Mid-trimester Amniotic Fluid Interleukins (IL-1β, IL-10 and IL-18) as Possible Predictors of Preterm Delivery

K. PUCHNER¹, C. IAVAZZO¹, D. GOURGIOTIS², M. BOUTSIKOU¹, S. BAKA¹, D. HASSIAKOS^{1,3}, E. KOUSKOUNI¹, E. ECONOMOU¹, A. MALAMITSI-PUCHNER¹ and G. CREATSAS¹

¹Second Department of Obstetrics and Gynecology, University of Athens, Aretaieion Hospital, Athens, Greece;

²Research Laboratories, Second Department of Pediatrics, University of Athens,

A. & P. Kyriakou Hospital, Athens, Greece;

³Department of Fetal Medicine 'LITO' Maternity Hospital Athens, Greece

Abstract. Aim: Strong evidence implicates chronic intraamniotic inflammation in the etiology of preterm delivery. The purpose of this study was to determine whether amniotic fluid IL-1\beta, IL-10 and IL-18 concentrations in women undergoing mid-trimester amniocentesis can identify those at risk for preterm labor or preterm rupture of membranes. Patients and Methods: A case-control study was conducted to compare mid-trimester concentrations of amniotic fluid IL-1 β , IL-10 and IL-18 in women delivering at term or preterm. Out of 362 women included in the study, 38 presented with preterm labor. Thirty-eight women with term delivery, matched for chronological and gestational age served as controls. Women with abnormal fetal karyotypes or major anomalies were excluded. IL-1\beta, IL-10 and IL-18 concentrations were determined by ELISA. Conditional logistic regression was applied in the statistical analysis. Results: IL-1 β was found to be positively and significantly associated with preterm delivery. Specifically, for every unit increase in IL-1 β , women were on average 7.2 (OR: 7.2, CI: 1.94-26.77, p=0.003) times more likely to deliver preterm. IL-18 levels as well as gender were significantly associated with preterm delivery. Specifically, for every unit increase in IL-18, women were on average 1% less likely to have a preterm delivery (OR: 0.99, CI: 0.98-0.99, p=0.04). On the other hand, IL-10 was not significantly associated with preterm delivery. Conclusion: Mid-trimester IL-1 β concentrations are positively associated with preterm delivery. Therefore, IL- 1β , determined on the occasion of mid-trimester amniocentesis could possibly

amniotic fluid concentrations of various cytokines (including IL-6, TNF α , ITAC, ADAM-8, beta-defensins) (7-11). Interleukin 1- β (IL-1 β) is a 17 kDa cytokine (12), produced by macrophages, monocytes and dendritic cells as a rapid response to bacterial antigen stimuli, and is a characteristic mediator of inflammation (13). IL-10 is an 18-kDa cytokine

mediator of inflammation (13). IL-10 is an 18-kDa cytokine (14), capable of inhibiting the production of cytokines by activated Th2 cells (15). IL-18 shows structural homology with the IL-1 family of cytokines (16) and is synthesized by macrophages, monocytes, keratinocytes, and epithelial cells (17).

In this study, it was hypothesized that IL-1 β , IL-10 and IL-18 might be implicated in the preterm delivery pathway and therefore, they could serve as possible predictors of the former. In this respect, their amniotic fluid concentrations were determined on the occasion of genetic amniocentesis and correlated them with preterm labor and/or premature rupture of membranes.

Correspondence to: C. Iavazzo, 38, Seizani Str., Nea Ionia, Athens, Greece 14231. Tel: +30 6948054119, e-mail: christosiavazzo@hotmail.com

Key Words: Preterm labor, premature rupture of membranes, cytokines, prediction, interleukins, amniotic fluid.

Patients and Methods

This was a prospective matched case-control study, performed in collaboration of the Second Department of Obstetrics and Gynecology, University of Athens and the Department of Fetal Medicine of 'LITO' Maternity Hospital in Athens, during the period September 2005 - December 2006. The study population consisted of Greek women with singleton pregnancies who

IL-18 concentrations are not elevated in mid-trimester amniotic fluid and probably cannot serve this purpose.

The incidence of preterm labor varies from 7-12.8% of

deliveries (1, 2) and accounts for 75% of perinatal mortality

and morbidity (e.g. blindness, deafness, developmental delay,

cerebral palsy and chronic lung disease (3, 4). Although the etiology of preterm birth is multifactorial (5), the impact of

intrauterine inflammation/infection is pronounced, being

implicated in nearly 30% of cases (6). Inflammation/infection-

associated preterm delivery is characterized by increased

serve as a marker of preterm delivery. In contrast, IL-10 and

0258-851X/2011 \$2.00+.40

presented for genetic amniocentesis. Women with twin pregnancies and women with known history of uterine abnormalities, cone biopsy, significant vaginal bleeding and fetal malformations were excluded from the study.

Preterm labor was defined as labor before 37 weeks of gestation with regular uterine contractions (at least two uterine contractions at a 10-minute interval during a 30-minute period) in combination with characteristic cervical changes (18). Preterm premature rupture of the fetal membranes was defined as the rupture of the amniotic membranes with amniotic fluid release more than one hour before the onset of preterm labor (19).

Gestational age was calculated from the date of last menstruation and was confirmed by routine ultrasound in the second trimester (16-19 weeks of gestation). Microbial invasion was defined as growth of any bacteria (aerobic or anaerobic) in the amniotic fluid cultures except for coagulase-negative Staphylococcus, (which was considered skin contamination) and/or positive PCR for Mycoplasma hominis and Chlamydia trachomatis. All patients were followed until delivery for the occurrence of pregnancy complications by their doctors. An independent investigator kept the medical records and entered maternal and perinatal data into a database. The Ethics Committee of the host teaching hospital approved the study. Each woman gave informed consent before enrolment in the study and completed a questionnaire regarding personal and family data.

Ultrasound-guided transabdominal amniocentesis with a 21gauge needle was performed under aseptic conditions in 362 women during genetic amniocentesis. Amniocentesis was performed for advanced maternal age, and/or increased risk for aneuploidy during nuchal translucency ultrasound. The first 0.5 ml of collected amniotic fluid was discarded to avoid maternal contamination. A volume of 20 ml of amniotic fluid was aspirated from each woman (15 ml was used for genetic diagnosis). One ml of the uncentrifuged amniotic fluid was then transported immediately to the laboratory and cultured for aerobic and anaerobic bacteria. Another 1 ml of the uncentrifuged amniotic fluid was also tested by polymerase chain reaction (PCR) for Mycoplasma hominis and/or Chlamydia trachomatis detection. The remaining 3 ml of amniotic fluid were placed immediately in a refrigerator (+4°C) and were centrifuged within the next six hours at 3000 g and +4°C for 10 min. The supernatant was stored in polypropylene tubes at -80°C until analysis. Amniotic fluid IL-1β and IL-10 concentrations were determined by an enzyme-linked immunosorbent assay (ELISA) (IL-1\beta, IL-10 and IL-18 by Bender MedSystems GmBH, A-1030Vienna, Austria) while IL-18 concentrations were determined by an enzyme-linked immunosorbent assay (ELISA) (MBL, Naka-ku Nagoya, Japan) (sandwich ELISA technique). The amniotic fluid samples ran in duplicates. For IL-B, the intra-assay coefficient of variation (CV) was 6.7%, the inter-assay CV 8.1% and the detection limit 0.05 pg/ml. For IL-1β, the intra-assay CV was 5.61%, the inter-assay CV 6.25% and the detection limit 125 pg/ml. For IL-10, the intraassay CV was 3.2%, the inter-assay CV 5.6% and the detection limit 1 pg/ml. (Intra- and inter-assay coefficients of CV refer to the laboratory characteristics of each kit used). Laboratory personnel were blinded to the clinical history of the women involved.

Statistical analysis. All data, except for age, gestational age at delivery and gestational age at amniocentesis, followed a normal distribution (Kolmogorov-Smirnov test). Independent samples *t*-test

was applied to detect differences between groups where continuous variables were normally distributed. Otherwise, the Mann-Whitney U-test was applied. Pearson's χ^2 test was used to detect differences between categorical variables.

Conditional logistic regression analysis was used to examine the possible associations of human IL-1 β , IL-10 and IL-18 with preterm labor. Women with a preterm labor were defined as cases (N=38), while for each case a woman matched for age who delivered at term served as control (N=38).

Furthermore, subgroup analysis was conducted in order to examine any possible association of human IL-1 β , IL-10, and IL-18 in women with preterm labor who delivered before 32 weeks of gestation (N=8 in each group). Finally, a subgroup of women who had preterm labor and premature rupture of membranes were evaluated (n=18 in each group). Results are presented as odds ratios (OR) and 95% confidence intervals (CI). Statistical analysis was performed by using SPSS 11.5 edition. A p-value of <0.05 was considered to be statistically significant.

Results

Out of the 360 women included in the study, 38 presented with spontaneous preterm delivery (incidence:10.56%), and out of that 38, 18 delivered after premature rupture of membranes (incidence: 5%). Ten women were excluded after amniocentesis because of fetal chromosomal abnormalities (two with trisomy 18, two with trisomy 21, one with Turner syndrome, one with Klinefelter syndrome and four with less common pathological karyotypes). Ten women were lost to follow-up. Four infants were delivered by caesarean section before the onset of labor for maternal (severe preeclampsia) or fetal reasons (compromised fetal growth or umbilical Doppler flow abnormalities). Two women who delivered within 30 days following amniocentesis were excluded from the study, as their delivery was considered related to the procedure of amniocentesis (20, 21).

The demographic data of the study population are presented in Tables I and II. No statistical significant differences were found in mean maternal age, gestational age at amniotic fluid sampling and parity between the groups of women delivering preterm and at term. Amniotic fluid cultures for common bacteria in the participating mothers were negative, while *Mycoplasma hominis* and *Chlamydia trachomatis* were identified in 2/338 and 2/338, respectively. However, one of the two women with *Mycoplasma hominis* and one of the two women with *Chlamydia trachomatis* delivered preterm. No correlation was found between PCR detection of *Mycoplasma hominis* and/or *Chlamydia trachomatis* with preterm labor prediction.

IL-1 β , IL-10, IL-18 concentrations in cases of preterm delivery (n=38) and controls (n=38), as well as in cases of premature rupture of membranes (n=18) and controls (n=18) are presented in Tables III and IV, respectively. IL-1 β was found to be positively associated with preterm delivery and this association was statistically significant. Specifically, for

Table I. Demographic data for women with preterm labor who delivered before 37 weeks of gestation (cases, n=38) and women delivering at term (controls, n=38).

Group	Preterm labor (n=38) mean±SD/median (range)	Controls (n=38) mean±SD/median (range)	<i>P</i> -value
Gestational age at delivery (weeks)	35.43 (22-37.0)	38.28 (37-40)	< 0.001
Gestational age at amniocentesis (weeks)	17.86 (15.14-25.43)	17.43 (15.86-19.71)	0.166
Birth weight (g)	2447.2±590.51	3232.78±308.75	< 0.001
Age (years)	37 (27-43)	37 (27-44)	0.883
Parity, n (%)			0.554
0	27 (71.0)	25 (65.8)	
≥1	11 (29.0)	13 (34.2)	
Smoking before pregnancy, n (%)	14 (36.8)	12 (31.6)	0.569
Smoking during pregnancy, n (%)	6 (15.8)	9 (23.7)	0.419
Alcohol consumption during pregnancy, n (%)	1 (2.6)	4 (10.5)	0.174
Use of drugs, n (%)	17(44.7)	21 (55.3)	0.280
Mode of delivery, n (%)			0.063
Vaginal	6 (15.8)	15 (39.5)	
Ceasarean	32 (84.2)	23 (60.5)	
Neonate gender, n (%)			
Male	26 (68.4)	14 (36.8)	0.011
Female	12 (31.6)	24 (63.2)	

Table II. Demographic data for women with preterm labor and premature rupture of membranes (cases, n=18) and women delivering at term (controls, n=18).

Groups	Preterm labor and premature rupture of membranes (n=18) Mean±SD	Controls (n=18) Mean±SD	P-value
Gestational age at delivery (weeks)	33.58±4.23	38.59±0.86	< 0.001
Gestational age at amniocentesis (weeks)	19.0±2.64	17.85±1.0	0.096
Birth weight (g)	2483.53±669.22	3252.35±353.53	< 0.001
Age (years)	34.44±9.4	36.67±3.94	0.362
Parity, n (%)			0.272
0	11 (61.1)	8 (44.4)	
≥1	7 (38.9)	10 (55.6)	
Smoking before pregnancy, n (%)	6 (33.3)	6 (33.3)	0.903
Smoking during pregnancy, n (%)	3 (16.7)	4 (22.2)	0.735
Alcohol consumption during pregnancy, n (%)	1 (5.6)	2 (11.1)	0.581
Use of drugs, n (%)	6 (33.3)	9 (50)	0.288
Mode of delivery, n (%)			0.227
Vaginal	4 (22.2)	8 (44.4)	
Ceasarean	14 (77.8)	10 (55.6)	
Neonate gender, n (%)			0.038
Male	15 (83.3)	9 (50)	
Female	3 (16.7)	9 (50)	

every unit increase in IL-1 β women were on average 7.2 times more likely to have a preterm delivery (OR: 7.2, CI: 1.94-26.77, p=0.003) (Figure 1). IL-18 levels, as well as gender, were significantly associated with preterm delivery. Specifically, for every unit increase in IL-18 women were on average 1% less likely to have a preterm delivery (OR:

0.99, CI: 0.98-0.99, p=0.04) (Figure 2). Furthermore, women who delivered female neonates were 84% less likely to have a preterm delivery as compared to women who delivered males (OR: 0.16, CI: 0.40-0.71, p=0.016). IL-10 was not found to be significantly associated with preterm delivery (Figure 3).

Table III. IL-1 β , IL-10, IL-18 concentrations in cases of preterm delivery (n=38) and controls (n=38).

Group	Preterm labor (n=38) Mean±SD/median (range)	Control (n=38) Mean±SD/median (range)
IL-1β (pg/ml) IL-10 (pg/ml) IL-18 (pg/ml)	1.50 (0.48-8.93) 3.07 (1.59-8.34) 90.69±69.45	0.72 (0.46-2.01) 3.26 (1.59-21.94) 127.56±68.52

Table IV. IL-1 β , IL-10, IL-18 concentrations in cases with premature rupture membranes (n=18) and controls (n=18).

Group	Preterm labor and premature rupture of membranes (n=18) Mean±SD/median (range)	Controls (n=18) Mean±SD/median (range)
IL-1β (pg/ml) IL-10 (pg/ml) IL-18 (pg/ml)	1.53 (0.64-5.73) 2.73 (0.98-8.05) 86.83±74.54	0.72 (0.51-1.84) 3.33 (1.71-15.52) 139.85±87.06

In the subgroup of women with preterm labor who delivered before 32 weeks of gestation, IL-1 β , IL-10 and IL-18 were not found to be significantly associated with preterm delivery. IL-10 and IL-18 were not found to be significantly associated with preterm delivery in this sub-analysis.

Discussion

The results of this study indicate that amniotic fluid IL-1 β concentrations are positively associated with preterm delivery, as well as preterm premature rupture of membranes. It has been previously documented that IL-1 β is produced by epithelial cells of the amnion, chorion, syncytiotrophoblast and decidua (22) as a procytokine molecule, cleaved by caspase-1 into its active form (23). Moreover, it has been shown that high levels of IL-1 β are present in amniotic fluid samples with detectable bacteria (24). For the participating mothers in the current study, amniotic fluid cultures for common bacteria were negative, while *Mycoplasma hominis* and *Chlamydia hominis* were only identified in 2/338 and 2/338, respectively.

IL-1 β is an important regulator of myometrial CRH-receptor-1 gene expression, a fact that might play a role in the uterine change from quiescence to contractility and labor (25, 26). It also indirectly increases oxytocin secretion in primary cultures of human deciduas through the cyclooxygenase-2 pathway (27) and modulates calcium mobilization from intracellular stores preparing the pregnant uterus for labor (28). IL-1 β also enhances MMP-1 and

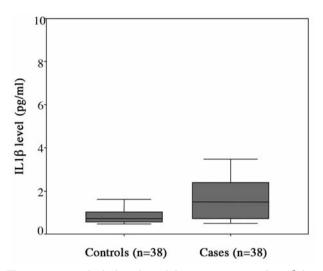


Figure 1. Box and whisker plots of the concentrations of IL-1 β from women with spontaneous preterm delivery who delivered before 37 weeks of gestation (cases) and women with full-term deliveries. Each box represents the median concentration with the interquartile range (25th and 75th percentiles).

MMP-3 expression in decidual cells, possibly leading to premature membrane rupture (29).

It has been shown that the major source of IL-10 is activated Th2 cells (30). However, other cells have also been implicated in IL-10 production, including Th1 (31), stimulated B-cells (32), LPS-activated monocytes (33), macrophages (34), dendritic cells (34, 35) and regulatory Tcells (36). Gestational tissues such as cytotrophoblast (37), syncytiotrophoblast (37), chorion (38) and decidua (39, 40), express IL-10 mRNA (41, 42) and produce IL-10. IL-10 is a pleiotropic anti-inflammatory cytokine (30, 43) acting in the down-regulation of the immune response (23) by inhibiting the IL-1β, IL-6 and PGE₂ production by choriodecidua (44). Nevertheless, IL-10 enhances production of cytokines and matrix metalloproteinases within the amnion (45). IL-10 also acts as an anti-inflammatory cytokine in the placenta (42, 46) by inhibiting the production of inflammatory cytokines such as IFN-γ, IL-1α, IL-1b, GM-CSF, G-CSF, TNFα, IL-6, IL-8 and IL-12 β (34). More specifically, the effects of IL-10 in fetal membranes depend on the precise tissue site. It inhibits cytokine production and matrix metalloproteinase expression in the choriodecidua (47). Furthermore, in rats, exogenously administered IL-10 attenuates fetal loss and growth restriction induced by LPS (48), whereas exogenous IL-10 has been shown to block intrauterine contractions in monkeys (49). Moreover, IL-10 has been shown to prolong gestation and to diminish fetal demise in a rat model when administered alone or in combination with antibiotics (50). For this reason IL-10 administration has been proposed as an anti-inflammatory agent in the treatment of preterm labor (49, 51).

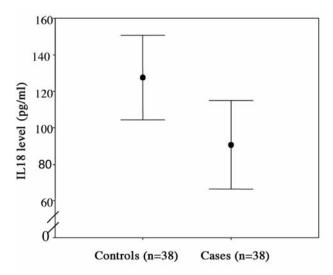


Figure 2. Mean concentrations of IL-18 from women with spontaneous preterm delivery who delivered before 37 weeks of gestation (cases) and women with full-term deliveries. Error bars represent the 95% CI.

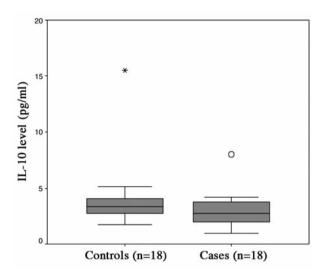


Figure 3. Box and whisker plots of the concentrations of IL-10 from women with spontaneous preterm delivery who delivered before 37 weeks of gestation (cases) and had premature rupture of membranes and women with full-term deliveries. Each box represents the median concentration with the interquartile range (25th and 75th percentiles).

IL-10 has been detected in the amniotic fluid during the second trimester of pregnancy (23), however, controversial findings relating to its concentrations throughout pregnancy have been reported (52). Gotsch et al. (23) showed that IL-10 is detectable in amniotic fluid and that its median levels do not change throughout pregnancy from mid-trimester to term. In contrast, Grieg et al. showed that amniotic fluid IL-10 concentrations increase throughout pregnancy and are elevated in patients with preterm labor associated with intrauterine infection (53). However, no differences in IL-10 concentrations have been documented in cases with preterm or term labor, or in cases with chorioamnionitis (54). Jones et al., using monolayer decidual cultures, reported that basal decidual IL-10 production rates did not change significantly with the onset of labor (55). In contrast, Simpson et al. (56) showed that the basal production of IL-10 by choriodecidual explants decreases after labor onset. This finding might be associated with the increase in the production of IL-6 and PGE₂ (56). To the Authors' knowledge the current study is the third investigating the role of mid-trimester amniotic fluid IL-10 concentrations as a possible predictor of preterm labor. No statistically significant association with preterm delivery was found in this study.

IL-18 is a physiological constituent of amniotic fluid, maternal plasma and cord blood and its levels increase with advancing gestational age (57). Sources of IL-18 during pregnancy include the chorion and the decidua, as IL-18 mRNA is expressed in the choriodecidual interface, but not the amnion (58). IL-18 has a critical role in host defense by

providing a cervical/decidual barrier against microbial invasion of the amniotic fluid (59). Its proinflammatory properties are based on the promotion of nuclear translocation of NF κ B (58), the production of TNF- α , IFN- γ and IL-1 β (17, 60) and the induction of both Th1 and Th2 response (61). IL-18 acts on macrophages and induces a response in order to enhance the immune response of the chorioamniotic membranes (62). More specifically, IL-18 activates the Th1 response by inducing IFN- γ production in T-cells, as well as NK cells (63). Moreover, IL-18 is a pro-apoptotic cytokine. IL-18 can increase cell membrane associated Fas expression and induce FasL, which will bind to Fas. This leads to intracellular signaling which facilitates procaspase-8 to its active form. This active caspase-8 is an effector for other caspases and leads to proteolysis and programmed cell death or apoptosis (58).

Amniotic fluid IL-18 was found to be significantly associated with histological chorioamnionitis in patients with preterm labor, whereas in premature rupture of membranes the results are controversial (58, 64). In animal models, the enhanced preterm labor rate was associated with decreased IL-18 (16). Moreover, it has been shown that there is a correlation between low levels of IL-18 in maternal serum and preterm labor. It has also been shown that cervical and amniotic fluid IL-18 levels are significantly higher in women in preterm labor when compared with women at term who are not in labor (59). The levels of IL-18 were higher in cases with microbial invasion (59). In the current study, no statistically significant association of IL-18 with preterm delivery was identified.

In conclusion, elevated amniotic fluid IL-1 β levels at the time of mid-trimester genetic amniocentesis might possibly serve as a predictive factor for preterm labor, with or without premature rupture of membranes. Future studies should further elucidate the possible roles of IL-1 β , IL-10, IL-18 in the preterm labor pathway.

References

- 1 Berkowitz GS and Papiernik E: Epidemiology of preterm birth. Epidemiol Rev *15*(2): 414-443, 1993.
- 2 Hamilton BE, Martin JA and Ventura SJ: Births: preliminary data for 2005. Natl Vital Stat Rep 55(11): 1-18, 2006.
- 3 Creasy RK: Preterm birth prevention: Where are we? Am J Obstet Gynecol 168(4): 1223-1230, 1993.
- 4 Speer CP: New insights into the pathogenesis of pulmonary inflammation in preterm infants. Biol Neonate 79: 205-209, 2001.
- 5 Ananth CV, Misra DP, Demissie K and Smulian JC: Rates of preterm delivery among Black women and White women in the United States over two decades: an age-period-cohort analysis. Am J Epidemiol 154(7): 657-665, 2001.
- 6 Ananth CV and Vintzileos AM: Epidemiology of preterm birth and its clinical subtypes. J Matern Fetal Neonatal Med 19(12): 773-782, 2006.
- 7 Romero R, Yoon BH, Mazor M, Gomez R, Diamond M, Kenney J, Ramirez M, Fidel P, Sorokin Y, Cotton D and Sehgal P: The diagnostic and prognostic value of amniotic fluid white blood cell count, glucose, interleukin-6 and Gram stain in patients with preterm labor and intact membranes. Am J Obstet Gynecol 169(4): 805-816, 1993.
- 8 Maymon E, Ghezzi F, Edwin S, Mazor M, Yoon BH, Gomez R and Romero R: The tumor necrosis factor [alpha] and its soluble receptor profile in term and preterm parturition. Am J Obstet Gynecol 181(5): 1142-1148, 1999.
- 9 Iavazzo C, Tassis K, Gourgiotis D, Boutsikou M, Baka S, Hassiakos D, Hadjithomas A, Botsis D and Malamitsi-Puchner A: The role of human beta defensins 2 and 3 in the second trimester amniotic fluid in predicting preterm labor and premature rupture of membranes. Arch Gynecol Obstet 281(5): 793-799, 2010.
- 10 Malamitsi-Puchner A, Vrachnis N, Samoli E, Baka S, Iliodromiti Z, Puchner KP, Malligianis P and Hassiakos D: Possible early prediction of preterm birth by determination of novel proinflammatory factors in midtrimester amniotic fluid. Ann NY Acad Sci 1092: 440-449, 2006.
- 11 Vrachnis N, Malamitsi-Puchner A, Samoli E, Botsis D, Iliodromiti Z, Baka S, Hassiakos D and Creatsas G: Elevated mid-trimester amniotic fluid ADAM-8 concentrations as a potential risk factor for preterm delivery. J Soc Gynecol Investig 13(3): 186-190, 2006.
- 12 Thornberry NA, Bull HG, Calaycay JR, Chapman KT, Howard AD, Kostura MJ, Miller DK, Molineaux SM, Weidner JR and Aunins J: A novel heterodimeric cysteine protease is required for interleukin-1 beta processing in monocytes. Nature 356(6372): 768-774, 1992.
- 13 Li YH, Brauner A, Jonsson B, van der Ploeg I, Soder O and Holst M: Ureaplasma urealyticum-induced production of proinflammatory cytokines by macrophages. Pediatr Res 48(1): 114-119, 2000.

- 14 Mosmann TR: Interleukin-10. In: The Cytokine Handbook. Thomson A (ed.). San Diego, CA: Academy Press Inc. pp. 223-237, 1994
- 15 Fiorentino DF, Bond MW and Mosmann TR: Two types of mouse T-helper cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. J Exp Med *170*: 2081-2095, 1989.
- 16 Wang X, Hagberg H, Mallard C, Zhu C, Hedtjärn M, Tiger CF, Eriksson K, Rosen A and Jacobsson B: Disruption of interleukin-18, but not interleukin-1, increases vulnerability to preterm delivery and fetal mortality after intrauterine inflammation. Am J Pathol 169(3): 967-976, 2006
- 17 Gracie JA, Robertson SE and McInnes IB: Interleukin-18. J Leukoc Biol 73(2): 213-224, 2003.
- 18 Crane JM and Hutchens D: Transvaginal sonographic measurement of cervical length to predict preterm birth in asymptomatic women at increased risk: a systematic review. Ultrasound Obstet Gynecol *31*(*5*): 579-587, 2008.
- 19 Canavan TP, Simhan HN and Caritis S: An evidence-based approach to the evaluation and treatment of premature rupture of membranes: Part I. Obstet Gynecol Surv 59(9): 669-677, 2004.
- 20 Brumfield CG, Lin S, Conner W, Cosper P, Davis RO and Owen J: Pregnancy outcome following genetic amniocentesis at 11-14 versus 16-19 weeks' gestation. Obstet Gynecol 88(1): 114-118, 1996.
- 21 Roper EC, Konje JC, De Chazal RC, Duckett DP, Oppenheimer CA and Taylor DJ: Genetic amniocentesis: gestation-specific pregnancy outcome and comparison of outcome following early and traditional amniocentesis. Prenat Diagn 19(9): 803-807, 1999.
- 22 Holcberg G, Amash A, Sapir O, Sheiner E, Levy S, Myatt L and Huleihel M: Perfusion with lipopolysaccharide differently affects the secretion of interleukin-1 beta and interleukin-1 receptor antagonist by term and preterm human placentae. Placenta 29(7): 593-601, 2008.
- 23 Gotsch F, Romero R, Kusanovic JP, Erez O, Espinoza J, Kim CJ, Vaisbuch E, Than NG, Mazaki-Tovi S, Chaiworapongsa T, Mazor M, Yoon BH, Edwin S, Gomez R, Mittal P, Hassan SS and Sharma S: The anti-inflammatory limb of the immune response in preterm labor, intra-amniotic infection/inflammation, and spontaneous parturition at term: a role for interleukin-10. J Matern Fetal Neonatal Med 21(8): 529-547, 2008.
- 24 Pütz I, Lohbreyer M, Winkler M and Rath W: Appearance of inflammatory cytokines interleukin-1 beta and interleukin-6 in amniotic fluid during labor and in intrauterine pathogen colonization. Z Geburtshilfe Neonatol 202(1): 14-18, 1998.
- 25 Markovic D, Vatish M, Gu M, Slater D, Newton R, Lehnert H and Grammatopoulos DK: The onset of labor alters corticotropin-releasing hormone type 1 receptor variant expression in human myometrium: putative role of interleukin-1beta. Endocrinology 148(7): 3205-13, 2007.
- 26 Chevillard G, Derjuga A, Devost D, Zingg HH and Blank V: Identification of interleukin-1beta regulated genes in uterine smooth muscle cells. Reproduction *134*(*6*): 811-822, 2007.
- 27 Friebe-Hoffmann U, Baston DM, Hoffmann TK, Chiao JP and Rauk PN: The influence of interleukin-1beta on oxytocin signalling in primary cells of human decidua. Regul Pept *142(3)*: 78-85, 2007.
- 28 Orsi NM and Tribe RM: Cytokine networks and the regulation of uterine function in pregnancy and parturition. J Neuroendocrinol *20*(*4*): 462-469, 2008.

- 29 Dunn CL, Kelly RW and Critchley HO: Decidualization of the human endometrial stromal cell: an enigmatic transformation. Reprod Biomed Online 7: 151-161, 2003
- 30 Murray PJ: Understanding and exploiting the endogenous interleukin-10/STAT3-mediated anti-inflammatory response. Curr Opin Pharmacol 6: 379-386, 2006.
- 31 Anderson CF, Oukka M, Kuchroo VJ and Sacks D: CD4(+)CD25(-)Foxp3(-) Th1 cells are the source of IL-10-mediated immune suppression in chronic cutaneous leishmaniasis. J Exp Med 204: 285-297, 2007.
- 32 Benjamin D, Knobloch TJ and Dayton MA: Human B-cell interleukin-10: B-cell lines derived from patients with acquired immunodeficiency syndrome and Burkitt's lymphoma constitutively secrete large quantities of interleukin-10. Blood 80: 1289-1298, 1992.
- 33 de Waal MR, Abrams J, Bennett B, Figdor CG and de Vries JE: Interleukin 10 (IL-10) inhibits cytokine synthesis by human monocytes: An autoregulatory role of IL-10 produced by monocytes. J Exp Med 174: 1209-1220, 1991.
- 34 Boonstra A, Rajsbaum R, Holman M, Marques R, Asselin-Paturel C, Pereira JP, Bates EE, Akira S, Vieira P and Liu YJ: Macrophages and myeloid dendritic cells, but not plasmacytoid dendritic cells, produce IL-10 in response to MyD88- and TRIF-dependent TLR signals, and TLR-independent signals. J Immunol 177: 7551-7558, 2006.
- 35 Dillon S, Agrawal A, Van Dyke T, Landreth G, McCauley L, Koh A, Maliszewski C, Akira S and Pulendran B: A Toll-like receptor 2 ligand stimulates Th2 responses in vivo, via induction of extracellular signal-regulated kinase mitogenactivated protein kinase and c-Fos in dendritic cells. J Immunol 172: 4733-4743, 2004.
- 36 Uhlig HH, Coombes J, Mottet C, Izcue A, Thompson C, Fanger A, Tannapfel A, Fontenot JD, Ramsdell F and Powrie F: Characterization of Foxp3+CD4+CD25+ and IL-10-secreting CD4+CD25+ T-cells during cure of colitis. J Immunol 177: 5852-5860, 2006.
- 37 Hanna N, Hanna I, Hleb M, Wagner E, Dougherty J, Balkundi D, Padbury J and Sharma S: Gestational age-dependent expression of IL-10 and its receptor in human placental tissues and isolated cytotrophoblasts. J Immunol 164: 5721-5728, 2000
- 38 Marvin KW, Keelan JA, Eykholt RL, Sato TA and Mitchell MD: Use of cDNA arrays to generate differential expression profiles for inflammatory genes in human gestational membranes delivered at term and preterm. Mol Hum Reprod 8: 399-408, 2002.
- 39 Gustafsson C, Hummerdal P, Matthiesen L, Berg G, Ekerfelt C and Ernerudh J: Cytokine secretion in decidual mononuclear cells from term human pregnancy with or without labour: ELISPOT detection of IFN-gamma, IL-4, IL-10, TGF-beta and TNF-alpha. J Reprod Immunol 71: 41-56, 2006.
- 40 Lidstrom C, Matthiesen L, Berg G, Sharma S, Ernerudh J and Ekerfelt C: Cytokine secretion patterns of NK cells and macrophages in early human pregnancy decidua and blood: Implications for suppressor macrophages in decidua. Am J Reprod Immunol 50: 444-452, 2003.
- 41 Makhseed M, Raghupathy R, El-Shazly S, Azizieh F, Al-Harmi JA and Al-Azemi MMK: Pro-inflammmatory maternal cytokines profile in preterm delivery. Am J Reprod Immunol 49: 308-318, 2003.

- 42 Newton ER: Preterm labor, preterm premature rupture of membranes and chorioamnionitis. Clin Perinatol *32*: 571-600, 2005
- 43 Thellin O, Coumans B, Zorzi W, Igout A and Heinen E: Tolerance to the foeto-placental 'graft': ten ways to support a child for nine months. Curr Opin Immunol *12*: 731-737, 2000.
- 44 Sato TA, Keelan JA and Mitchell MD: Critical paracrine interactions between TNF-alpha and IL-10 regulate lipopolysaccharide-stimulated human choriodecidual cytokine and prostaglandin E₂ production. J Immunol 170(1): 158-166, 2003
- 45 Zaga-Clavellina V, Garcia-Lopez G, Flores-Herrera H, Espejel-Nuñez A, Flores-Pliego A, Soriano-Becerril D, Maida-Claros R, Merchant-Larios H and Vadillo-Ortega F: *In vitro* secretion profiles of interleukin (IL)-1beta, IL-6, IL-8, IL-10, and TNF alpha after selective infection with *Escherichia coli* in human fetal membranes. Reprod Biol Endocrinol 5: 46, 2007.
- 46 Zaga V, Estrada-Gutierrez G, Beltran-Montoya J, Maida-Claros R, Lopez-Vancell R and Vadillo-Ortega F: Secretion of interleukin-1 beta and tumor necrosis factor alpha by whole fetal membranes depends on initial interactions of amnion or chorion with lipopolysaccharides or group B streptococci. Biol Reprod 71: 1296-1302, 2004.
- 47 Deb k, Chaturvedi MM and Jaiswal YK: Comprehending the role of LPS in Gram-negative bacterial vaginosis: ogling into the causes of unfulfilled child-wish. Arch Gynecol Obstet 270: 133-146, 2004.
- 48 Rivera DL, Olister SM, Liu X, Thompson JH, Zhang XJ, Pennline K, Azuero R, Clark DA and Miller MJ: Interleukin-10 attenuates experimental fetal growth restriction and demise. FASEB J 12: 189-197, 1998.
- 49 Sadowsky DW, Novy MJ, Witkin SS and Gravett MG: Dexamethasone or interleukin-10 blocks interleukin-1betainduced uterine contractions in pregnant rhesus monkeys. Am J Obstet Gynecol 188: 252-263, 2003.
- 50 Terrone DA, Rinehart BK, Granger JP, Barrilleaux PS, Martin JN and Bennet WA: Interleukin-10 administration and bacterial endotoxin-induced preterm birth in a rat model. Obstet Gynecol 98: 476, 2001.
- 51 Robertson SA, Skinner RJ and Care AS: Essential role for IL-10 in resistance to lipopolysaccharide-induced preterm labor in mice. J Immunol 177: 4888-4896, 2006.
- 52 Trautman MS, Collmer D, Edwin SS, Mitchell MD and Dudley DJ: Expression of interleukin-10 in human gestational tissues. J Soc Gynecol Investig 4: 247, 1997.
- 53 Grieg PC, Herbert WNP, Robinette BL and Teot LA: Amniotic fluid interleukin-10 concentrations increase through pregnancy and are elevated in patients with preterm labor associated with intrauterine infection. Am J Obstet Gynecol 173: 1223, 1995.
- 54 Dudley DJ, Edwin SS, Dangerfield A, Jackson K and Trautman MS: Regulation of decidual cell and chorion cell production of interleukin-10 by purified bacterial products. Am J Reprod Immunol 38: 246, 1997.
- 55 Jones CA, Finlay-Jones JJ and Hart PH: Type-1 and type-2 cytokines in human late-gestation decidual tissue. Biol Reprod 57: 303, 1997.
- 56 Simpson KL, Keelan JA and Mitchell MD: Labor-associated changes in interleukin-10 production and its regulation by immunomodulators in human choriodecidua. J Clin Endocrinol Metab 83(12): 4332-4337, 1998.

- 57 Pacora P, Romero R, Maymon E, Gervasi MT, Gomez R, Edwin SS and Yoon BH: Participation of the novel cytokine interleukin 18 in the host response to intra-amniotic infection. Am J Obstet Gynecol 183: 1138-1143, 2000.
- 58 Menon R, Lombardi SJ and Fortunato SJ: IL-18, a product of choriodecidual cells, increases during premature rupture of membranes but fails to turn on the Fas-FasL-mediated apoptosis pathway. J Assist Reprod Genet 18: 276-284, 2001.
- 59 Jacobsson B, Holst RM, Mattsby-Baltzer I, Nikolaitchouk N, Wennerholm UB and Hagberg H: Interleukin-18 in cervical mucus and amniotic fluid: Relationship to microbial invasion of the amniotic fluid, intra-amniotic inflammation and preterm delivery. Brit J Obstet Gynaecol 110: 598-603, 2003.
- 60 Laskarin G, Strbo N, Bogovic CT, Juretic K, Ledee BN, Chaouat G and Rukavina D: Physiological role of IL-15 and IL-18 at the maternal-fetal interface. Chem Immunol Allergy 89: 10-25, 2005.
- 61 Reddy P: Interleukin-18: recent advances. Curr Opin Hematol 11(6): 405-410, 2004

- 62 Emmrich P, Horn LC and Seifert U: Morphologic findings in fetuses and placentas of late abortion in the 2nd trimester. Zentralbl Gynakol 120(8): 399-405, 1998.
- 63 Hook CE, Matyszak MK and Gaston JS: Infection of epithelial and dendritic cells by *Chlamydia trachomatis* results in IL-18 and IL-12 production, leading to interferon-gamma production by human natural killer cells. FEMS Immunol Med Microbiol 45: 113-120, 2005.
- 64 Holst RM, Laurini R, Jacobsson B, Samuelsson E, Sävman K, Doverhag C, Wennerholm UB and Hagberg H: Expression of cytokines and chemokines in cervical and amniotic fluid: relationship to histological chorioamnionitis. J Matern Fetal Neonatal Med 20(12): 885-893, 2007.

Received September 28, 2010 Revised October 26, 2010 Accepted October 28, 2010