

## Effect of EGFR Antagonists Gefitinib (Iressa) and C225 (Cetuximab) on MnSOD-Plasmid Liposome Transgene Radiosensitization of a Murine Squamous Cell Carcinoma Cell Line

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**Abstract.** Radiation therapy of tumors of the head and neck region is compromised by dose limiting toxicity of normal tissues including the oral cavity and oropharyngeal mucosa. MnSOD-Plasmid Liposome (MnSOD-PL) intraoral gene therapy has been demonstrated to decrease normal tissue toxicity and also improve survival in mice with orthotopic SCC-VII squamous cell tumors on the floor of the mouth. Furthermore, intravenous administration of MnSOD-PL in mice with orthotopic tumors, or addition of MnSOD-PL to tumor cell lines *in vitro* produces a radiosensitizing effect attributable to differences in antioxidant pool responses of tumor cells compared to normal tissues following irradiation. To determine whether EGF receptor (EGFR) antagonists Iressa, or Cetuximab provided further improvement of radiation killing of squamous cell tumors, MnSOD-PL transfected or control SCCVII tumor cells were irradiated *in vitro*, and then the effect of EGFR receptor antagonists was tested. Cells transfected with MnSOD-PL were relatively radiosensitive  $D_0=1.244\pm 0.126$  Gy compared to control  $D_0=3.246\pm 0.087$  ( $p<0.0001$ ). Clonogenic radiation survival curves of SCCVII cells demonstrated radiosensitization by Iressa  $D_0=2.770\pm 0.134$  Gy ( $p=0.0264$ ), but no significant radiosensitizing effect of Cetuximab  $D_0=3.193\pm 0.309$  ( $p=0.7338$ ). The combination of MnSOD-PL plus Iressa further increased radiosensitivity of SCC-VII cells *in vitro*  $D_0=0.785\pm 0.01064$  ( $p<0.0001$ ). The results suggest some synergy of the effectiveness of the EGFR antagonist Iressa on increasing the radiation killing of SCC-VII cells that supplements MnSOD-PL tumor radiosensitization.

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**Key Words:** Manganese superoxide dismutase, EGFR antagonist, radiosensitization, tumor redox balance.

The epidermal growth factor receptor (EGFR) signal transduction cascade has been shown to be important in the etiology of squamous cell carcinomas of the head and neck region (1). Expression of EGFR has been found to be of particular importance in the diagnosis of squamous cell carcinomas of the head and neck (2, 3). Targeted therapies against squamous cell tumors have been developed in recent years with a focus on inhibiting the EGFR signal transduction pathway (4, 5). In particular two categories of cytotoxic therapies have been developed. Gefitinib (Iressa) is a small molecule targeting the EGFR (5-7). Another targeted therapy approach has utilized a monoclonal antibody to the EGFR C225, Cetuximab (8), and this agent has been shown to target EGFR positive cells.

Gefitinib (Iressa) has significant effects at inhibiting both squamous cell and hematopoietic cell growth *in vitro* (9-10), and, in early clinical trials, this agent has successfully elicited a therapeutic response in head and neck (11, 12) and lung cancers (13). Clinical trials with Cetuximab for squamous cell carcinoma of the head and neck region have also been very promising (14-17). While there is controversy as to the potential mechanism of action and general utilization of Iressa or Cetuximab (18-22), the relevance of the EGFR signal transduction pathway in human squamous cell cancers has been established (22-24).

Radiotherapy of squamous cell tumors of the head and neck regions is known to produce significant tumor responses, but normal tissue side-effects reflect as mucositis and salivary gland dysfunction (24-28). Tumor cell responses to irradiation include stimulation of cellular repopulation (24, 25) and induction of angiogenic pathways which serve to restimulate tumor recovery from radiation. Angiogenic responses in squamous cell tumors of the head and neck, as well as lung cancer, are known to involve the induction of endothelial cell recruitment by both irradiation and the hypoxic tumor microenvironment (29-32).

Modern radiotherapy techniques including conformal radiotherapy fields (33) and intensity modulation (IMRT)

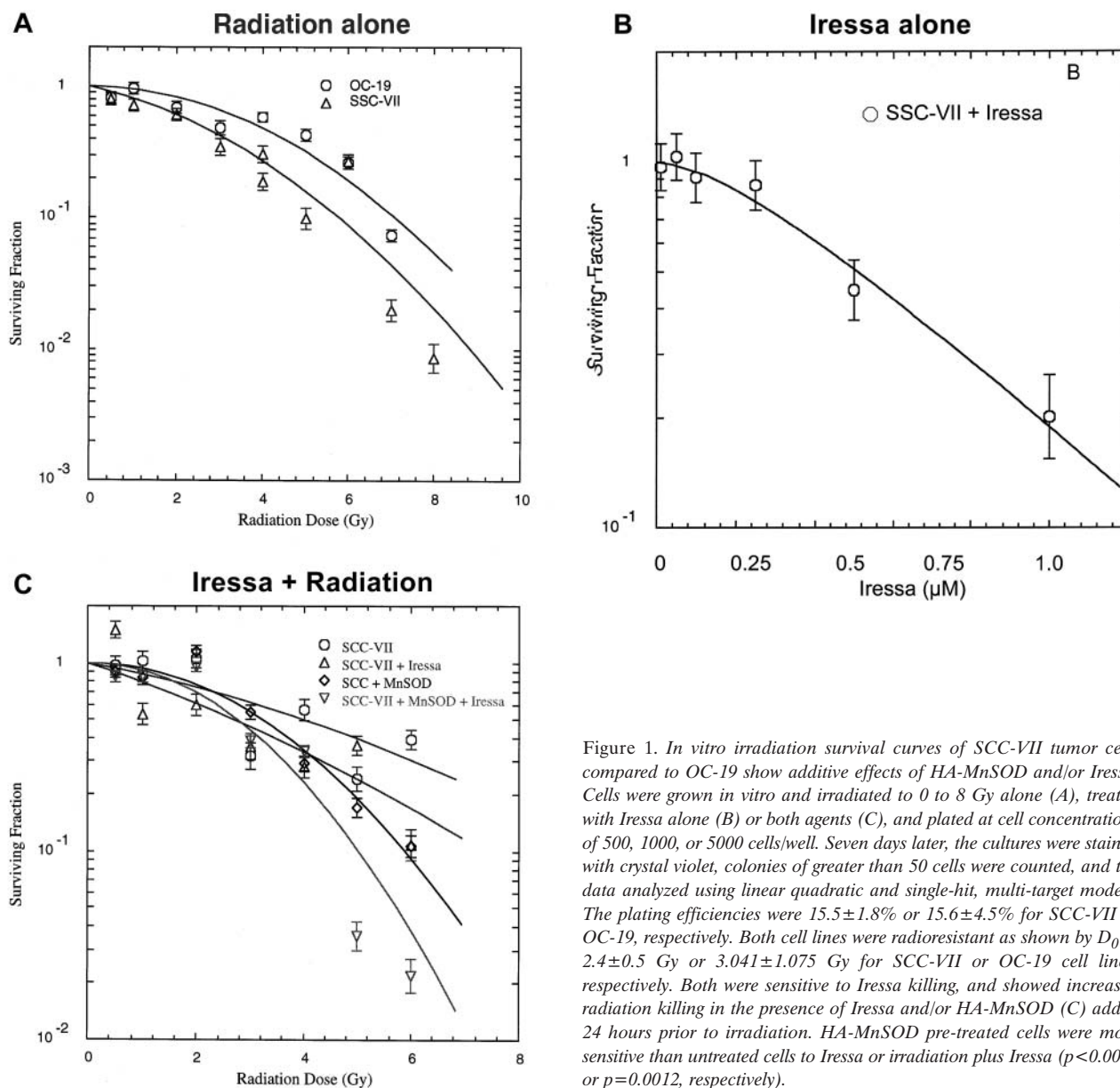


Figure 1. *In vitro* irradiation survival curves of SCC-VII tumor cells compared to OC-19 show additive effects of HA-MnSOD and/or Iressa. Cells were grown *in vitro* and irradiated to 0 to 8 Gy alone (A), treated with Iressa alone (B) or both agents (C), and plated at cell concentrations of 500, 1000, or 5000 cells/well. Seven days later, the cultures were stained with crystal violet, colonies of greater than 50 cells were counted, and the data analyzed using linear quadratic and single-hit, multi-target models. The plating efficiencies were 15.5±1.8% or 15.6±4.5% for SCC-VII or OC-19, respectively. Both cell lines were radioresistant as shown by D<sub>0</sub> of 2.4±0.5 Gy or 3.041±1.075 Gy for SCC-VII or OC-19 cell lines, respectively. Both were sensitive to Iressa killing, and showed increased radiation killing in the presence of Iressa and/or HA-MnSOD (C) added 24 hours prior to irradiation. HA-MnSOD pre-treated cells were more sensitive than untreated cells to Iressa or irradiation plus Iressa (*p*<0.0001 or *p*=0.0012, respectively).

(34-40), have facilitated irradiation dose escalation to treatment volumes in the head and neck region. However, the toxicity of concomitant or sequential chemotherapy necessitates the development of better targeted therapies that can both radiosensitize the tumor and prevent the growth of distant metastasis (41-45). Tumor cells adjacent to normal tissue areas respond to ionizing irradiation with the production of free radicals including: hydroxyl, superoxide, nitric oxide, peroxynitrite and hydrogen peroxide (ROS) causing initial DNA strand breaks, and apoptosis resulting from mitochondrial damage (46, 47). The secondary elaboration of cytokines from irradiation damaged tissues

causes secondary normal tissue damage effects (48, 49). The ROS response to ionizing irradiation may secondarily stimulate the EGF signal transduction pathway as part of the repopulation response to irradiation (50-52).

Normal tissue radioprotective antioxidant gene therapy using MnSOD-Plasmid Liposomes (MnSOD-PL) has been demonstrated to produce paradoxical tumor radiosensitization in orthotopic tumors, perhaps by indirect effects on the tumor repopulation response (46), but also by other mechanisms which may involve different responses of tumor cells compared to normal cells to the oxidative stress induced by irradiation (47).

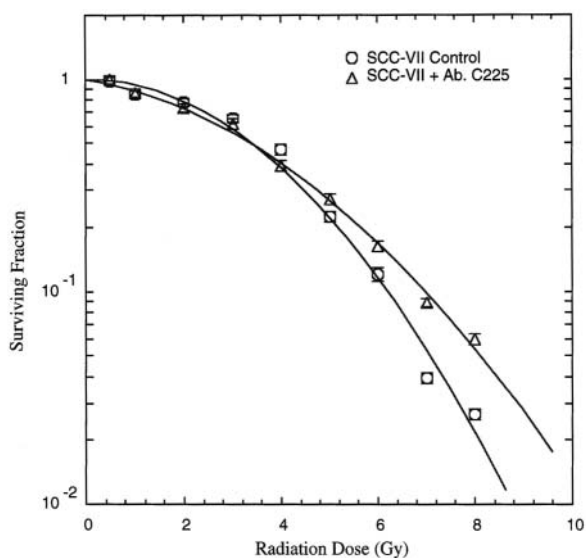


Figure 2. Effect of Cetuximab in irradiation killing of SCC-VII cells *in vitro*. Cells were irradiated after 24 hours incubation in 40  $\mu\text{g/ml}$  C225 or control antibody. Cells were plated at  $5 \times 10^3$ ,  $1 \times 10^4$ , or  $5 \times 10^4$  cells per dish in 4 well tissue culture plates and counted colonies of  $\geq 50$  cells scored at 7 days. Results are presented as described in the methods.

In the present study the SCC-VII squamous cell floor of the mouth murine cancer cell line (63) was utilized to evaluate the potential tumor radiosensitizing effects of Iressa, compared to Cetuximab, and also to determine whether radiosensitization of tumor cells by either agent might further enhance MnSOD-PL mediated tumor radiosensitization (64).

## Materials and Methods

**Cell lines.** The SCC-VII murine squamous cell carcinoma cell line and OC-19, a human squamous cell carcinoma cell line, were passaged *in vitro* according to previously published methods (63). SCC-VII and OC-19 cells were generously provided by Dr. Jennifer Grandis, University of Pittsburgh Cancer Institute, USA.

**Fractionated irradiation.** Cells were irradiated *in vitro* using a 4 MeV linear accelerator (Varian) over doses from 0 to 800 cGy as previously reported (65). Radiation survival curves were plotted according to previously published methods (65, 66) using single-hit multi-target and linear quadratic models.

**Iressa and Cetuximab.** Clinical grade EGFR antagonists Iressa and Cetuximab were generously provided by Dr. Jennifer Grandis, University of Pittsburgh Cancer Institute, and prepared for *in vitro* culture according to the manufacturer's guidelines.

**MnSOD-Plasmid Liposomes (MnSOD-PL).** MnSOD-PL were prepared according to previously published methods (62, 63). SCC-VII cells were transfected *in vitro* 24 hours prior to irradiation. An

epitope-tagged HA-MnSOD was utilized to confirm that over 80% of cells were transfected and were producing transgene protein at the time of the radiation survival curve assays (67).

**Statistics.** Radiation survival curves were evaluated by statistical analysis according to previously published methods (65, 66). Student *t*-test was used to compare the different experimental groups (65, 66).

## Results

**SCC-VII murine squamous cell carcinoma cells are sensitive to Iressa-mediated cytotoxicity.** The radiation survival curve of SCC-VII squamous cell carcinoma cells, and a human squamous cell carcinoma cell line OC-19 were first carried out. Results, shown in Figure 1A, demonstrate significant radioresistance of both cell lines *in vitro*.  $D_0$  of SCC-VII cells were  $3.246 \pm 0.087$  Gy, and that for OC-19 cells was  $3.460 \pm 0.419$  Gy.

Treatment of SCC-VII cells with Iressa (Figure 1B) showed significant cytotoxicity over the dose range of 0.25 to 1.0 micromolar Iressa.

**Iressa supplements MnSOD-PL transfection-mediated radiosensitization of SCC-VII cells *in vitro*.** SCC-VII cells were transfected with MnSOD-PL as reported in "Materials and Methods". Radiation survival curves demonstrated significant radiosensitivity of transfected compared to untreated cells (Figure 1C). HA-MnSOD transfection was utilized to confirm transgene expression in over 80% of the transfected cells and significant radiosensitization production by MnSOD-PL transfection ( $D_0$   $1.249 \pm 0.126$  Gy ( $p < 0.0001$ )).

The addition of Iressa to MnSOD-PL treatment (24 hours prior to irradiation) further increased radiosensitization of SCC-VII cells (Figure 1C)  $D_0$   $0.785 \pm 0.010$  ( $p < 0.0001$ ). These results were significant compared to MnSOD-PL alone or Iressa alone ( $p = 0.0216$  and  $0.0001$ , respectively).

**Cetuximab is not detectably radiosensitizing for SCC-VII cells *in vitro*.** SCC-VII cells were treated *in vitro* with Cetuximab at a concentration of 10 to 40  $\mu\text{g/ml}$ . Over the dose range tested there was no significant spontaneous cell killing by Cetuximab. Irradiation of SCC-VII cells in the presence of 40  $\mu\text{g/ml}$  Cetuximab produced no significant radiosensitization (Figure 2).

## Discussion

Targeted therapies have shown great potential in the treatment of tumors of the head and neck region (19-22). A concern for the addition of targeted therapies to chemoradiotherapy has been over the possible toxicity of these agents, or their exacerbation of radiotherapy/chemotherapy toxicity (53-55). Other targeted therapies including hypoxic

cell sensitizers (Tirapazamine) have shown encouraging results when added to chemotherapy or radiotherapy (56).

Attempts to decrease the toxicity of chemoradiotherapy-induced mucositis have included addition of free radical scavenger compounds, such as Amifostine (Ethyol) (57), Pilocarpine (58), or combinations of agents (58-61).

A recently reported radioprotector of potential therapeutic value is the antioxidant transgene MnSOD, delivered by plasmid liposomes (62-64). MnSOD-PL gene therapy, by targeting transgene product to the mitochondria of oral mucosa cells (65, 66), protects normal tissues, in part through preventing the irradiation-induced cell cycling (67). In the present study MnSOD-PL transfection of a squamous cell tumor cell line of SCC-VII, murine squamous cell carcinoma, induced radiosensitization *in vitro*. These results confirm and extend previously conducted studies (63). Targeted EGFR antagonist Iressa was synergistic with MnSOD-PL in providing further radiosensitization of this tumor cell line *in vitro*. In contrast the monoclonal antibody anti-EGFR Cetuximab was not radiosensitizing for SCC-VII cells *in vitro*. This could be due to the fact that the small molecule Iressa entered cells more rapidly *in vitro* and was able to exert a specific therapeutic toxic effect on the murine squamous cell tumor line resulting in further radiosensitization. This effect was not seen with the antibody treatment. Alternatively, lack of significant expression of an epitope of EGFR, responsive to the Cetuximab monoclonal antibody, on the surface of murine SCC-VII cells may have been responsible for the lack of further radiosensitization. Further studies are required to determine how the successful radiosensitization by Iressa can be translated to potential therapeutic benefit in orthotopic tumor models and normal tissue protection and simultaneous tumor radiosensitization by MnSOD-PL.

## References

- Cohen EW: Role of epidermal growth factor receptor pathway-targeted therapy in patients with recurrent and/or metastatic squamous cell carcinoma of the head and neck. *J Clin Oncol* 24: 2659-2665, 2006.
- Li S, Kim JS, Kim JM, Cho MJ, Yoon WH, Song KS, Yeo SG and Kim JS: Epidermal growth factor receptor as a prognostic factor in locally advanced rectal cancer patients treated with preoperative chemoradiation. *Int J Radiat Oncol Biol Phys* 65: 705-712, 2006.
- Bentzen SM, Atasoy BM, Daley FM, Dische S, Richman PI, Saunders MI, Trott KR and Wilson GD: Epidermal growth factor receptor expression in pretreatment biopsies from head and neck squamous cell carcinoma as a predictive factor for a benefit from accelerated radiation therapy in a randomized controlled trial. *J Clin Oncol* 23: 5560-5567, 2005.
- Wang J, Zhang X, Thomas SM, Grandis JR, Wells A, Chen Z and Ferris RL: Chemokine receptor 7 activates phosphoinositide-3 kinase-mediated invasive and prosurvival pathways in head and neck cancer cells independent of EGFR. *Oncogene* 24: 5897-5904, 2005.
- Chun PY, Feng FY, Scheurer AM, Davis MA, Lawrence TS and Nyati MK: Synergistic effects of gemcitabine and gefitinib in the treatment of head and neck carcinoma. *Cancer Res* 66: 981-990, 2006.
- Ji H, Zhao X, Yuza Y, Shimamura T, Li D, Protopopov A, Jung BL, McNamara K, Xia H, Glatt KA, Thomas RK, Sasaki H, Horner JW, Eck M, Mitchell A, Sun Y, Al-Hashem R, Bronson RT, Rabindran SK, Discifani CM, Maher E, Shapiro GI, Meyerson M and Wong KK: Epidermal growth factor receptor variant III mutations in lung tumorigenesis and sensitivity to tyrosine kinase inhibitors. *PNAS* 103: 7817-7822, 2006.
- Pino MS, Shrader M, Baker CH, Cognetti F, Xiong HQ, Abbruzzese JL and McConkey DJ: Transforming growth factor  $\alpha$  expression drives constitutive epidermal growth factor receptor pathway activation and sensitivity to gefitinib (Iressa) in human pancreatic cancer cell lines. *Cancer Res* 66: 3802-3812, 2006.
- Luwor RB, Lu Y, Li X, Mendelsohn J and Fan Z: The antiepidermal growth factor receptor monoclonal antibody cetuximab/C225 reduces hypoxia-inducible factor-1 alpha, leading to transcriptional inhibition of vascular endothelial growth factor expression. *Oncogene* 24: 4433-4441, 2005.
- Stegmaier K, Corsello SM, Ross KN, Wong JS, DeAngelo DJ and Golub TR: Gefitinib induces myeloid differentiation of acute myeloid leukemia. *Blood* 106: 2841-2848, 2005.
- Kassouf W, Dinney CPN, Brown G, McConkey DJ, Diehl AJ, Bar-Eli M and Adam L: Uncoupling between epidermal growth factor receptor and downstream signals defines resistance to the antiproliferative effect of gefitinib in bladder cancer cells. *Cancer Res* 65: 10524-10535, 2005.
- Wirth LJ, Haddad RI, Lindeman NI, Zhao X, Lee JC, Joshi VA, Norris CM and Posner MR: Phase I study of gefitinib plus celecoxib in recurrent or metastatic squamous cell carcinoma of the head and neck. *J Clin Oncol* 23: 6976-6981, 2005.
- Siegel-Lakhai WS, Beijnen JH and Schellens JHM: Current knowledge and future directions of the selective epidermal growth factor receptor inhibitors erlotinib (Tarceva) and gefitinib (Iressa). *The Oncologist* 10: 579-589, 2005.
- Cappuzzo F, Varella-Garcia M, Shigematsu H, Domenichini I, Bartolini S, Ceresoli GL, Rossi E, Ludovini V, Gregorc V, Toschi L, Franklin WA, Crino L, Gazdar AF, Bunn PA and Hirsch FR: Increased HER2 gene copy number is associated with response to gefitinib therapy in epidermal growth factor receptor-positive non-small cell lung cancer patients. *J Clin Oncol* 23: 5007-5018, 2005.
- Bonner JA, Harari PM, Giralto J, Azarnia N, Shin DM, Cohen RB, Jones CU, Sur R, Raben D, Jassem J, Ove R, Kies MS, Baselga J, Youssoufian H, Amellal N, Rowinsky EK and Ang KK: Radiotherapy plus cetuximab for squamous-cell carcinoma of the head and neck. *N Engl J Med* 354: 567-578, 2006.
- Pfister DG, Su YB, Kraus DH, Wolden SL, Lis E, Alif TB, Zahalsky AJ, Lake S, Needle MN, Shaha AR, Shah JP and Zelefsky MJ: Concurrent cetuximab, cisplatin, and concomitant boost radiotherapy for locoregionally advanced, squamous cell head and neck cancer: a pilot phase II study of a new combined-modality paradigm. *J Clin Oncol* 24: 1072-1078, 2006.
- Li X, Luwor R, Lu Y, Liang K and Fan Z: Enhancement of antitumor activity of the anti-EGF receptor monoclonal antibody cetuximab/C225 by perifosine in PTEN-deficient cancer cells. *Oncogene* 25: 525-535, 2006.

- 17 Dittmann K, Mayer C and Rodemann H-P: Inhibition of radiation-induced EGFR nuclear import by C225 (cetuximab) suppresses DNA-PK activity. *Radiother Oncol* 76: 157-161, 2005.
- 18 Zhang X, Gureasko J, Shen K, Cole PA and Kuriyan J: An allosteric mechanism for activation of the kinase domain of epidermal growth factor receptor. *Cell* 125: 1137-1149, 2006.
- 19 Baumann M and Krause M: Targeting the epidermal growth factor receptor in radiotherapy: radiobiological mechanisms, preclinical and clinical results. *Radiother Oncol* 72: 257-266, 2004.
- 20 Song J and Chen C: Emerging role of EGFR-targeted therapies and radiation in head and neck cancer. *Oncology*, pp. 1757-1767, 2004.
- 21 Posner MR: Paradigm shift in the treatment of head and neck cancer: the role of neoadjuvant chemotherapy. *The Oncologist* 10: 11-19, 2005.
- 22 Saba NF, Khuri FR and Shin DM: Targeting the epidermal growth factor receptor trials in head and neck and lung cancer. *Oncology*, pp. 153, 2006.
- 23 Baselga J and Arteaga CL: Critical update and emerging trends in epidermal growth factor receptor targeting in cancer. *J Clin Oncol* 23: 2445-2459, 2005.
- 24 Lammering G, Valerie K, Lin P-S, Hewit TH and Schmidt-Ullrich RK: Radiation-induced activation of a common variant of EGFR confers enhanced radioresistance. *Radiother Oncol* 72: 267-273, 2004.
- 25 Kim JJ and Tannock IF: Repopulation of cancer cells during therapy: an important cause of treatment failure. *Nature* 5: 516-523, 2005.
- 26 Ciccolini F, Mandl C, Holz-Wenig G, Kehlenbach A and Hellwig A: Prospective isolation of late development multipotent precursors whose migration is prompted by EGFR. *Developmental Biology* 284: 112-125, 2005.
- 27 Olabisi OO, Mahon GM, Kostenko EV, Liu Z, Ozer HL and Whitehead IP: Bcr interacts with components of the endosomal sorting complex required for transport-1 and is required for epidermal growth factor receptor turnover. *Cancer Res* 66: 6250-6260, 2006.
- 28 Pore N, Jiang Z, Gupta A, Cerniglia G, Kao GD and Maity A: EGFR tyrosine kinase inhibitors decrease VEGF expression by both hypoxia-inducible factor (HIF)-1-independent and HIF-1-dependent mechanisms. *Cancer Res* 66: 3197-3204, 2006.
- 29 Nelin LD, Chicoine LG, Reber KM, English BK, Young TL and Liu Y: Cytokine-induced endothelial arginase expression is dependent on epidermal growth factor receptor. *Am J Respir Cell Mol Biol* 33: 394-401, 2005.
- 30 Krause M, Ostermann G, Petersen C, Yaromina A, Hessel F, Harstrick A, van der Kogel AJ, Thames HD and Baumann M: Decreased repopulation as well as increased reoxygenation contribute to the improvement in local control after targeting of the EGFR by C225 during fractionated irradiation. *Radiother Oncol* 76: 162-167, 2005.
- 31 Eicheler W, Krause M, Hessel F, Zips D and Baumann M: Kinetics of EGFR expression during fractionated irradiation varies between different human squamous cell carcinoma lines in nude mice. *Radiother Oncol* 76: 151-156, 2005.
- 32 Toulany M, Dittmann K, Kruger M, Baumann M and Rodemann HP: Radioresistance of K-Ras mutated human tumor cells is mediated through EGFR-dependent activation of P13K-AKT pathway. *Radiother Oncol* 76: 143-150, 2005.
- 33 Bourhis J, Lapeyre M, Tortochaux J, Rives M, Aghili M, Bourdin S, Lesaunier F, Benassi T, Lemanski C, Geoffrois L, Lusinchi A, Verrelle P, Bardet E, Julieron M, Wibault P, Luboinski M and Benhamou E: Phase III randomized trial of very accelerated radiation therapy compared with conventional radiation therapy in squamous cell head and neck cancer: a GORTEC trial. *J Clin Oncol* 24: 2873-2878, 2006.
- 34 Wang D, Schultz CJ, Jursinic PA, Bialkowski M, Zhu XR, Brown WD, Rand SD, Michel MA, Campbell BH, Wong S, Li XA and Wilson JF: Initial experience of FDG-PET/CT guided IMRT of head-and-neck carcinoma. *Int J Radiation Oncol Biol Phys* 65: 143-151, 2006.
- 35 Bensadoun R-J, Benezery K, Dassonville O, Magne N, Poissonnet G, Ramaoli A, Lemanski C, Bourdin S, Tortochaux J, Peyrade F, Marcy P-Y, Chamorey E, Vallicioni J, Seng H, Aizieu C, Gery B, Chauvel P, Schneider M, Santini J, Demard F and Calais G: French multicenter phase III randomized study testing concurrent twice-a-day radiotherapy and cisplatin/5-fluorouracil chemotherapy (BiRCF) in unresectable pharyngeal carcinoma: results at 2 years (FNCLCC-GORTEC). *Int J Radiation Oncol Biol Phys* 64: 983-994, 2006.
- 36 Tishler RB, Posner MR, Norris CM, Mahadevan A, Sullivan C, Goguen L, Wirth LJ, Costello R, Case MA, Stowell S, Sammartino D, Busse PM and Haddad RI: Concurrent weekly docetaxel and concomitant boost radiation therapy in the treatment of locally advanced squamous cell cancer of the head and neck. *Int J Radiation Oncol Biol Phys* 65: 1036-1044, 2006.
- 37 Semrau R, Mueller R-P, Stuetzer H, Staar S, Schroeder U, Guntinas-Lichius O, Kocher M, Eich HT, Dietz A, Flentje M, Rudat V, Volling P, Schroeder M and Eckel HE: Efficacy of intensified hyperfractionated and accelerated radiotherapy and concurrent chemotherapy with carboplatin and 5-fluorouracil: updated results of a randomized multicentric trial in advanced head-and-neck cancer. *Int J Radiation Oncol Biol Phys* 64: 1308-1316, 2006.
- 38 Saarlahti K, Kouri M, Collan J, Hamalainen T, Atula T, Joensuu H and Tenhunen M: Intensity modulated radiotherapy for head and neck cancer: evidence for preserved salivary gland function. *Radiother Oncol* 74: 251-258, 2005.
- 39 Yao M, Dornfeld KJ, Buatti JM, Skwarchuk M, Tan H, Nguyen T, Wacha J, Bayouth JE, Funk GF, Smith RB, Graham SM, Chang K and Hoffman HT: Intensity-modulated radiation treatment for head-and-neck squamous cell carcinoma-the University of Iowa experience. *Int J Radiation Oncol Biol Phys* 63: 410-421, 2005.
- 40 Jabbari S, Kim HM, Feng M, Lin A, Tsien C, Elshaikh M, Terrel JE, Murdoch-Kinch C and Eisbruch A: Matched case-control study of quality of life and xerostomia after intensity-modulated radiotherapy or standard radiotherapy for head-and-neck cancer: initial report. *Int J Radiation Oncol Biol Phys* 63: 725-731, 2005.
- 41 Chinnaiyan P, Huang S, Vallabhaneni G, Armstrong E, Varambally S, Tomlins SA, Chinnaiyan AM and Harari PM: Mechanisms of enhanced radiation response following epidermal growth factor receptor signaling inhibition by Erlotinib (Tarceva). *Cancer Res* 65: 3328-3335, 2005.
- 42 Jimeno A, Rubio-Viqueira B, Amador ML, Oppenheimer D, Bouraoud N, Kulesza P, Sebastiani V, Maitra A and Hidalgo M: Epidermal growth factor receptor dynamics influences response to epidermal growth factor receptor targeted agents. *Cancer Res* 65: 3003-3011, 2005.

- 43 Oda K, Matsuoka Y, Funahashi A and Kitano H: A comprehensive pathway map of epidermal growth factor receptor signaling. *Molecular Systems Biology* 10: 1038, 2005.
- 44 Chen YR, Fu YN, Lin CH, Yang ST, Hu SF, Chen YT, Tsai SF and Huang SF: Distinctive activation patterns in constitutively active and gefitinib-sensitive EGFR mutants. *Oncogene* 25: 1205-1215, 2006.
- 45 Mattila E, Pellinen T, Nevo J, Vuoriluoto K, Arjonen A and Ivaska J: Negative regulation of EGFR signaling through integrin- $\alpha_1\beta_1$ -mediated activation of protein tyrosine phosphatase TCPTP. *Nature Cell Biology* 7: 78-86, 2005.
- 46 Wang M, Kirk JS, Venkataraman S, Domann FE, Zhang HJ, Schafer FQ, Flanagan SW, Weydert CJ, Spitz DR, Buettner GR and Oberley LW: Manganese superoxide dismutase suppresses hypoxic induction of hypoxia-inducible factor-1 $\alpha$  and vascular endothelial growth factor. *Oncogene* 24: 8154-8166, 2005.
- 47 Rosenberg A and Knox S: Radiation sensitization with redox modulators: a promising approach. *Int J Radiation Oncol Biol Phys* 64: 343-354, 2006.
- 48 Mamot C, Drummond DC, Noble CO, Kallab V, Guo Z, Hong K, Kirpotin DB and Park JW: Epidermal growth factor receptor-targeted immunoliposomes significantly enhance the efficacy of multiple anticancer drugs *in vivo*. *Cancer Res* 65: 11631, 2005.
- 49 Perez-Soler R, Delord JP, Halpern A, Kelly K, Krueger J, Sureda BM, von Pawel J, Temel J, Siena S, Soulieres D, Saltz L and Leyden J: HER1/EGFR inhibitor-associated rash: future directions for management and investigation outcomes from the HER1/EGFR inhibitor rash management forum. *The Oncologist* 10: 345-356, 2005.
- 50 Ahmad KA, Iskandar KB, Hirpara JL, Clement MV and Pervaiz S: Hydrogen peroxide-mediated cytosolic acidification is a signal for mitochondrial translocation of Bax during drug-induced apoptosis of tumor cells. *Cancer Res* 64: 7867-7878, 2004.
- 51 Kim YM, Kim KE, Koh GY, Ho Y-S and Lee K-J: Hydrogen peroxide produced by angiopoietin-1 mediates angiogenesis. *Cancer Res* 66: 6167-6174, 2006.
- 52 Koptyra M, Falinski R, Nowicki MO, Stoklosa T, Majsterek I, Nieborowska-Skorska M, Biasiak J and Skorski T: BCR/ABL kinase induces self-mutagenesis via reactive oxygen species to encode imatinib resistance. *Blood* 108: 319-327, 2006.
- 53 Konings AWT, Faber H, Cotteleer F, Vissink A and Coppes RP: Secondary radiation damage as the main cause for unexpected volume effects: a histopathologic study of the parotid gland. *Int J Radiation Oncol Biol Phys* 64: 98-105, 2006.
- 54 Kahn ST and Johnstone PAS: Management of xerostomia related to radiotherapy for head and neck cancer. *Oncology* 19: 1827-1833, 2005.
- 55 Spielberger R, Stiff P, Bensinger W, Gentile T, Weisdorf D, Kewalramani T, Shea T, Yanovich S, Hansen K, Noga S, McCarty J, LeMaistre CF, Sung EC, Blazar BR, Elhardt D, Chen M-G and Emmanouilides C: Palifermin for oral mucositis after intensive therapy for hematologic cancers. *N Engl J Med* 351: 2590-2598, 2004.
- 56 Rischin D, Hicks RJ, Fisher R, Binns D, Corry J, Porceddu S and Peters LJ: Prognostic significance of [ $^{18}\text{F}$ ] - misonidazole positron emission tomography - detected tumor hypoxia in patients with advanced head and neck cancer randomly assigned to chemoradiation with or without tirapazamine: a substudy of trans-tasman radiation oncology group study 98.02. *J Clin Oncol* 24: 2098-2104, 2006.
- 57 Cassatt DR, Fazenbaker CA, Bachy CM, Kifle G and McCarthy MP: Amifostine (ethyol) protects rats from mucositis resulting from fractionated or hyperfractionated radiation exposure. *Int J Radiation Oncol Biol Phys* 61: 901-907, 2005.
- 58 Hakim SG, Kosmehl H, Lauer I, Nadrowitz R, Wedel T and Sieg P: Comparative study on the protection profile of lidocaine, Amifostine, and pilocarpin on the parotid gland during radiotherapy. *Cancer Res* 65: 10486, 2005.
- 59 Buentzel J, Micke O, Adamietz IA, Monnier A, Glatzel M and De Vries A: Intravenous Amifostine during chemoradiotherapy for head-and-neck cancer: a randomized placebo-controlled phase III study. *Int J Radiation Oncol Biol Phys* 64: 684-691, 2006.
- 60 Wasserman TH, Brizel DM, Henke M, Monnier A, Eschwege F, Sauer R and Strnad V: Influence of intravenous Amifostine on xerostomia, tumor control, and survival after radiotherapy for head-and-neck cancer: 2 year followup of a prospective, randomized, phase III trial. *Int J Radiation Oncol Biol Phys* 63: 985-990, 2005.
- 61 Konings AWT, Faber Hette, Vissink A and Coppes RP: Radioprotective effect of Amifostine on parotid gland functioning is region dependent. *Int J Radiation Oncol Biol Phys* 63: 1584-1591, 2005.
- 62 Epperly MW, Defilippi S, Sikora C, Gretton J, Kalend K and Greenberger JS: Intratracheal injection of manganese superoxide dismutase (MnSOD) plasmid/liposomes protects normal lung but not orthotopic tumors from irradiation. *Gene Ther* 7: 1011-1018, 2000.
- 63 Guo HL, Seixas-Silva JA, Epperly MW, Gretton JE, Shin DM and Greenberger JS: Prevention of irradiation-induced oral cavity mucositis by plasmid/liposome delivery of the human manganese superoxide dismutase (MnSOD) transgene. *Radiation Research* 159: 361-370, 2003.
- 64 Guo HL, Epperly MW, Bernarding M, Nie S, Gretton J, Jefferson M and Greenberger JS: Manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) intratracheal gene therapy reduction of irradiation-induced inflammatory cytokines does not protect orthotopic lewis lung carcinomas. *In Vivo* 17: 13-22, 2003.
- 65 Epperly MW, Sikora C, Defilippi S, Gretton J, Zhan Q, Kufe DW and Greenberger JS: MnSOD inhibits irradiation-induced apoptosis by stabilization of the mitochondrial membrane against the effects of SAP kinases p38 and Jnk1 translocation. *Radiation Res* 157: 568-577, 2002.
- 66 Epperly MW, Gretton JE, Bernarding M, Nie S, Rasul B and Greenberger JS: Mitochondrial localization of copper/zinc superoxide dismutase (Cu/ZnSOD) confers radioprotective functions *in vitro* and *in vivo*. *Radiation Res* 160: 568-578, 2003.
- 67 Epperly MW, Carpenter M, Agarwal A, Mitra P, Nie S and Greenberger JS: Intra-oral manganese superoxide dismutase plasmid liposome radioprotective gene therapy decreases ionizing irradiation-induced murine mucosal cell cycling and apoptosis. *In Vivo* 18: 401-410, 2004.

Received August 2, 2006  
Accepted September 4, 2006