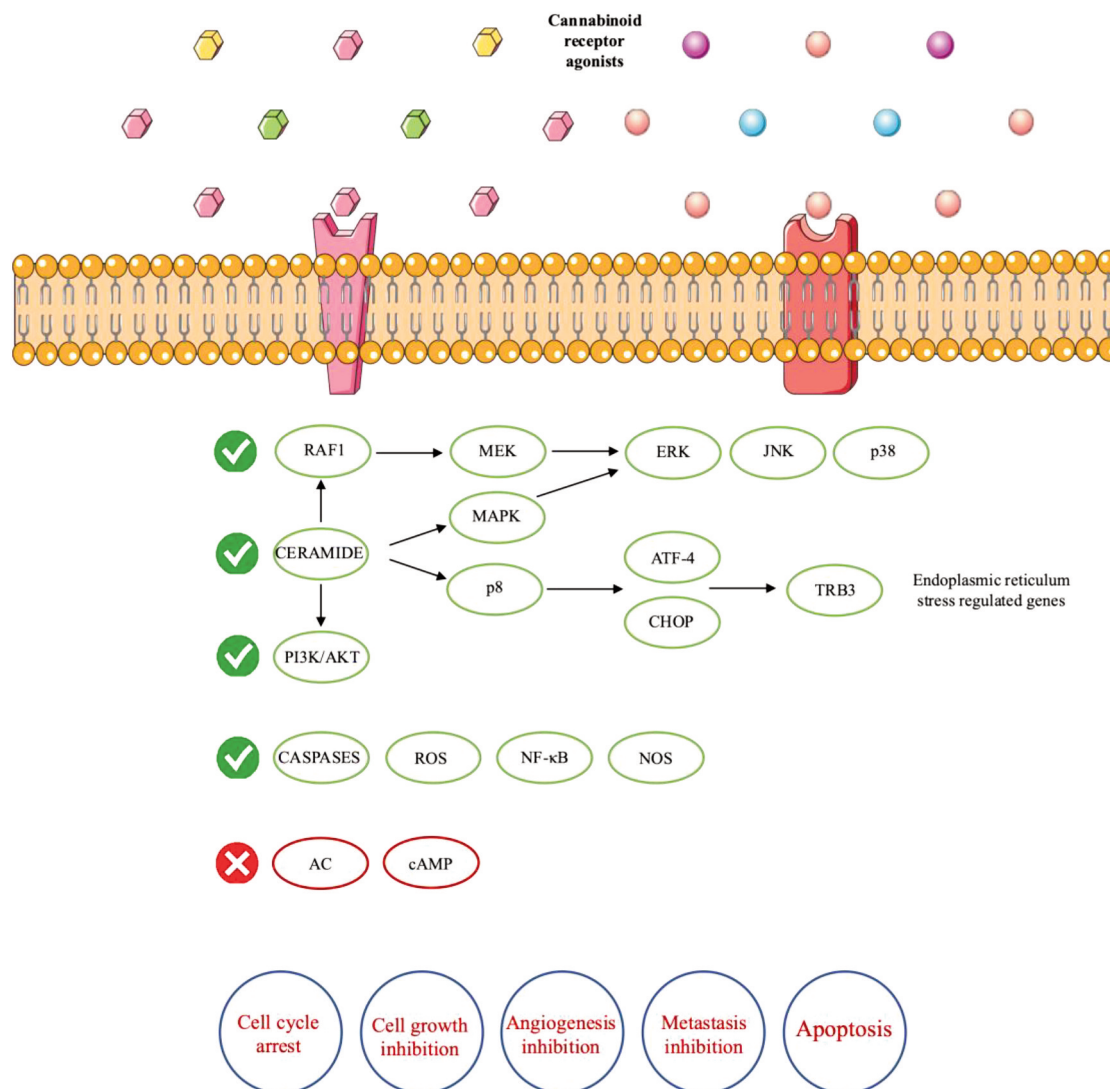


Figure 2. Mechanism of actions and anticancer effects of cannabinoid receptors. AC: Adenylyl cyclase; AKT: protein kinase B also known as AKT; ATF-4: Activating transcription factor 4; cAMP: cyclic adenosine monophosphate; CHOP: C-homologous protein; ERK: extracellular-signal-regulated kinase; JNK: c-Jun N-terminal kinase; MAPK: mitogen-activated protein kinase; MEK: mitogen-activated protein kinase kinase; NF-κB: nuclear factor kappa-light-chain-enhancer of activated B-cells; NOS: nitric oxide synthase; PI3K: phosphatidylinositol-4,5-bisphosphate 3-kinase; ROS: reactive oxygen species; TRIB3: Tribbles homolog 3.

signaling intermediates in the apoptotic cascade to induce apoptosis. Hence, given the incidence and the severity of these diseases, novel therapeutic targets need to be identified in order to combat cancer. Research into the molecular basis of cancer shows that cannabinoid receptors are promising targets.

The antiproliferative potential of cannabinoids was first reported in the early 1970s by Munson *et al*. It was shown that oral administration of tetrahydrocannabinol in mice inhibits Lewis lung adenocarcinoma tumor growth (17). Later, in the 1990s, studies reported that either endogenous,

or synthetic cannabinoids induce antitumor effects on a wide spectrum of tumor cells in culture. In particular, it was found that in several types of cancer cells such as glioma, lymphoma, prostate, breast, pancreatic and skin cancer cells, cannabinoids reduced tumor size and induced apoptosis (9, 18-22). Studies in animal tumor models, such as Wistar rats inoculated with C6 gliomas, and xenografts in athymic nude mice implanted with KiMol or MBA-MD-231 breast cancer cells, found that cannabinoid receptor agonists exerted antiproliferative actions (22). As a variety of biochemical and pharmacological approaches showed, these antitumor



effects require the efficiency of CB1 and CB2. Elective cannabinoid receptor agonists and antagonists, which ligate to CB1 and CB2, can be used in order to study their expression and their mode of action (apoptotic or stimulatory) in malignant cells. However, at certain doses and in specific cellular contexts, some cannabinoids have been reported to induce the proliferation of cancer cells *in vitro* (23).

As was mentioned above, cannabinoid receptors are activated by specific G-protein coupling with cannabinoids. This binding induces several cellular pathways, as cAMP–protein kinase A pathway is inhibited and the activity of  $\text{Ca}^{2+}$  and  $\text{K}^{+}$  channels is modulated. These changes lead to the inhibition of neurotransmitter release. Moreover, CB1 participates in a variety of different cellular signaling pathways that affect the control of cell fate. In particular, CB1 receptor coupling urges the activation of mitogen-activated protein kinase (MAPK) cascades, such as the extracellular-signal-regulated kinase (ERK), the stress-activated kinases Jun amino-terminal kinase (JNK) and p38 MAPK (25-29). These signaling cascades exert dominant roles in the regulation of cell proliferation and differentiation. Cannabinoid-induced MAPK stimulation has been reported in neural cell lines, primary neural cells, lymphoid cells, vascular endothelial cells, and Chinese hamster ovary cells that were transfected with cannabinoid receptor complementary DNAs (10, 24-26, 28). On the other hand, an *in vitro* study in a neuronal-like cell line showed that CB1 receptor activation attenuates ERK (29).

Furthermore, cannabinoid receptors participate in the activation of the phosphatidylinositol 3-kinase (PI3K)-AKT serine/threonine kinase 1 (AKT) survival pathway (30-32). Particularly, the phosphorylation of AKT induces the inhibition of nuclear translocation of forkhead transcription factors (33, 34). As a result, the expression of pro-apoptotic proteins is prevented. However, a study reported that cannabinoid receptors negatively regulated AKT activation. Some systems showed that PI3K is an upstream component of cannabinoid-induced ERK activation, while others reported contradictory results (35, 36). Moreover, some conflicting data claim that nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B) and nitric oxide synthase (NOS) are other targets of CB1 activation, which are involved in the control of cell fate. Nevertheless, it is not clear if these components are activated or inhibited by cannabinoids (30, 37).

Cannabinoid receptors are also involved in the modulation of sphingolipid-metabolizing pathways. CB1 activation induces sphingomyelin breakdown, resulting in an increase in ceramide levels (38, 39). Ceramide is a lipid second messenger that stimulates apoptosis and cell-cycle arrest. Interestingly, these actions are not based on the G-protein structure, but they are cannabinoid receptor-dependent. It is

reported that an adaptor factor associated with neutral sphingomyelinase activation (FAN) is involved (40). Furthermore, cannabinoid receptor activation has the potential to urge a sustained peak of ceramide expression through enhanced *de novo* synthesis (32, 39).

Overall, cannabinoid receptors are implicated in antitumor effects by a variety of different mechanisms, as their activation stimulates transformed-cell death, inhibits transformed-cell proliferation and limits tumor angiogenesis and metastasis. As mentioned above, apoptosis is induced by *de novo* ceramide accumulation, sustained ERK activation and AKT inhibition (34, 41) (Figure 2). In glioma cells, ceramide levels are increased after cannabinoid receptor activation (41). This increase induces both the activation of RAF1–mitogen-activated protein kinase kinase (MEK)–ERK signaling cascade and the AKT inhibition. It is known that ERK activation stimulates cell proliferation. In any case, many factors can affect the relation between ERK activation and cell fate, given that prolonged ERK stimulation mediates cell-cycle arrest and even cell death (42). Mimeault *et al.* reported that in prostate tumor cells, pharmacological inhibition of *de novo* ceramide synthesis led to prevention of cannabinoid-induced cell death (43).

A study on breast cancer cells found that CB1 receptor activation induced cell-cycle arrest at the  $\text{G}_1$ -S transition by inhibiting adenylyl cyclase and the cAMP signaling cascade. Given that the phosphorylation of protein kinase A reduces RAF1 expression, cannabinoid receptors block the inhibition of RAF1. As a result, the RAF1–MEK–ERK pathway is stimulated (44). Corresponding research in thyroid epithelioma cells transformed with the Kirsten rat sarcoma (*KRAS*) oncogene showed that CB1 receptor activation also blocks the cell cycle at the  $\text{G}_1$ -S transition (45). However, the underlying mechanisms of cannabinoid action on the cell cycle remain obscure. Similar results of cell-cycle arrest following cannabinoid receptor activation have been reported in skin and prostate cancer cells. In these studies, growth-factor-receptor signaling is inhibited (43, 46). It is likely that a general underlying mechanism of cannabinoid antitumor action exists.

Angiogenesis is a very important process that allows tumors to grow beyond a minimal size. Therefore, the prevention of the angiogenic process is an extremely promising approach in order to eradicate tumors (47). Studies in mice with glioma or skin carcinoma showed that cannabinoid administration induced the modification of vascular hyperplasia into blood vessels that are characterized by small and differentiated capillaries (46). These modifications are associated with reduced expression of pro-angiogenic cytokines, such as vascular endothelial growth factor (46-48). Moreover, in vascular endothelial cells, cannabinoid receptor stimulation restrained cell migration and survival. In mice injected with lung cancer

cells, cannabinoid receptor activation inhibited tumor metastasis (48). This might be a result of diminished activity and expression of matrix metalloproteinase 2, an enzyme that induces tissue proteolysis during angiogenesis and metastasis (46).

It is understandable that cannabinoids have been reported as useful antitumor agents in animal cancer models. Concerning toxicity, studies claim that cannabinoids have a good safety profile and offer relief to patients with cancer. However, their antitumor activity in humans has not been demonstrated. The Spanish Ministry of Health has approved a phase I clinical trial which aims at investigating the effect of administration of  $\Delta^9$ -tetrahydrocannabinol on the growth of recurrent glioblastoma multiforme. Guzman *et al.* found  $\Delta^9$ -tetrahydrocannabinol to have antiproliferative action on tumor cells (49). However, Massi *et al.* reported that cannabidiol treatment led to stimulation of apoptosis in glioma cells. Moreover, they showed that the same treatment eliminated tumor growth through activation of caspases and reactive oxygen species (ROS) (50). Although the mechanisms of action of cannabinoids remain obscure, it is clear that cannabinoid receptors have a prominent role in treatment of cancer.

**Cannabinoids against pancreatic cancer.** Pancreatic cancer has evolved as one of the deadliest diseases. Pancreatic cancer accounts for about 3% of all cancer in the United States and about 7% of all cancer-related deaths (51). According to the Surveillance Epidemiology and End Results program, the 5-year survival rate of patients with pancreatic cancer is 4%. This figure is the lowest of all cancer types (52, 53). However, the high mortality rate is a result of limited early diagnosis, ineffective chemotherapy, and poor radiotherapy response. Moreover, 50% of patients with early pancreatic cancer do not show symptoms. After the diagnosis, the 4-year survival rate can rise to 78% after resection of pancreatic cancer with a size smaller than 2 cm (54). The standard chemotherapy for pancreatic cancer is gemcitabine (55). Unfortunately, the efficacy of this current therapy is limited due to the complicated tumor microenvironment, which hinders efficient drug delivery to the cell target (56). Therefore, the scientific community is trying to target the molecular mechanisms of pancreatic cancer (57, 58). For instance, activation of KRAS oncoprotein, inactivation of p16<sup>INK4A</sup>, overexpression of cyclo-oxygenase-2 and loss of p53 effects are the main targets of the molecular pathology of pancreatic cancer (53, 54, 59). Recent therapeutic strategies include combinations of these compounds and gemcitabine in order to enhance the efficacy of pancreatic cancer treatment (54, 57).

An *in vitro* study investigated the antitumor effects of CB receptors and their potential role in the human pancreatic cancer cell line MIA PaCa-2 (60). According to this study,

cannabinoid derivatives were cytotoxic *via* a receptor-independent mechanism. In particular, both selective CB1 and CB2 receptors agonists and antagonists induced a significant cytotoxic effect. The absence of CB2 receptors clearly prevented a potential role of these receptors in cannabinoid cytotoxicity. Moreover, CB1 receptor agonist and antagonist, at nanomolar concentrations, did not significantly alter cell viability in serum-free medium, which is a condition that avoids non-specific interactions with serum components. Given that cannabinoids are lipophilic components, it is understandable that cytotoxicity can arise by their crossing the cell membrane and rupturing cellular networks, resulting in cell death. Internucleosomal degradation of DNA by CB1 antagonist *N*-(piperidin-1-yl)-5-(4-iodophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (AM251) showed that apoptosis might contribute to its antitumor effect. AM251 also induced a significant time-response action on caspase 3/7, indicating that these two enzymes of apoptosis were a target of cannabinoids. It is possible that apoptosis might be mediated *via* a caspase-dependent pathway. The gene-expression profile of MIA PaCa-2 cells after AM251 treatment was examined by microarray analysis and AM251 was found to act on cell cycle-related pathways, as well as on critical steps in cell proliferation including Janus kinase (JAK)/signal transducers and activators of transcription (STAT) and MAPK signaling cascades. We should mention that in that study, AM251 was found to synergistically enhance the anticancer activity of 5-fluorouracil, which is a very common agent used in the treatment of pancreatic cancer (57). Hence, AM251 is highlighted as a novel compound for pancreatic cancer therapy (60).

In recent years, there has been increasing interest in cannabinoids as a therapeutic regiment against pancreatic cancer (18, 19, 60). About 95% of clinical cases of pancreatic cancer are ductal adenocarcinomas. Michalski *et al.* studied immunoreactivity for CB1 and CB2 receptors, as well as for the endocannabinoid metabolizing enzymes fatty acid amide hydrolase (FAAH) and monoacyl glycerol lipase (MGLL) in human tissues of pancreatic ductal adenocarcinoma. Interestingly, significant variation was reported in receptors, and FAAH and MGLL levels between healthy people and patients suffering from pancreatic cancer (61). As CB1 and CB2 receptors levels were up-regulated in pancreatic cancer, it is assumed that cannabinoid receptors have dominant role in pancreatic carcinogenesis. We should mention that in their study, endocannabinoid levels in pancreatic cancer tissues did not change, possibly due to the strong desmoplastic reaction in pancreatic cancer. Researchers found an inverse association between CB1 receptor quantity and survival and an association between FAAH or MGLL levels in cancer cells and survival rate. On the contrary, other studies claim that cannabinoids are

involved in antiproliferative activities on pancreatic cancer growth. In any case, the heterogeneous expression patterns of cannabinoid receptors indicate their potential use as clinical biomarkers (18, 60, 62). Guo *et al.* showed that CB2R is overexpressed in both human pancreatic adenocarcinoma tissues and cell lines. Fluorescent substances such as NIR760-XLP6 binding to CB2R can permit early recognition of pancreatic cancer tissue and earlier diagnosis (62). Michalski *et al.* also reported that cannabinoid receptors may be associated with the development of pancreatic cancer pain, as both CB1 and CB2 receptor expression were inversely correlated with pain symptoms of patients. Nevertheless, given that there was no association between FAAH/MGLL levels and pain scores, endocannabinoid inactivation through FAAH or MGLL is probably not involved in pancreatic cancer pain (61).

Other studies investigated cannabinoid receptors in pancreatic cancer both *in vitro* and *in vivo* (18, 63). Researchers showed that cannabinoid receptors are more highly expressed in human pancreatic tumor cell lines and tumor biopsies than in normal pancreatic tissue. *In vitro* studies in MiaPaCa2 and Panc1 cell lines reported that cannabinoid administration led to apoptosis, reduced cell viability and augmented the levels of ceramide and caspase 3. The combination of cannabinoid and O<sub>2</sub>/O<sub>3</sub> induced cytotoxicity and augmented chemosensitivity to gemcitabine and paclitaxel (63). Moreover, mRNA levels of the p8 protein, which is associated with stress, were up-regulated (19). Interestingly, when CB2 receptor was blocked, all the above effects were prevented (18). These results agree with other observations that had shown that CB2 receptor is associated with the anticancer effect of cannabinoids in gliomas and lymphomas, as well as skin and prostate carcinomas (9, 46, 64, 65). CB2-selective activation is not related to the typical marijuana-like psychoactive effects that we meet in CB1 receptor activation (5). Researchers found that CB2 receptor-dependent accumulation of *de novo*-synthesized ceramide induced p8 up-regulation. p8 was involved, *via* its down-stream endoplasmic reticulum stress-related targets, in the stimulation of activating transcription factor 4 (ATF-4) and proapoptotic protein TRB3 in apoptosis stimulated by  $\Delta^9$ -tetrahydrocannabinol. Furthermore, study in tumor xenografts showed that cannabinoid treatment reduced tumor growth and inhibited the spreading of pancreatic cancer cells. Similarly, to the *in vitro* studies, apoptotic death and increase of TRB3 expression was observed in pancreatic tumor cells, while there were no changes in normal tissue (18). Hence, CB2 receptor is reported to induce apoptosis of pancreatic tumor cells, setting the basis for a novel therapeutic approach for the treatment of pancreatic malignancy.

A recent study investigated the correlation of gemcitabine with the synthetic cannabinoids arachidonylcyclopropamide

(ACPA) or W405833 (GW) (66). ACPA is a highly selective agonist for CB1 receptor, while GW is a selective agonist for CB2 receptor (67). Gemcitabine induced both CB1 and CB2 receptors *via* a nuclear factor- $\kappa$ B (NF- $\kappa$ B)-dependent molecular mechanism. The co-treatment prevented pancreatic adenocarcinoma and cell growth, as it induced ROS production. As a result, endoplasmic reticulum stress and autophagic cell death were provoked. Free radical scavenger *N*-acetyl-cysteine and the specific NF- $\kappa$ B inhibitor BAY 11-7085 prevented the antitumor synergism, indicating that the stimulation of ROS by gemcitabine and cannabinoid co-treatment, and of NF- $\kappa$ B by gemcitabine were essential for these actions. In human pancreatic tumor cells xenografted in nude mice, combined treatment significantly prevented tumor growth. Similar effects on the tumor size and volume were observed in another study, in which ALAM027 and ALAM108, two new cannabinoid derivatives were used in human pancreatic female mice bearing tumor cell xenografts. In the same study, ALAM108 demonstrated higher cytotoxicity at lower concentrations against pancreatic cancer cell lines (68). Furthermore, cannabinoid treatment activated transcriptional factor X-box-binding protein-1 (XBP-1) splicing, glucose-regulated protein 78 (GRP78) and CCAAT/enhancer-binding protein (C/EBP) as well as homologous protein (CHOP) gene induction (69, 70). These genes are the molecular switch that stimulate autophagic or apoptotic cell death signals. It is noteworthy that these endoplasmic reticulum stress-related genes were also enhanced by gemcitabine. This evidence shows that these genes might be involved in gemcitabine/cannabinoid co-treatment. We should mention that according to Donadelli *et al.*, although ACPA or GW induced cell-cycle arrest at the G<sub>1</sub> phase, they did not induce apoptosis. In addition, cannabinoid treatment partially, but significantly, inhibited apoptosis stimulated by gemcitabine (66). These data are contradictory to those of Carracedo *et al.* but it is possible that this discrepancy is a result of the different cannabinoid receptor agonists that were used (18).

Another study of Panc1 cells showed that treatment for 12 h with increasing amounts ACPA or GW had antiproliferative effects. GW inhibited tumor growth significantly and more intensively than ACPA. Moreover, treatment of Panc1 cells with cannabinoids led to the down-regulation of phosphorylated alanine aminopeptidase (ANPEP) (71). ANPEP is a zinc-dependent metallo-exopeptidase found on the surface of tumor cells. ANPEP degrades the extracellular matrix and, as a result, it induces tumor invasion, angiogenesis and metastasis (72). It constitutes a transcriptional target of RAS signaling pathways. ANPEP inhibitors have been reported to be effective anticancer agents (73). It is noteworthy that ANPEP has been proposed as a novel prognostic biomarker for patients with pancreatic cancer (74).



Proteomic investigation showed that cannabinoid treatment induced several isoforms of phospho-keratins 18 (KRT18) (SSP6411, SSP6402, SSP6410) (71). KRT18 is a filament protein associated with mitosis, apoptotic death, signaling pathways and invasion. Phosphorylation of KRT18 at Ser52 is necessary for filament reconstruction at cell-cycle arrest, as during apoptosis; intermediate filaments reorganize into granular structures enriched for phosphorylated KRT18 (75). Phosphorylation of KRT19 at Ser33 is associated with 14-3-3 proteins but it does not seem essential for apoptosis (76). GW strongly induced KRT18 phosphorylation both at Ser52 and Ser53, while total KRT18 expressed was down-regulated (71). Presumably, CB2 receptor activation, by augmenting KRT18 phosphorylation, induces the collapse of the keratin cytoskeleton and cell death (77).

AMP-activated protein kinase (AMPK) plays an important role in autophagy, as it inhibits mammalian target of rapamycin complex 1 (mTORC1), the major regulator of cell growth (78). AMPK is a crucial molecule in the autophagic pathway induced by cannabinoids (79). Dando *et al.* found that ACPA and GW synthetic cannabinoids induced the activation of AMPK in pancreatic adenocarcinoma cells, as they increased the cellular AMP/adenosine triphosphate (ATP) ratio (80). Cannabinoid treatment reduced the phosphorylation level of p70S6K, a direct target of mTORC1, as well as increasing AMPK phosphorylation. Furthermore, AMPK is reported to be an intracellular energy sensor and plays an active role in maintaining intracellular homeostasis during stress challenges (81). For instance, oxidative stress has been found to induce AMPK (82). ROS were essential for the increase in AMP/ATP ratio, which in turn led to the activation of AMPK by cannabinoids and as a result to autophagy. Their study reported that treatment with either ACPA or GW elevated the nicotinamide adenine dinucleotide (NADH) level and this increase was prevented in presence of the radical scavenger *N*-acetyl-L-cysteine. These results suggest that ROS production induced by cannabinoids might disrupt the electron transport chain, which in turn inhibits the Krebs cycle, leading to NADH accumulation and inhibition of oxidative phosphorylation, which could further elevate the ROS level (80). The p-21-activated kinase 1 mediated pathway is responsible for the anticancer effects of cannabinoids. Yang *et al.* conducted a study in which they tested the antitumor effects of cannabinoids in cell lines and mouse models. The p-21 pathway is particularly related to the actions of the KRAS pathway. Cannabinoids act through the inhibition of these pathways. Thus, the interaction between pancreatic cells and pancreatic stromal cells was reduced, leading to reduction of the proliferation of cancer cells and of tumor growth. Furthermore, antitumor immunity was restored through the down-regulation of programmed death-ligand 1 (PD-L1), leading to a better immune response and reduction of tumor size (83).

Concomitantly, endocannabinoid treatment increased expression of phosphatase and tensin homolog (PTEN), which is a tumor suppressor mutated in different types of human cancer. PTEN is a major regulator of AKT, which, in turn, up-regulates c-MYC (84). Recently, it has been shown that c-MYC is involved in the transcription of genes that lead to the expression of kinase isoform M2 (PKM2) (85, 86). PKM2 is expressed in cancer and promotes aerobic glycolysis (85, 87). Cannabinoids were shown to determine the down-regulation of PKM2, *via* the inhibition of c-MYC (80). Overall, in pancreatic adenocarcinoma cells treated with cannabinoids, autophagy that was induced by cannabinoids is highly associated with inhibition of energetic metabolism, which, in turn, is related to cannabinoid-dependent ROS production.

All the studies mentioned above demonstrated different oncogenic pathways which can be affected through cannabinoid receptors. The action of cannabinoids might be potentiated by the use of other chemotherapeutic agents or modalities in order to exert a synergistic antitumor activity (65). Yasmin-Karim *et al.* showed in both *in vitro* and *in vivo* studies that the use of cannabinoids against pancreatic malignancy can also be potentiated through the use of radiation and can prolong survival (88). As a result, their simultaneous use with chemotherapy and radiation can probably reduce the side-effects of both and improve the clinical outcome. All these studies offer promising results in the field of pancreatic cancer treatment. Thus, further research should take place in order to explain the mechanisms of action of cannabinoids and prove their clinical significance against pancreatic malignancy, especially in combination with other therapeutic methods.

## Conclusion

In recent decades, the genetic and pharmaceutical manipulation of the cannabinoid system has gained significant interest in the emergence of new drugs and their actions, both in cancer and various diseases. Natural and synthetic analog of the marijuana plant *Cannabis sativa L.* affect several human diseases such as neurological, cardiovascular, and autoimmune disorders, pain, inflammation, and diabetes as well as cancer. As a result, the medicinal use of cannabis has potential against a variety of diseases; the US Food and Drug Administration has already approved the application of certain relative drug regimens (89).

Several effects of cannabinoids are mediated through the activation of cannabinoid receptors. Two cannabinoid receptors have been identified and cloned from mammalian tissues: the 'central' CB1 receptor and the 'peripheral' CB2 receptor, while a few more are under investigation. These receptors and their antagonists are proving to be meaningful as they are able to have a targeted impact on cancer cells whilst sparing normal cells. The differential expression of

CB1 and CB2 receptors in different cell lines is justified by the variation of the effects of cannabinoids. Low expression of cannabinoid receptors may induce cell proliferation and metastasis, as the antitumor immune response is suppressed, while overexpression of cannabinoid receptors could have antitumor effects.

Concerning pancreatic cancer, the cannabinoid system does not seem to play a relative role in cell proliferation. However, it was found that the CB1 receptor antagonist AM251 might be an important compound for the development of promising diarylpyrazole derivatives active against pancreatic cancer (60). Moreover, cannabinoid administration induced apoptosis of pancreatic tumor cells. In particular, apoptosis was a result of CB2 receptor activity and ceramide-dependent up-regulation of p8, ATF-4 and TRB3 stress-related genes (18). It is understandable that the observed effects of these two studies are contradictory. However, it is clear that cannabinoids have therapeutic potential for the treatment of pancreatic cancer and probably diagnostic value. Unfortunately, we cannot completely explain these effects, as the knowledge about the mechanism of action of cannabinoid receptors is limited.

In any case, there is evidence that cannabinoid receptors should be investigated as chemotherapeutic agents either as combination therapy boosting palliative treatment, or as monotherapy. In future clinical studies, researchers should evaluate different combinations of cannabinoid agents along with chemotherapeutic regimens and clarify the optimum ratio in order to achieve down-regulation of malignant cells and avoid tumor cell proliferation. Hence, further studies are required to elucidate the exact mechanism of action of cannabinoid receptors and their possible future potential against pancreatic cancer. After all, the clarification of the aforementioned factors and the creation of more evidence-based data are preconditions for the approval of such cannabinoid-based medicinal products by the Food and Drug Administration.

## Conflicts of Interest

All the Authors declare that there are no conflicts of interest.

## Authors' Contributions

Nikolaos Garmpis, Christos Damaskos and Dimitrios Dimitroulis designed the study. Nikolaos Garmpis, Christos Damaskos, Dimitrios Dimitroulis and Anna Garmpi wrote the article. Nikolaos Garmpis, Christos Damaskos, Anna Garmpi, Evangelos Diamantis, Panagiotis Sarantis, Vasiliki E. Georgakopoulou, Alexandros Patsouras, Athanasios Syllaios, Georgios Kyriakos, Evangelos Koustas, Christos Vallilas and Petros Papalexis collected the data. Christos Damaskos, Panagiotis Sarantis, Efstathios A. Antoniou, Konstantinos Kontzoglou and Dimitrios Dimitroulis offered scientific advice. Dionysios Prevezanos and Markos Despotidis revised the article. Gregory Kouraklis critically revised the article and was the supervisor.

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