The *In Vitro* Activity of Phenothiazines Against *Mycobacterium avium*: Potential of Thioridazine for Therapy of the Co-infected AIDS Patient

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Abstract. Patients presenting with Acquired Immune Deficiency Syndrome (AIDS) are predisposed to co-infection with Mycobacterium avium. The management of such patients is problematic due to underlying immuno-incompetence and the high resistance of M. avium to most non-toxic compounds. Therefore, the need for effective agents is obvious. Because phenothiazines, especially the relatively mild thioridazine, have significant activity against Mycobacterium tuberculosis, we investigated the in vitro activity of chlorpromazine, thioridazine, promazine, promethazine and desipramine against a reference and clinical strains of M. avium. The results obtained show that whereas all of the phenothiazines employed in this study had an minimum inhibitory concentration (MIC) against the strains studied that ranged from ca. 10 to >50 mg/L, as was previously shown for M. tuberculosis, thioridazine was the most active of the group against M. avium.

Phenothiazines have been shown to have *in vitro* activity against sensitive and drug-resistant strains of *Mycobacterium tuberculosis* (1-4). Nevertheless, the concentrations of these agents necessary to inhibit the growth of these bacteria is well beyond that clinically possible (5-8). However, chlorpromazine has activity against intracellular antibiotic-susceptible (9, 10) and antibiotic-resistant intracellular

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Key Words: Mycobacterium avium, phenothiazines, thioridazine, antimycobacterial activities in vitro, AIDS.

strains of *M. tuberculosis* (10) at concentrations in the medium which are similar (9) or below (10) those that are clinically employed. Because chlorpromazine causes frequent and serious side-effects when administered chronically (11), it is not a drug of choice for the management of multidrug-resistant (MDR-TB) infections. However, since the milder phenothiazine, thioridazine, is as effective *in vitro* (3, 4) and *ex vivo* (phagocytosed) MDR-TB (10) as is chlorpromazine (10), the use of thioridazine for the management of MDR-TB, where all other derivatives have failed, has been suggested (7, 12).

Chlorpromazine was shown by Crowle *et al.*, in 1992, to have activity against *M. avium* that had been phagocytosed by human macrophages at concentrations in the medium which were similar to those employed for the therapy of psychosis (9). Based upon this evidence, we studied the *in vitro* activity of chlorpromazine and four other phenothiazines conventionally employed for the management of psychosis against strains of *M. avium*.

Materials and Methods

Materials. Chlorpromazine (CPZ), thioridazine (TZ), promazine (PMZ), promethazine (PMTZ) and desipramine (DSP) were purchased from Sigma Aldrich Quimica SA, Madrid, Spain. Stock solutions of each phenothiazine were freshly prepared, as previously described (4).

Bacteria. The strains of Mycobacterium avium employed in this study were the M. avium CIP 14 031 002-S4 (Pasteur Institute, Paris, France), which has been maintained in our laboratory since 1996, and a clinical isolate obtained from a patient presenting with AIDS. Isolation of M. avium was done in BACTEC MGIT 960 tubes (Middlebrook 7H9 broth) (Becton-Dickinson Diagnostic Instrument Systems, Towson, MD, USA), and identification as the M. avium complex was conducted by the Accuprobe hybridisation probes (Gen-Probe Inc., San Diego, CA, USA), methods routinely

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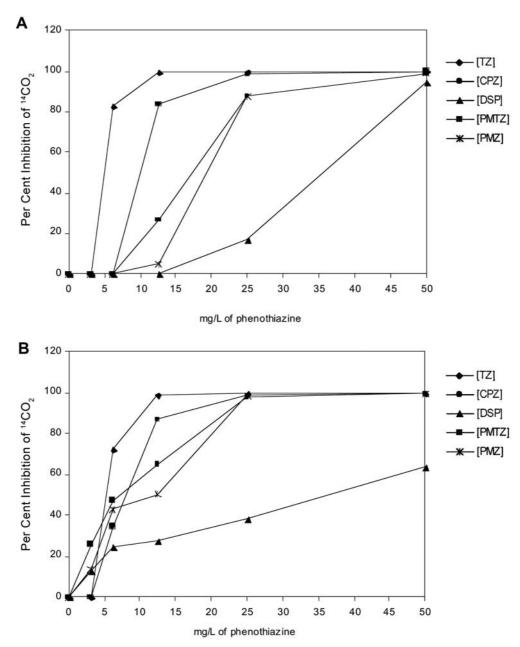


Figure 1. The in vitro activity of chlorpromazine (CPZ), thioridazine (TZ), promazine (PMZ), promethazine (PMTZ) and desipramine (DSP) against Mycobacterium avium. Mycobacterium avium strain CIP 14 031 002-S4 (Pasteur Institute, Paris, France) (A) and a Mycobacterium avium strain isolated from an AIDS patient (B) were incubated in Bactec 12B vials that contained concentrations of each phenothiazine that ranged from 0.0 to 50 mg/L. The experiment was repeated twice and essentially the same quantitative data was obtained. The results represent the data obtained from one such experiment and describe the activity against both strains.

employed in the Mycobacteriology Laboratory of the Institute of Hygiene and Tropical Medicine of Lisbon, Portugal (13).

Method for assessment of the in vitro effects of the phenothiazines against M. avium. M. avium strains were cultured in Middlebrook 7H9 broth, Bactec 12B vials (Becton-Dickinson Diagnostic Instrument Systems), until maximum growth was achieved. From these cultures, aliquots were transferred to vials and the optical

density adjusted to a 0.5 McFarland standard. These vials served as the source for the inoculation of triplicate Bactec 12B vials that contained concentrations of each phenothiazine that ranged from 0.0 to 50 mg/L. A separate 1:100 and 1:1000 dilution of the adjusted inoculum was made and transferred to Bactec 12B vials that contained no drug; this served as the proportional control as defined by the recommended Bactec proportional method for *M. avium* (14). The vials were incubated at 37°C and the contents

periodically assessed by the Bactec 460 TB instrument (Becton-Dickinson Diagnostic Instrument Systems) for ¹⁴CO₂ generated from the metabolism of ¹⁴C palmitic acid. The method for assessment of growth has been previously described (2, 4, 6). The MIC that completely inhibits the growth of each strain is defined for each of the phenothiazines evaluated (4).

Results

The in vitro activity of the phenothiazines CPZ, TZ, PMZ, PMTZ and DSP are presented in Figure 1. Briefly, the ascending order of activity, as defined by the MIC (lowest concentration that produces 100% inhibition of growth assessed by the inhibition of the ¹⁴CO₂ generated) in mg/L, was as follows: TZ (10), CPZ (25), PMZ (25-50), PMTZ (25-50) and DSP (>50). Because it has generally been assumed that the resistance of M. avium to drugs employed for the management of pulmonary tuberculosis produced by M. tuberculosis is due to the cell wall of the former, it was surprising to discover that M. avium is more sensitive to TZ than is M. tuberculosis, the latter being completed inhibited in its growth by concentrations of TZ that exceed 25 mg/L (4). However, with respect to CPZ, PMZ, PMTZ and DSP, the sensitivity of M. avium to these agents is practically identical to that produced against M. tuberculosis (4). Because the assays were conducted in triplicate and the experiments repeated at least twice, the difference between the MICs of TZ for M. avium versus M. tuberculosis were considered to be significant.

Discussion

Patients presenting with AIDS are at risk of acquiring a variety of opportunistic infections (15). Although each of these infections may cause death to any given AIDS patient, pulmonary infections caused by *M. avium* produce the highest mortality (15, 16) since effective drugs are few and relatively ineffective against this organism (16). Currently, azithromycin and clarithromycin are the two most effective agents for the therapy of *M. avium* pulmonary nontubercular infections (16). However, there is now evidence that these agents produce ototoxicity (17) and arrhythmia (18), respectively. The need for effective drugs for the management of this infection is obvious.

The results obtained in our current study indicate that each of the phenothiazines studied had *in vitro* activity against the ATCC and clinical strains. The response of the reference and clinical strains was quantitatively very similar, thereby suggesting that other clinical strains will show similar responses. The order of increasing effectiveness against these strains was TZ, CPZ, PMZ, PMTZ and DSP. TZ was by far the most effective of the phenothiazines evaluated, presenting with an MIC of *ca.* 10 mg/L. Surprisingly, *M. avium* is far more sensitive to the phenothiazine TZ than are

strains of *M. tuberculosis* (7, 8). Nevertheless, the MIC of 6 mg/L for TZ is not clinically achievable. However, TZ does enhance the killing of intracellular *M. tuberculosis* (10) when the concentration of this agent in the medium is of the order of 0.1 mg/L and, hence, within clinical range and below any associated with toxicity (10, 19, 20). Killing is considered to be the result of the agent being concentrated by the macrophage to levels equivalent to those that produced *in vitro* killing (10, 19, 20) and, because phenothiazines are concentrated by lysosomes (21, 22), the killing of the bacterium by macrophages that have little killing activity of their own (10) is probably due to the fusion of the phagosome with the lysosome containing the highly concentrated phenothiazine (10, 19, 20).

The greater sensitivity of *M. avium* to TZ as opposed to that produced against *M. tuberculosis* is interesting. Although at this time we cannot provide an adequate explanation for this difference, we can confidently conclude that the cell wall of *M. avium*, generally considered to be the cause for its resistance to anti-TB agents, does not lead to greater resistance to TZ.

The results of the current study are not sufficient to recommend that TZ be used for the management of AIDS patients co-infected with *M. avium*. Furthermore, even when the study that evaluated the killing activity of TZ against phagocytosed *M. avium* showed that the concentration required for such killing was comparable to that expected to be present in the plasma of patients chronically treated with this agent, the use of the mild TZ is not without risk, as shown by recent studies demonstrating that, for a very small number of patients, TZ is associated with serious arrhythmia such as "torsade de pointes" (23). However, the relationship is now in question since the number of risk factors presented by the patient may be more significant for the arrhythmia than that produced by the agent itself (24).

Conclusion

The phenothiazines CPZ, TZ, PMZ, PMTZ and DSP have *in vitro* activity against *M. avium*. TZ was the most effective of the phenothiazines, having an MIC of *ca.* 10 mg/L. Although TZ seems to be the drug of choice among the phenothiazines tested, it is not at this time recommended for the management of AIDS patients coinfected with *M. avium*.

Acknowledgements

We would like to thank the Institute of Hygiene and Tropical Medicine of Lisbon, Portugal and its Scientific Council for the support given to this project. Special thanks are to be given to the Management Committee Members of Cost Action B16 of the European Commission for their many helpful suggestions. This work was supported by grant EU-FSE/FEDER-POCTI-37579/FCB/2001

provided by the Fundação para a Ciência e a Tecnologia (FCT) of Portugal. M. Martins was supported by grant SFRH/BD/14319/2003 from the Fundação para a Ciência e a Tecnologia (FCT) of Portugal.

References

- 1 Kristiansen JE and Vergmann B: The antibacterial effect of selected phenothiazines and thioxanthenes on slow-growing mycobacteria. Acta Pathol Microbiol Immunol Scand (B) 94: 393-398, 1986.
- 2 Amaral L, Kristiansen JE, Abebe LS and Millet W: Inhibition of the respiration of multi-drug resistant clinical isolates of *Mycobacterium tuberculosis* by thioridazine: potential use for the initial therapy of freshly diagnosed tuberculosis. J Antimicrob Chemother 38: 1049-1053, 1996.
- 3 Kristiansen JE and Amaral L: The potential management of resistant infections with non-antibiotics. J Antimicrob Chemother 40: 319-327, 1997.
- 4 Bettencourt-Viveiros M, Bosne-David S and Amaral L: Comparative activity of phenothiazines against multi-drug resistant Mycobacterium tuberculosis. Int J Antimicrob Agents 16: 69-71, 2000.
- 5 Amaral L and Kristiansen JE: Phenothiazines: an alternative to conventional management of suspect multidrug resistant tuberculosis. Int J Antimicrob Agents 14: 173-176, 2000.
- 6 Viveiros M and Amaral L: Enhancement of antibiotic activity against poly-drug resistant *Mycobacterium tuberculosis* by phenothiazines. Int J Antimicrob Agents 17: 225-228, 2001.
- 7 Amaral L, Kristiansen JE, Viveiros M and Atouguia J: Activity of phenothiazines against antibiotic resistant *Mycobacterium* tuberculosis: a review supporting further studies that may elucidate the potential use of thioridazine as an anti-TB agent. J Antimicrob Chemother 47: 505-507, 2001.
- 8 Amaral L, Viveiros M and Kristiansen JE: Phenothiazines: potential alternatives for the management of antibiotic resistant infections of tuberculosis and malaria in developing countries. Trop Med Int Health 6: 1016-1022, 2001.
- 9 Crowle AJ, Douvas GS and May HH: Chlorpromazine: a drug potentially useful for treating mycobacterial infections. Exp Chemother 38: 410-419, 1992.
- 10 Ordway D, Viveiros M, Leandro C and Amaral L: Clinical concentrations of thioridazine kill intracellular multi-drug resistant *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 47: 917-922, 2003.
- 11 Amaral L and Kristiansen JE: Phenothiazines: potential management of Creutzfeldt Jakob Disease and its variants. Int J Antimicrob Agents 18: 411-417, 2001.
- 12 Amaral L, Viveiros M and Molnar J: Antimicrobial activity of phenothiazines. In Vivo 18: 725-732, 2004.
- 13 Reisner BS, Gatson AM and Wood GL: Use of Gen-Probe AccuProbes to identify *Mycobacterium avium* complex, *Mycobacterium tuberculosis* complex, *Mycobacterium kansasii*, and *Mycobacterium gordonae* directly from BACTEC TB broth cultures. J Clin Microbiol *32* 4: 2995-2998, 1994.

- 14 Inderlied CB and Nash KA: Antimycobacterial agents: in vitro susceptibility testing, spectra of activity, mechanisms of action and resistance, and assays for activity in biologic fluids. In: Lorian V (ed.): Antibiotics in Laboratory Medicine, 4th ed. Baltimore: Williams and Wilkins, pp. 127-175, 1996.
- 15 Bellamy R, Sangeetha S and Paton NI: Causes of death among patients with HIV in Singapore from 1985 to 2001: results from the Singapore HIV Observational Cohort Study (SHOCS). HIV Med 5: 289-295, 2004.
- 16 Hoffmann T and Brunner H: Model for simulation of HIV/AIDS and cost-effectiveness of preventing nontuberculous mycobacterial (MAC)-disease. Eur J Health Econ 5: 129-135, 2004.
- 17 Uzun C, Koten M, Adali MK, Yorulmaz F, Yagiz R and Karasalihoglu AR: Reversible ototoxic effect of azithromycin and clarithromycin on transiently evoked otoacoustic emissions in guinea pigs. J Laryngol Otol 115: 622-628, 2001.
- 18 Ohtani H, Taninaka C, Hanada E, Kotaki H, Sato H, Sawada Y and Iga T: Comparative pharmacodynamic analysis of Q-T interval prolongation induced by the macrolides clarithromycin, roxithromycin, and azithromycin in rats. Antimicrob Agents Chemother 44: 2630-2637, 2000.
- 19 Ordway D, Viveiros M, Leandro C, Jorge Arroz M, Molnar J, Kristiansen JE and Amaral L: Chlorpromazine has intracellular killing activity against phagocytosed *Staphylococcus aureus* at clinical concentrations. J Infect Chemother 8: 227-231, 2002.
- 20 Ordway D, Viveiros M, Leandro C, Arroz MJ and Amaral L: Intracellular activity of clinical concentrations of phenothiazines including thioridiazine against phagocytosed *Staphylococcus aureus*. Int J Antimicrob Agents 20: 34-43, 2002.
- 21 Daniel WA, Wojcikowski J and Palucha A: Intracellular distribution of psychotropic drugs in the grey and white matter of the brain: the role of lysosomal trapping. Br J Pharmacol *134*: 807-814, 2001.
- 22 Wojcikowski J and Daniel WA: Thioridazine-fluoxetine interaction at the level of the distribution process in vivo. Pol J Pharmacol 54: 647-654, 2002.
- 23 Vieweg WV and Wood MA: Tricyclic antidepressants, QT interval prolongation, and torsade de pointes. Psychosomatics 45: 371-377, 2004.
- 24 Titier K, Girodet PO, Verdoux H, Molimard M, Begaud B, Haverkamp W, Lader M and Moore N: Atypical antipsychotics: from potassium channels to torsade de pointes and sudden death. Drug Saf 28: 35-51, 2005.

Received March 1, 2005 Accepted May 4, 2005