

PD-1 Expression and its Correlation With Prognosis in Clear Cell Renal Cell Carcinoma

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Abstract. *Background/Aim:* Programmed death ligand-1 (PD-L1) and programmed death protein 1 (PD-1) expression levels in many tumors and their correlation with prognosis have been actively studied. However, studies on PD-1 expression and its prognostic value in clear cell renal cell carcinoma (ccRCC) are limited and controversial. In this study, we describe the expression of PD-1 and its prognostic significance and association with clinical features in patients with ccRCC. *Materials and Methods:* We obtained clinicopathological data from 166 patients with ccRCC who were treated at Gyeongsang National University Hospital, Jinju, Korea between January 2000 and December 2009. Tissue microarray blocks were made using representative paraffin blocks of ccRCC specimens. Two pathologists analyzed PD-L1 and PD-1 expression in both tumor and inflammatory cells. *Results:* PD-1 expression in tumor-infiltrating inflammatory cells was significantly correlated with unfavorable disease-free survival (DFS) ($p < 0.001$) and disease-specific survival (DSS) ($p = 0.002$) in univariate analysis. A statistically significant correlation between PD-1 expression and unfavorable DFS ($p = 0.025$) was observed in multivariate analysis. *Conclusion:* PD-1 expression in tumor-infiltrating inflammatory cells serves as an independent prognostic factor for unfavorable DSS in patients with ccRCC.

Clear cell renal cell carcinoma (ccRCC) accounts for ~80% of all kidney cancers and is a major lethal genitourinary malignancy (1-3). The prognosis of ccRCC depends on clinicopathologic factors, such as stage, size, grade and necrosis (4). Some studies have been performed to identify gene signatures of ccRCC, molecular prognostication strategies and biomarkers (5-7).

Recently, programmed death ligand-1 (PD-L1) and programmed death protein (PD-1) have been under the spotlight. PD-L1 and PD-1 expression in many tumors and their correlation with prognosis have been largely studied (8-13). Recent studies have also demonstrated the expression of PD-L1 and its correlation with prognosis in ccRCC and have reported that elevated levels of PD-L1 expression have a negative prognostic role in ccRCC (14-17). However, studies regarding PD-1 expression and its prognostic role in ccRCC are scarce and controversial (16, 18, 19).

In this study, we describe the expression of PD-1 and its prognostic significance and association with clinical features in patients with ccRCC.

Materials and Methods

Patients and clinicopathological data. By reviewing electronic clinical charts, we obtained clinicopathologic data of ccRCC patients at Gyeongsang National University Hospital, Jinju, Korea between January 2000 and December 2009. One hundred sixty-six patients with ccRCC were enrolled in this study. The tumors were staged according to the guidelines of the eighth edition of the American Joint Committee on Cancer tumor-node-metastasis (TNM) staging system. Recurrence was diagnosed by surgical biopsy or radiologically. Disease-free survival (DFS) was defined as the period from the date of the surgery to the date of cancer recurrence. Disease-specific survival was defined as the period from the date of the surgery to the date of death due to ccRCC.

Two pathologists reviewed hematoxylin and eosin-stained glass slides. This study was approved by the Institutional Review Board of Gyeongsang National University Hospital with a waiver of informed consent (GNUH 2020-04-006).

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Key Words: Clear cell renal cell carcinoma, PD-1, immuno-histochemistry, survival analysis, prognosis.

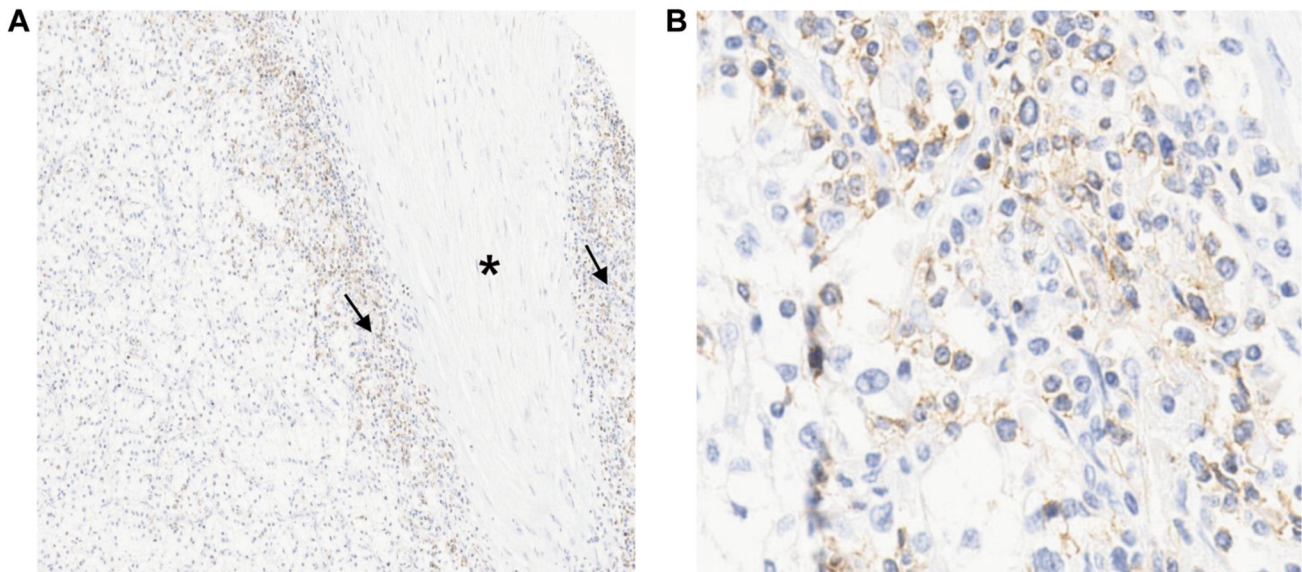


Figure 1. *PD-1* expression in ccRCC. (A) *PD-1* positive inflammatory cells are infiltrating the tumors (arrow) around the fibrotic band (star) (Original magnification ×100). (B) *PD-1* staining of inflammatory cells shows membranous pattern. (Original magnification ×400). *PD-1*: Programmed death protein 1; ccRCC: clear cell renal cell carcinoma.

Tissue microarray. Specimens were obtained surgically and fixed overnight in neutral buffered formalin (20%). The samples were embedded in paraffin blocks. Two cores of 2-mm representative tissue were collected from each paraffin block and transplanted onto new recipient tissue microarray (TMA) blocks. Two cores were collected from the center and periphery of the tumor.

Immunohistochemical analysis. Primary antibodies against PD-L1 (1:200, EIL3N, Cell Signaling Technology, Danvers, MA, USA) and PD-1 (1:100, ab52587, Abcam, Cambridge, UK) were used to evaluate protein expression. The immunohistochemical (IHC) method is described in our previous report (20). PD-L1 scoring (SP142) was based on the percentage of PD-L1-expressing tumor cells or inflammatory cells with any intensity (21). To increase the reproducibility, we evaluated the intensity of PD-1 expression of membranous staining on tumor cells and inflammatory cells simply as negative or positive for any percentage (Figure 1).

Statistical analysis. The correlation between PD-1 expression and clinicopathological parameters was analyzed by using chi-square tests. The prognostic significance of the clinicopathological data for DFS and DSS was evaluated by Cox proportional hazards regression analysis. Survival probability was also analyzed by the Kaplan-Meier method with the log-rank test for DFS and DSS. The results were considered statistically significant when the *p*-value was less than 0.05. SPSS version 21.0 (IBM Corp, Armonk, NY, USA) was used for the analysis.

Results

Clinicopathological data. The clinicopathological data of patients with ccRCC (*n*=166) are summarized in Table I. The mean age of the patients was 59 years (range=32-84 years).

There were 124 (74.7%), 12 (7.2%), 26 (15.7%), and 4 (2.4%) patients with T stage 1, 2, 3 and 4 disease, respectively. The Fuhrman nuclear grades were as follows: 30 (18.1%) were grade 1, 110 (66.3%) were grade 2, 20 (12.1%) were grade 3, and 6 (3.6%) were grade 4.

PD-L1 and PD-1 expression. Of the 332 cores, 58 cores had hemosiderin pigment or artificial brown pigment, and one core was not available due to loss of the specimen. In total, 59 cores were not informative. Ultimately, 273 available cores were used for the analysis.

Of the 273 cores evaluated, 123 showed PD-1 expression in inflammatory cells and no PD-1 expression in tumor cells. No cores showed PD-L1 expression in tumor and inflammatory cells. The positive control for PD-L1 showed high expression. PD-1-positive inflammatory cells showed a membranous staining pattern.

Relationship between PD-1 expression and clinicopathological data. The relationships between PD-1 expression and clinicopathological data are shown in Table II. PD-1 expression was significantly related to male sex (*p*=0.046) and T stage=1 (*p*<0.001) but not to age (*p*=0.597) or Fuhrman nuclear grade (*p*=0.086).

PD-1 expression and survival analysis. Univariate and multivariate analyses of survival according to PD-1 expression in 273 available cores are shown in Table III. In the univariate analysis, several variables were associated with poor DFS, including age ≥60 (*p*=0.001), Fuhrman grades 3

Table I. Clinicopathological information of 166 patients with clear cell renal cell carcinoma.

Variable	Value (%)
Age, median [range]	59 [32~84]
Gender (M/F)	120/46 (72.3/27.7)
Follow-up period, mean (year)	4.05
T stage	
1a	99 (59.6)
1b	25 (15.1)
2a	9 (5.4)
2b	3 (1.8)
3a	24 (14.5)
3b	2 (1.2)
4	4 (2.4)
Fuhrman nuclear grade	
1	30 (18.1)
2	110 (66.3)
3	20 (12.1)
4	6 (3.6)
Available cores*/total cores**	273/332 (82.5/100)
PD-1 expression in available cores	123 cores (44.9)
PD-L1 expression in available cores	0 cores (0%)
Total number of patients	166

*58 cores had hemosiderin pigment or artificial brown pigment, and one core was not available due to loss of specimen. In total, 59 cores were not informative; **two cores were obtained from paraffin specimens of each 166 patients with clear cell renal cell carcinoma. Two cores were collected from the central and peripheral areas of the tumor. A total of 273 available cores exhibited a full circle with a 2 mm diameter without specimen loss. PD-1: Programmed death protein 1; PD-L1: programmed cell death ligand-1.

and 4 ($p<0.001$), T stage ≥ 2 ($p<0.001$) and PD-1 expression ($p<0.001$). Poor DSS was also associated with age ≥ 60 ($p=0.021$), Fuhrman grades 3 and 4 ($p<0.001$), T stage ≥ 2 ($p<0.001$) and PD-1 expression ($p=0.002$). Moreover, multivariate analysis revealed that PD-1 expression was an independent factor for unfavorable DFS (hazard ratio=2.171; 95% confidence interval=1.103-4.275; $p=0.025$).

Discussion

Tumors are usually infiltrated by inflammatory cells. The composition of inflammatory cells and the number of each subtype differ between different tumors. These inflammatory cells interact with each other and affect the growth of tumors. In addition, these inflammatory cells can be related to a good prognosis or a poor prognosis in patients (22). Tumor microenvironments are mostly composed of tumor-infiltrating lymphocytes including CD4+ and CD8+ T cells (23). These T cells are regulated by positive and negative signals (24). Among inhibitory immune checkpoint proteins, PD-1 and PD-L1 have been actively studied, and their relationships to cancer, infectious disease, transplantation,

Table II. Correlations among clinicopathological factors and PD-1 expression.

Variables	273 available cores		<i>p</i> -Value
	PD-1 negative	PD-1 positive	
Age (years)			0.597
≤ 59	78	60	
≥ 60	72	63	
Gender			0.046
Male	98	94	
Female	52	29	
Fuhrman nuclear grade			0.086
1,2	129	96	
3,4	21	27	
T stage			<0.001
1	129	69	
≥ 2	21	54	

PD-1: Programmed death protein 1; PC: PD-1 positive cells. Bold values indicate statistical significance.

and autoimmunity have been demonstrated (25-27). For example, a strong correlation between PD-1 and PD-L1 expression and prognosis in cancer patients has been actively demonstrated (8, 11, 12, 15, 16, 25).

Additionally, there has been significant research on PD-1 and PD-L1 and ccRCC. Usually, PD-L1 is expressed in both immune cells and tumor cells, while PD-1 is mainly expressed in tumor infiltrating lymphocytes (28). Many previous reports have also shown that PD-L1 is an independent poor prognostic factor for ccRCC (16, 17, 19). However, previous studies on the correlation between PD-1 and ccRCC are limited and controversial. Stenzel *et al.* (19) reported that high PD-1 expression in inflammatory cells is significantly associated with favorable cancer-specific survival. In contrast, Ueda *et al.* (16) reported that PD-1 expression is significantly associated with adverse outcomes in metastatic ccRCC.

Our results showed that PD-1 expression was significantly correlated with male sex and higher T stage. PD-1 expression was related to unfavorable DFS and DSS in the univariate analysis of patients with ccRCC. Moreover, PD-1 expression was related to unfavorable DFS in multivariate analysis in patients with ccRCC. Based on our results, we hypothesize that PD-1 expression inhibits T cell-mediated antitumor responses in ccRCC. This finding suggests that PD-1 pathway blockade will enhance antitumor effects in ccRCC.

Our study has limitation. Tissue microarray is a valuable technique evaluating immunohistochemical markers in tumors and used as an alternative for whole tissue sections (29). However, the possibility of intratumoral heterogeneity in biomarker expression is concerned in small tissue microarray specimen (30). There have been a few studies on PD-L1

Table III. Cox proportional hazards analysis of PD-1 expression in 273 available cores.

Variables	Univariate analysis				Multivariate analysis			
	DFS		DSS		DFS		DSS	
	HR (95%CI)	p-Value	HR (95%CI)	p-Value	HR (95%CI)	p-Value	HR (95% I)	p-Value
Age (years) (≤ 59 vs. ≥ 60)	2.704 (1.479-4.945)	0.001	2.280 (1.133-4.588)	0.021	1.681 (0.905-3.123)	0.100	1.303 (0.641-2.653)	0.465
Gender (M vs. F)	0.475 (0.223-1.012)	0.054	1.018 (0.459-2.259)	0.965				
Fuhrman (1,2 vs. 3,4)	5.020 (2.866-8.794)	<0.001	6.058 (3.115-11.784)	<0.001	3.092 (1.684-5.679)	<0.001	4.058 (2.016-8.168)	<0.001
T stage (1 vs. ≥ 2)	17.578 (8.504-36.334)	<0.001	25.431 (8.956-72.219)	<0.001	11.855 (5.300-26.517)	<0.001	19.366 (6.426-58.368)	<0.001
PD-1 (- vs. +)	4.449 (2.319-8.535)	<0.001	3.271 (1.567-6.827)	0.002	2.171 (1.103-4.275)	0.025	0.972 (0.441-2.142)	0.944

PD-1: Programmed death protein 1; DFS: disease-free survival; DSS: disease-specific survival; HR: hazard ratio; CI: confidence interval. Bold values indicate statistical significance.

expression and heterogeneity. Intratumoral and intertumoral heterogeneity was present in more than half of the cases (31). However, to our knowledge, there have been no studies on the intratumoral and intertumoral heterogeneity of PD-1 expression in inflammatory cells. Therefore, further studies of PD-1 expression in ccRCC with whole tumor sections are needed.

Conclusion

This study showed that PD-1 expression in tumor infiltrating inflammatory cells serves as an independent factor for unfavorable DSS in patients with ccRCC.

Conflicts of Interest

The Authors declare that they have no competing interests in relation to this study.

Authors' Contributions

MHK and DHS conceived and designed this study. MHK, GHK, JHL, JSL, DCK, JWY, JMN, HJA and DHS collected samples, performed pathological diagnosis and analyzed the immunostained samples. MHK and DHS analyzed all the data. MHK and DHS wrote the first draft of the manuscript. GHK, JHL, JSL, DCK, JWY, JMN, HJA and DHS critically reviewed and corrected the manuscript. All Authors reviewed and approved the final version of the manuscript.

References

- 1 Siegel RL, Miller KD and Jemal A: Cancer statistics, 2019. *CA Cancer J Clin* 69(1): 7-34, 2019. PMID: 30620402. DOI: 10.3322/caac.21551
- 2 Escudier B, Porta C, Schmidinger M, Rioux-Leclercq N, Bex A, Khoo V, Grünwald V, Gillessen S, Horwich A and ESMO Guidelines Committee: Renal cell carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 30(5): 706-720, 2019. PMID: 30788497. DOI: 10.1093/annonc/mdz056
- 3 Patard JJ, Leray E, Rioux-Leclercq N, Cindolo L, Ficarra V, Zisman A, De La Taille A, Tostain J, Artibani W, Abbou CC, Lobel B, Guillé F, Chopin DK, Mulders PF, Wood CG, Swanson DA, Figlin RA, Belldegrun AS and Pantuck AJ: Prognostic value of histologic subtypes in renal cell carcinoma: a multicenter experience. *J Clin Oncol* 23(12): 2763-2771, 2005. PMID: 15837991. DOI: 10.1200/JCO.2005.07.055
- 4 Leibovich BC, Blute ML, Cheville JC, Lohse CM, Frank I, Kwon ED, Weaver AL, Parker AS and Zincke H: Prediction of progression after radical nephrectomy for patients with clear cell renal cell carcinoma: A stratification tool for prospective clinical trials. *Cancer* 97(7): 1663-1671, 2003. PMID: 12655523. DOI: 10.1002/cncr.11234
- 5 Brooks SA, Brannon AR, Parker JS, Fisher JC, Sen O, Kattan MW, Hakimi AA, Hsieh JJ, Choueiri TK, Tamboli P, Maranchie JK, Hinds P, Miller CR, Nielsen ME and Rathmell WK: ClearCode34: A prognostic risk predictor for localized clear cell renal cell carcinoma. *Eur Urol* 66(1): 77-84, 2014. PMID: 24613583. DOI: 10.1016/j.eururo.2014.02.035
- 6 Rini B, Goddard A, Knezevic D, Maddala T, Zhou M, Aydin H, Campbell S, Elson P, Koscielny S, Lopatin M, Svedman C, Martini JF, Williams JA, Verkarre V, Radulescu C, Neuzillet Y, Hemmerlé I, Timsit MO, Tsiatis AC, Bonham M, Lebre T, Mejean A and Escudier B: A 16-gene assay to predict recurrence after surgery in localised renal cell carcinoma: Development and validation studies. *Lancet Oncol* 16(6): 676-685, 2015. PMID: 25979595. DOI: 10.1016/S1470-2045(15)70167-1
- 7 Rasti A, Mehrzama M, Madjd Z, Abolhasani M, Saeednejad Zanjani L and Asgari M: Co-expression of cancer stem cell markers OCT4 and NANOG predicts poor prognosis in renal

- cell carcinomas. *Sci Rep* 8(1): 11739, 2018. PMID: 30082842. DOI: 10.1038/s41598-018-30168-4
- 8 Tamura T, Ohira M, Tanaka H, Muguruma K, Toyokawa T, Kubo N, Sakurai K, Amano R, Kimura K, Shibutani M, Maeda K and Hirakawa K: Programmed death-1 ligand-1 (pdl1) expression is associated with the prognosis of patients with stage II/III gastric cancer. *Anticancer Res* 35(10): 5369-5376, 2015. PMID: 26408698.
 - 9 Kerr KM, Tsao MS, Nicholson AG, Yatabe Y, Wistuba II, Hirsch FR and IASLC Pathology Committee: Programmed death-ligand 1 immunohistochemistry in lung cancer: In what state is this art? *J Thorac Oncol* 10(7): 985-989, 2015. PMID: 26134220. DOI: 10.1097/JTO.0000000000000526
 - 10 Kim SH, Go SI, Song DH, Park SW, Kim HR, Jang I, Kim JD, Lee JS and Lee GW: Prognostic impact of CD8 and programmed death-ligand 1 expression in patients with resectable non-small cell lung cancer. *Br J Cancer* 120(5): 547-554, 2019. PMID: 30745585. DOI: 10.1038/s41416-019-0398-5
 - 11 Ni X, Sun X, Wang D, Chen Y, Zhang Y, Li W, Wang L and Suo J: The clinicopathological and prognostic value of programmed death-ligand 1 in colorectal cancer: A meta-analysis. *Clin Transl Oncol* 21(5): 674-686, 2019. PMID: 30392153. DOI: 10.1007/s12094-018-1970-9
 - 12 Planes-Laine G, Rochigneux P, Bertucci F, Chrétien AS, Viens P, Sabatier R and Gonçalves A: PD-1/PD-L1 targeting in breast cancer: The first clinical evidences are emerging, a literature review. *Cancers (Basel)* 11(7): 1033, 2019. PMID: 31336685. DOI: 10.3390/cancers11071033
 - 13 Zhao JJ, Zhou ZQ, Wang P, Chen CL, Liu Y, Pan QZ, Zhu Q, Tang Y, Weng DS and Xia JC: Orchestration of immune checkpoints in tumor immune contexture and their prognostic significance in esophageal squamous cell carcinoma. *Cancer Manag Res* 10: 6457-6468, 2018. PMID: 30568505. DOI: 10.2147/CMAR.S181949
 - 14 Iacovelli R, Nolè F, Verri E, Renne G, Paglino C, Santoni M, Cossu Rocca M, Giglione P, Aurilio G, Cullurà D, Cascinu S and Porta C: Prognostic role of PD-L1 expression in renal cell carcinoma. A systematic review and meta-analysis. *Target Oncol* 11(2): 143-148, 2016. PMID: 26429561. DOI: 10.1007/s11523-015-0392-7
 - 15 Xu F, Xu L, Wang Q, An G, Feng G and Liu F: Clinicopathological and prognostic value of programmed death ligand-1 (PD-L1) in renal cell carcinoma: A meta-analysis. *Int J Clin Exp Med* 8(9): 14595-14603, 2015. PMID: 26628942.
 - 16 Ueda K, Suekane S, Kurose H, Chikui K, Nakiri M, Nishihara K, Matsuo M, Kawahara A, Yano H and Igawa T: Prognostic value of PD-1 and PD-L1 expression in patients with metastatic clear cell renal cell carcinoma. *Urol Oncol* 36(11): 499.e9-499.e16, 2018. PMID: 30131293. DOI: 10.1016/j.urolonc.2018.07.003
 - 17 Leite KR, Reis ST, Junior JP, Zerati M, Gomes Dde O, Camara-Lopes LH and Srougi M: PD-L1 expression in renal cell carcinoma clear cell type is related to unfavorable prognosis. *Diagn Pathol* 10: 189, 2015. PMID: 26470780. DOI: 10.1186/s13000-015-0414-x
 - 18 Erlmeier F, Weichert W, Schrader AJ, Autenrieth M, Hartmann A, Steffens S and Ivanyi P: Prognostic impact of PD-1 and its ligands in renal cell carcinoma. *Med Oncol* 34(6): 99, 2017. PMID: 28432616. DOI: 10.1007/s12032-017-0961-y
 - 19 Stenzel PJ, Schindeldecker M, Tagscherer KE, Foersch S, Herpel E, Hohenfellner M, Hatiboglu G, Alt J, Thomas C, Haferkamp A, Roth W and Macher-Goeppinger S: Prognostic and predictive value of tumor-infiltrating leukocytes and of immune checkpoint molecules PD1 and PDL1 in clear cell renal cell carcinoma. *Transl Oncol* 13(2): 336-345, 2020. PMID: 31881506. DOI: 10.1016/j.tranon.2019.11.002
 - 20 Song DH, Ko GH, Lee JH, Lee JS, Kim HC, Yang JW, Heo RW, Roh GS, Han SY and Kim DC: Myoferlin expression in non-small cell lung cancer: Prognostic role and correlation with VEGFR-2 expression. *Oncol Lett* 11(2): 998-1006, 2016. PMID: 26893682. DOI: 10.3892/ol.2015.3988
 - 21 Teixidó C, Vilarinho N, Reyes R and Reguart N: PD-L1 expression testing in non-small cell lung cancer. *Ther Adv Med Oncol* 10: 1758835918763493, 2018. PMID: 29662547. DOI: 10.1177/1758835918763493
 - 22 Whiteside TL: The tumor microenvironment and its role in promoting tumor growth. *Oncogene* 27(45): 5904-5912, 2008. PMID: 18836471. DOI: 10.1038/onc.2008.271
 - 23 Whiteside TL: The local tumor microenvironment. In: *General Principles of Tumor Immunotherapy*. HL Kaufman, JD Wolchok (eds.), Springer, pp. 145-167, 2007.
 - 24 Chen L: Co-inhibitory molecules of the B7-CD28 family in the control of T-cell immunity. *Nat Rev Immunol* 4(5): 336-347, 2004. PMID: 15122199. DOI: 10.1038/nri1349
 - 25 Okazaki T and Honjo T: PD-1 and PD-1 ligands: From discovery to clinical application. *Int Immunol* 19(7): 813-824, 2007. PMID: 17606980. DOI: 10.1093/intimm/dxm057
 - 26 Pardoll DM: The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* 12(4): 252-264, 2012. PMID: 22437870. DOI: 10.1038/nrc3239
 - 27 An HJ, Ko GH, Lee JH, Lee JS, Kim DC, Yang JW, Kim MH, Kim JP, Jung EJ and Song DH: Programmed death-ligand 1 expression and its correlation with lymph node metastasis in papillary thyroid carcinoma. *J Pathol Transl Med* 52(1): 9-13, 2018. PMID: 28994272. DOI: 10.4132/jptm.2017.07.26
 - 28 McDermott DF and Atkins MB: Immune therapy for kidney cancer: A second dawn? *Semin Oncol* 40(4): 492-498, 2013. PMID: 23972713. DOI: 10.1053/j.seminoncol.2013.05.008
 - 29 Khouja MH, Baekelandt M, Sarab A, Nesland JM and Holm R: Limitations of tissue microarrays compared with whole tissue sections in survival analysis. *Oncol Lett* 1(5): 827-831, 2010. PMID: 22966388. DOI: 10.3892/ol_00000145
 - 30 Nassar A, Radhakrishnan A, Cabrero IA, Cotsonis GA and Cohen C: Intratumoral heterogeneity of immunohistochemical marker expression in breast carcinoma: A tissue microarray-based study. *Appl Immunohistochem Mol Morphol* 18(5): 433-441, 2010. PMID: 20485156. DOI: 10.1097/PAI.0b013e3181dddb20
 - 31 Haragan A, Field JK, Davies MPA, Escriu C, Gruver A and Gosney JR: Heterogeneity of PD-L1 expression in non-small cell lung cancer: Implications for specimen sampling in predicting treatment response. *Lung Cancer* 134: 79-84, 2019. PMID: 31320000. DOI: 10.1016/j.lungcan.2019.06.005

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