

Review

Natural Compounds and Neuroprotection: Mechanisms of Action and Novel Delivery Systems

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Abstract. Neurodegeneration characterizes pathologic conditions, ranging from Alzheimer's disease to glaucoma, with devastating social and economic effects. It is a complex process implicating a series of molecular and cellular events, such as oxidative stress, mitochondrial dysfunction, protein misfolding, excitotoxicity and inflammation. Natural compounds, because of their broad spectrum of pharmacological and biological activities, could be possible candidates for the management of such multifactorial morbidities. However, their therapeutic potential against neurodegenerative diseases has been hampered by their poor bioavailability and subsequent insufficient delivery to the brain. This article provides an overview of the molecular mechanisms through which natural compounds exert their neuroprotective effects, as well as the development of novel natural compound-loaded delivery systems that could improve their neuroavailability.

Neurodegenerative diseases are characterized by progressive loss of structure or function of neurons and include a broad range of conditions, from Parkinson's (PD), Alzheimer's (AD), Huntington's disease (HD) to glaucoma (1). They represent rapidly growing causes of disability and even death, having profound social and economic implications. Neurodegeneration is the result of a complex cascade of

pathological events, including oxidative stress, mitochondrial dysfunction, inflammation and protein aggregation (2, 3). The increasing knowledge of the cellular and molecular events underlying the degenerative process has greatly stimulated research for identifying compounds capable of stopping or, at least, slowing the progress of neural deterioration.

Natural compounds are complex chemical multiple-target molecules found mainly in plants and microorganisms (4). These agents have been extensively studied regarding their antioxidant activities. However, in addition to their ability to prevent damage caused by oxidative stress, they have been shown to modulate multiple signal transduction pathways through direct effects on enzymes, such as kinases, regulatory proteins and receptors (4, 5). Furthermore, it has been suggested that many polyphenols exert some of their beneficial biological effects *via* chromatin remodeling and epigenetic modifications (6). This broad spectrum of pharmacological or biological activities has made them suitable candidates for the treatment of multifactorial diseases, such as cancer and neurodegenerative diseases (7-9). Indeed, there are studies suggesting a correlation between consumption of flavonoids and low population rates of dementia (10). However, their physicochemical properties are not drug-like and a number of challenges, concerning their stability and neuroavailability, need to be overcome in order to be established as effective therapeutics.

This review focuses on the neuroprotective role that natural compounds have through different molecular mechanisms on the biological processes involved in neurodegenerative diseases. Furthermore, the development of novel delivery systems, that could improve the neuroavailability and subsequently the neuroprotective activity of the natural compounds, is discussed.

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Neuroprotective Targets of Natural Compounds

Mitochondrial Dysfunction

Neurons are heavily dependent on mitochondria for survival because of their high energy requirement. Mitochondria are important in various essential cellular functions, including apoptosis, metabolism and calcium homeostasis. Regarding oxidative stress, they are both an important source of reactive oxygen species (ROS) production and a major target for ROS-induced cellular injury. Given the role of mitochondria as key regulators of cellular death and life, it is expected that alterations in their biology have implications in a wide array of diseases, such as neurodegenerative ones (11-13). A direct link between mitochondrial dysfunction and neurotoxic manifestations of anticancer drugs has also been established (14).

SIRT1, a sirtuin protein family member, is a nicotinamide adenine dinucleotide (NAD⁺)-dependent histone and protein deacetylase. A large number of studies have reported that activation and overexpression of SIRT1 are neuroprotective in both acute central nervous system (CNS) injuries and chronic neurodegenerative diseases (15). AMP-activated protein kinase (AMPK) enhances SIRT activity by increasing cellular NAD⁺ levels leading to the deacetylation and modulation of the activity of downstream SIRT1 targets like peroxisome proliferator-activated receptor- γ coactivator 1 alpha (PGC-1 α) (16), a master regulator of mitochondrial biogenesis (17). Therefore, the AMPK/SIRT1/PGC-1 α axis is strongly related to the orchestration of mitochondrial function and energy/redox status in mammalian cells.

Nature has been a major source of substances that can improve mitochondrial function (18). Among them polyphenols, such as luteolin, myricitrin, quercetin (QCT) and epigallocatechin gallate (EGCG) have been extensively studied regarding their beneficial effects on mitochondrial function both *in vitro* and *in vivo* (19-22). In addition to these, proanthocyanidins, a group of polyphenolic bioflavonoids, ameliorate the hydrogen peroxide-induced mitochondrial dysfunction *via* stimulating the mitochondrial membrane potential (MMP) and respiratory chain complex IV, while moderating the mitochondrial free radical production, ROS and mitochondrial superoxide (23). Also, hesperidin has been reported to enhance the mitochondrial complex I-IV enzymatic potential (22). On the other hand, resveratrol (RSV) may act directly or indirectly on the mitochondria with subsequent beneficial effects (reviewed by de Oliveira *et al.* (24)). The mechanism by which RSV modulates mitochondrial function is through regulation of cell signaling pathways and genes involved in mitochondrial biogenesis, endogenous antioxidant defense and oxidative phosphorylation (25, 26). In particular, RSV regulates the gene expression of antioxidant enzymes, such as manganese-

dependent superoxide dismutase (*Mn-SOD*) and modulates uncoupling protein 2 (UCP2) protein levels in mitochondrial membranes, both important in the redox maintenance of the organelle, possibly through SIRT1 activation (24, 27). It has also been shown in different *in vivo* and *in vitro* experimental models that the AMPK-SIRT1 pathway plays a pivot role in RSV-induced neuroprotection (28-30). Furthermore, co-administration of EGCG and RSV reversed the severe impairment of mitochondrial bioenergetics and biogenesis in hippocampal progenitor cells from a Down syndrome mouse model *via* activation of the AMPK/Sirt1/PGC-1 α axis as well (31). Finally, the neuroprotective role of viniferin, a stilbene RSV dimer, in an HD model was shown to be mediated through the AMPK/SIRT3 pathway (32).

Concerning anti-neoplastic agent-induced neurotoxicity, natural compounds have also been studied. Co-administration of curcumin (CUR) and QCT mitigated peripheral neurotoxicity induced by oxaliplatin through the restoration of glutathione S-transferase (GST), glutathione peroxidase (GPx) and MnSOD activity (11, 14, 33).

Neurodegeneration, as a complication of diabetes, it is known to be associated with dicarbonyl glycation and methylglyoxal (MGO), a major precursor of advanced glycation end products (AGEs), which target mainly mitochondrial proteins. Several *in vivo* and *in vitro* studies have pointed out that flavonoids could inhibit the formation of AGEs and prevent diabetic neurodegeneration (34, 35). Moreover, myricitrin was shown to alleviate MGO-induced mitochondrial dysfunction, possibly through modulation of the AGEs/Receptor for AGEs (RAGE)/Nuclear factor kappa B (NF- κ B) pathway (36).

Apoptosis

Apoptotic neuronal death is a common feature in the brain of patients suffering from many neurodegenerative diseases and either intrinsic (mitochondrial-mediated) or extrinsic (death receptor-mediated) (37) is controlled by several proteins (38). The Bcl-2 protein family plays an essential role in the regulation of the intrinsic pathway *via* monitoring mitochondrial membrane permeability and the release of the pro-apoptotic factor, cytochrome c, which promotes the caspase-9 activation (39, 40). Bcl-2 proteins include members that inhibit apoptosis (such as Bcl-2, Bcl-w, Bcl-xL, Mcl-1) or promote apoptosis (such as Bak, Bad, Bax, Bcl-rambo). It is the balance between these members that determines whether or not a cell will undergo apoptosis. On the other hand, the caspase cascade may be initiated at the endoplasmic reticulum (ER) as well (40) by stress conditions, which affect the folding of proteins in ER lumen (41, 42). Finally, in addition to ER-resident proteins and mitochondria, several stress-sensing transcription factors are stimulated and implicated in AD pathogenesis (43).

Many natural compounds with neuroprotective effects have been shown to act by directly affecting programmed cell death pathways. Bilobalide, the main terpenoid of Ginkgo biloba leaves, showed potent protective effects on neurons and Schwann cells (44), prevented ROS-induced apoptosis in early stages and attenuated the elevation of Bax, p53 and caspase-3 in PC12 cells (43, 45). Furthermore, bilobalide modified mitochondrial function through up-regulation of cytochrome c oxidase subunit I (46). Huperzine A, a sesquiterpene alkaloid, has been shown to inhibit apoptotic factors, such as caspase-3, Bax and p53 (47). Finally, ER stress-induced apoptosis was impeded by the bioflavonoid apigenin, through suppression of ROS accumulation, inhibition of caspase-12 and -3 activation and cleavage of poly (ADP-ribose) polymerase (PARP).

Excitotoxicity

Glutamate is a main excitatory neuro-transmitter in the CNS, including the retina (48). It is released in the presynaptic terminals for very brief periods of time and binds to a variety of receptor-linked channels in the postsynaptic membrane, resulting in the influx of Ca^{++} and the initiation of the action potential (49). There are three classes of glutamate-gated ion channels, known as α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA), kainate and N-methyl-D aspartate (NMDA) receptors. Glutamate presence in excessive amounts or for excessive periods of time can literally excite cells to apoptotic cell death, mainly due to high Ca^{++} level in the cytosol (50, 51). Furthermore, the disruption of energy metabolism during acute and chronic neurodegenerative disorders may lead to inefficient glutamate clearance or even inappropriate release, which cause elevated levels of glutamate and increased concentrations of cytosolic Ca^{++} (52). Finally, excitotoxicity can arise even with normal levels of glutamate, if NMDA receptor is activated (53).

Glutamate receptor-mediated excitotoxicity has been associated with several diseases of the brain, whereas *in vivo* and *in vitro* studies have shown that blocking the NMDA and the non-NMDA receptors simultaneously results in maximum protection against ischemic neurodegeneration (54). (-)-kainocephalin, a natural compound isolated from the fungus *Eupenicillium shearii*, has been shown to be a potent antagonist of particular subtypes of glutamate receptors (AMPA and NMDA, but not kainate) and to protect CNS neurons from excitotoxicity (55). Furthermore, myricetin inhibited glutamate-induced excitotoxicity in neurons; specifically, it affected NMDAR receptor (NMDAR) phosphorylation, which had, as a result, reduction of intracellular Ca^{++} overload. Myricetin also inhibited the glutamate-induced ROS production and the activity of caspase-3 by interacting with it. (56). On the other hand,

acacetin has been shown to effectively prevent kainic acid-induced *in vivo* excitotoxicity by inhibiting glutamate release from rat hippocampal synaptosomes *via* attenuation of voltage-dependent Ca^{++} entry (57).

Inflammation

Chronic inflammation in the CNS has been shown to be related to neuronal injury and death in neurodegenerative diseases (58). Activated microglia, the resident immune cells of the CNS, are one of the prime participants in neuroinflammation (59) and thought to contribute to neuronal death through the production of reactive nitrogen species (RNS) and ROS (60). The fibrillar β -amyloid peptide ($A\beta$) deposits also play a crucial role in tissue neuroinflammation (61).

NF-kappa B/Rel proteins are dimeric, sequence-specific transcription factors involved in the activation of a remarkably large number of genes, in response to tumor necrosis factor α (TNF- α), lipopolysaccharide (LPS) and other stressful stimuli that require rapid reprogramming of gene expression (62). NF-kappa B is sequestered and inactive in the cytoplasm bound to inhibitory I-kappa B proteins in unstimulated cells. Induction results in phosphorylation, ubiquitinylation and, ultimately, proteolytic degradation of Ikappa B and subsequent release and translocation of NF-kappa B to the nucleus where it activates the transcription of specific target genes (62). NF-kappaB induces production of various mediators, such as nitric oxide (NO), and regulates a number of inflammation- and oxidative stress-related genes, such as cyclooxygenase 2 (COX-2) (43, 63). The activation of NF-kappaB, particularly in the CNS, has been shown to trigger multicellular responses, including transactivation of inflammatory molecules and production of free radicals in glial cells, which are intricately associated with the initiation and progression of neurodegenerative diseases. (63). However, the final effect of NF-kappa B stimulation on neuronal survival and death potentially depends on parameters, such as cell type, developmental stages of cells, type of signal and the nature of activated NF-kappaB dimers (64-66).

Based on the fact that inflammation contributes to the continued loss of CNS neurons, even if in some cases it is not the primary causative process, the use of anti-inflammatory compounds has been proposed for diminishing the cumulative effects of inflammation in the brain (67). Flavonoids have been reported to exert their anti-inflammatory effect through intervention in a broad range of molecular pathways related to inflammation (68, 69), such as the release of pro-inflammatory cytokines and microglia activation (70). Daidzein and possibly other isoflavones as well have been reported to be neuroprotective due to their ability to dampen the induction of microglial activation and

subsequent release of soluble pro-inflammatory factors by inhibiting the oxidative induction of the p38 mitogen-activated protein kinase (MAPK)-NF-kappa B pathway (71).

Moreover, flavonoids, including EGCG, have been reported to modulate T cell response partially through inhibition of NF-kappa B signaling (72-74). In addition, a transgenic mouse model of amyotrophic lateral sclerosis treated with EGCG displayed increased neuron survival and diminished microglial activation (75).

Several studies have shown that NF-kappa B signal transduction is down-regulated by flavonoids (68) through activation of extracellular signal-related kinase 1,2 (ERK1/2) or inhibition of the IkappaB kinase activity (76). Since COX-2 is synthesized through an NF-kappa B-mediated pathway, another important target of flavonoids accounting for their anti-inflammatory effect could be COX-2 at the expression level. Indeed, many flavonoids were reported to be efficient COX inhibitors (68). On the other hand, polyphenols, such as CUR, RSV and catechins, have been reported to affect nuclear NF-kappaB expression and chromatin remodeling through modulation of histone deacetylase (HDACs) and DNA methyl transferase (DNMTs) activities (6). Interestingly, regarding CUR, it has been shown that it can also inhibit NF-kappaB signaling by acting as an agonist of the peroxisome proliferator-activated receptor γ (PPAR- γ) (77, 78).

Additionally, flavonoids have been shown to inhibit LPS-induced production of inflammatory cytokines in human monocytes (79) and astrocytes (80). Moreover, they have been demonstrated to inhibit the activity of kinases, such as protein kinase C (PKC), p38 MAPK and Jun N-terminal kinase/stress-activated protein kinase, that are important partners in inflammatory signal transduction pathways (81, 82).

Oxidative Stress

Oxidative stress is recognized as an essential factor in a variety of neurodegenerative diseases as a mechanism for age-related degenerative processes and as a mediator of the adverse effects of neurotoxicants (83). Oxidative stress causes damage to proteins, lipids, DNA and occurs when ROS accumulate in cells, from either excessive production or insufficient neutralization, due to an imbalance between antioxidant defense systems and production of ROS (84). Production of ROS and RNS may lead to irreversible deleterious modification of macromolecules like proteins and neuronal cell death. The brain is sensitive to accumulation of these reactive species due to its inadequate ability to neutralize their effects (85). In specific, it retains high concentration of transition metals, high aerobic metabolism and increased levels of ascorbic acid, all of them contributing to production of ROS/RNS and oxidized products (86), whereas it does not have a competent antioxidant defense system characterized by moderate

activity of antioxidant enzymes, such as superoxide dismutases (SODs) and catalase, as well as low levels of glutathione (87). Finally, many neurotransmitters are autoxidized to generate ROS and RNS like NO (86). In addition to these, another contributing factor to oxidative stress in neurodegenerative disorders is inflammation caused by activated microglia (86, 88).

The nuclear factor E2-related factor 2 (Nrf2) signaling pathway is primarily responsible for cellular defense against oxidative stress (89). Nrf2 is a fundamental transcription factor implicated in transcriptional activation of phase II detoxifying enzymes through antioxidant response element (ARE) (90, 91). It has also been reported that Nrf2 activation, in macrophages and microglia, down-regulates the NF-kappa B-related inflammatory responses (92, 93).

Natural compounds have received much attention as potent antioxidants and polyphenols have been the most investigated (94). Phenolic compounds, such as flavonoids, rosmarinic acid, ferulic, caffeic, chlorogenic, vanillic, p-hydroxybenzoic acid, protocatechuic acid and p-coumaric acid, were identified to contribute to the antioxidant potential by various scavenging assays (83, 86). Besides enhancing the efficiency of antioxidant gene regulation, natural antioxidants also exert their effects through additional mechanisms of action, including hydrogen atom transfer, electron donation, direct radical scavenging, metal chelation, restoration of endogenous antioxidant levels, activation of antioxidant enzymes, singlet oxygen quenching and activation of Nrf2 pathway (reviewed by Fraubberger *et al.* (95)).

Protein Misfolding

Most neurodegenerative disorders share a common feature. A protein accumulates in an insoluble form in the affected tissue (96). The identity of the proteins and the site of the deposits are distinguishing for each particular disorder (97). However, the majority of the affected proteins or protein fragments are, at least, partially unfolded under physiological conditions (96, 98) and all of them share a typical intramolecular cross-beta sheet conformation that leads to the formation of insoluble fibrillar structures (96). The mechanism of amyloid formation in protein misfolding disorders follows a process of seeded polymerization similar to one-dimensional crystal growth (96). Large fibrillar aggregates are formed by aggregates of a certain size or conformation, which stabilize and catalyze the addition of monomers (96, 99). Products of an alternative proteolytic processing of the amyloid precursor protein (APP) forms the extracellular protein deposits in AD (96), whereas intraneuronal deposits in the form of neurofibrillary tangles are formed by the microtubule-associated highly phosphorylated protein tau (96, 100-102). The other frequent protein misfolding disease, PD, is characterized by deposits

located in dopaminergic neurons of the substantia nigra, which primarily contain the alpha-synuclein protein (α S) but have also tau and A β as secondary components (96, 103).

Polyphenolic extract has been shown to decrease the formation of amyloid deposits (104, 105) and to significantly attenuate cognitive deterioration in a mouse model of AD (106) through its antioxidant activity (107) and the promotion of APP degradation into non-amyloidogenic peptides (108-110). It has also been reported that EGCG prevents the fibrillization of several proteins that are implicated in protein misfolding disorders by binding directly to them (96, 111). Moreover, it promotes the formation of spherical, stable, not cytotoxic aggregates, which have a lower β -sheet content than fibrils, and do not catalyze fibril formation (96). Derivatives of orcein have been reported not only to accelerate fibril formation of the AD-related A β peptide but also to deplete oligomeric and protofibrillar forms of the peptide (96). The phenylethanoid oleuropein aglycone (OLE), which arises from deglycosylation of oleuropein, found in the leaves and drupes of *Olea europaea*, was able to abolish the formation of toxic oligomers during the *in vitro* amyloid aggregation of the A β peptide, as well as promote fibril and plaque disaggregation, in various model organisms with subsequent relief of AD-like symptoms (112-115). OLE is also active against tau and human islet amyloid polypeptide (hIAPP) aggregation *in vitro* (116, 117). Finally, RSV derivatives significantly inhibited A β aggregation, disaggregated fibrils generated by self- and Cu(II)-induced A β aggregation and displayed antioxidant effect (118).

Neurotrophic Activity

Neurotrophins, including nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin 3 (NT3) and neurotrophin 4 (NT4), play essential roles in development, maintenance, repair and survival of particular neuronal populations (119), whereas the diminished function of neurotrophins and their receptors can result in neuronal damage, contributing in that way to the pathogenesis of neurodegenerative disorders (120).

Neurotrophins interact with two receptor types: neurotrophin receptor p75 (p75NTR) and the tropomyosin receptor kinase (Trk) receptors (TrkA, B and C) (121). The Trk receptors are dimerized upon neurotrophins binding and are activated by transphosphorylation of the cytoplasmic domain kinases, an event leading to the stimulation of the downstream signaling effectors MAPK, phosphatidylinositol-3-kinase (PI3K)/Akt and phospholipase C- γ 1 (121, 122).

Several studies have shown that various polyphenols, including flavonoids and acetylated flavonoid glycosides from *Scoparia dulcis*, are capable of enhancing the activity of NGF (123), whereas the stilbenoid compound RSV

produced neurotrophic effects on cultured dopamine neurons through promoting neurotrophic factors release (124).

A few polyphenolic compounds act as Trk receptors agonists, whereas many others activate downstream effectors, leading to a neurotrophic effect (120). 7,8-Dihydroxyflavone (7,8-DHF) triggers TrkB dimerization and tyrosine phosphorylation and stimulates downstream effectors (125). The flavonoid epicatechin has been reported to restore TrkA phosphorylation in diabetic animal models and to reduce diabetes-related neuronal cell death (120, 126). The diterpenes, kansuinin A, D, E, isolated from the roots of *Euphorbia kansui*, promoted the survival of TrkA- or TrkA- and B-expressing fibroblasts (127). The quinic acid metabolite (-)-3,5-dicaffeoyl-muco-quinic acid has been shown to enhance neurite outgrowth in PC12 cells through activation of signaling pathways similar to NGF, thus suggesting that this secondary metabolite specifically activates TrkA (128). Interestingly, a compound isolated from the fungus *Penicillium fellutanum*, fellutamide B, has been reported to possess unique neurotrophic activity through inhibition of the proteasome and increase in the production and secretion of NGF (129, 130). Many polyphenolic compounds, including flavonoids, have also been shown to increase the expression of NGF, glial cell line-derived neurotrophic factor (GDNF), BDNF, TrkA or TrkB, in various *in vivo* animal models (120) via activation of ERK-CREB-BDNF pathway, Nrf2/ARE signaling pathway or Akt/ Glycogen synthase kinase 3 β (GSK-3 β) signaling pathway (reviewed by Moosavi *et al.* (120)). Finally, the neuroprotective potency of natural compounds could be a consequence of less studied mechanisms, such as modulation of action of Na⁺/K⁺/2Cl⁻ co-transporter (NKCC1) or A2A receptor (131).

Neuronal Survival Signaling Pathways

Modulation of neuronal survival signal transduction pathways may be an attractive approach to the treatment of CNS diseases.

Wnt signaling contributes to normal neural development, neuronal homeostasis, axonogenesis, synaptic plasticity and the establishment of brain polarity (43, 132). GSK-3 β inhibition by CUR has been demonstrated to lead to Wnt/ β -catenin signaling activation and subsequent reduced A β production (133-135).

ERK pathway, a part of MAPKs, has been involved in many neuronal functions, such as proliferation, differentiation, survival and regulation of neuronal response to various growth factors (120). ERK1/2 activation by several polyphenolic compounds like luteolin promoted survival in various cell lines, enhanced neurite outgrowth and neuronal differentiation (136, 137). Moreover, liquiritin, icaritin and rutin have shown favorable effects against A β -

induced neurotoxicity *in vivo* through activation of MAPK and BDNF (120).

A number of studies have also reported that PI3K and its downstream effector Akt may be responsible for neuronal survival, increased neurite outgrowth, as well as other polyphenols-mediated neurotrophic actions (120). The PI3K/Akt pathway, for instance, has been implicated in the neuroprotective effect of CUR in a rat model of A β -induced cognitive impairment (138). Puerarin similarly, *via* stimulation of the ERK1/2 and PI3K/Akt pathways, potentiated NGF-induced neuritogenesis in PC12 cells and protected dopaminergic cells (139). Finally, activation of the MAPK/ERK and PI3K/Akt pathways, both downstream signaling effectors of BDNF, have been shown to account for the astilbin-induced improvement of depressive-like behaviors in mice models of depression (120).

CREB is a transcription regulator that recognizes Cyclic AMP response element (CRE) sequence and, once activated, up-regulates the expression of genes responsible for survival, growth, synaptic plasticity, dendritic spine formation, differentiation and long-term memory (120, 140). CUR, nobiletin and hesperetin have all been shown to induce CREB phosphorylation in PC12 cells, while green tea catechins and several flavonoids have been shown to activate CREB in various *in vivo* neurodegenerative models (120).

Autophagy

Autophagy is a multistep process implicating the formation of the autophagosomes, double-membrane structures, which fuse with lysosomes, (141) and their content (cellular metabolic waste, misfolded proteins) is degraded by hydrolytic enzymes. Autophagy is also crucial for mitochondrial turnover under physiological conditions, in a process known as mitophagy (142). Normal basal autophagy plays a fundamental role in the integrity of the CNS (141, 143). Therefore, stimulation of autophagy in the CNS would lead to neuroprotection and that could be the case for the beneficial effects of various compounds, including RSV (144). QCT-induced autophagy has been shown to alleviate Schwann cells' damage caused by high glucose (145). OLE was also found to induce autophagy and to reduce inflammation in a mouse model of A β deposition (115). Furthermore, RSV was shown recently to affect mitochondrial fission and mitophagy as well (24).

Epigenetics

Epigenetic mechanisms, such as DNA methylation, genomic imprinting, histone modification and regulation by microRNA, are important for normal development and maintenance of adult life and their dysregulation may also contribute to the susceptibility and complexity of

neurodegenerative diseases (146). Indeed, DNA methylation is involved in AD-related molecular mechanisms (147) (reviewed by Mastroeni *et al.* (148)).

Natural compounds, including polyphenols, alkaloids, terpenoids and organosulfur compounds, may play an essential role in modulating these epigenetic modifications (6, 28) and several of them have been reported to be able to alter DNA methylation and histone modifications leading to gene activation or silencing (6). Many polyphenols display their neuroprotective effects through activation of HDACs, whereas polyphenols with anti-inflammatory properties, such as catechins, RSV and CUR, were reported to modify NF-kappaB expression and chromatin remodeling through modulation of both HDACs and DNMTs activities (6).

Other Targets

It has been shown that AD is related to a reduction of cholinergic neurons' activity (149). Galantamine, an alkaloid used for the symptomatic treatment of AD, shows neuroprotective effect through activation of muscarinic acetylcholine receptors (150). Finally, flavonoids, as well as other natural compounds, have been reported to inhibit monoamine oxidases (MAO) (151), mitochondrial bound enzymes, which have been implicated in various processes leading to neurodegeneration, such as oxidative stress, neuroinflammation, triggering of apoptosis and glial activation (152).

Bioavailability Issues and Development of Novel Delivery Systems

Natural compounds usually target multiple signaling pathways and regulate gene expression broadly leading to a wide-ranging spectrum of activities, such as anti-inflammatory and antioxidant. These multi-targeting properties of natural compounds make them attractive candidates for the treatment of disorders, where a multitude of pathophysiological pathways is affected, like neurodegenerative diseases (7-9).

However, their physicochemical properties are not drug-like. Several natural compounds show limited stability as they are sensitive to degradation or are metabolized to inactive derivatives in circulation (153, 154). Indeed, tea catechins undergo extensive methylation, glucuronidation and sulfation (155). Also, RSV and CUR have low bioavailability due to their rapid metabolism and elimination (156, 157). Compound solubility might be an additional issue, since most of them show limited solubility in water (153). Moreover, restricted passage across the blood brain barrier (BBB) (158) and subsequent limited distribution to brain tissue contribute to poor bioavailability (153). But even when the compound reaches the brain and passes the BBB, it will be diluted and cleared in cerebrospinal fluid before

distributed into the brain parenchyma (159). Therefore, the properties of the compound delivery system are crucial for efficient biodistribution (159). Finally, relatively high local compound concentrations are required for a desirable effect because of the modest potency of natural therapeutic agents.

Given the rapid progression in the field of nanomedicine, research has now concentrated on shifting from microsystems to nanocarriers for the treatment of neurodegenerative disorders. The incorporation of nanoparticles with the specific formulation and physicochemical properties aims to improve the bioavailability, as well as the targeting and the controlled-release profiles of natural products (160).

In order to overcome solubility issues of natural compounds for *in vivo* application, the liposomal formulation has been applied as a broadly used strategy (153). Encapsulating highly lipophilic compounds, such as EGCG, RSV and CUR, which dissolve poorly in the bloodstream, can increase their water solubility and efficiency. It has been reported, that EGCG nanolipids' oral bioavailability was higher than free EGCG (161) and lipid core nanocapsules loaded with RSV increased RSV concentration in brain tissue, compared to free RSV. Several drug delivery systems have been tested for improved targeting of CUR, such as liposomes, solid lipid nanoparticles (SLNs), polymeric nanoparticles (poly(lactic-co-glycolic acid -PLGA), nanogels, micelles and complexes with dendrimer/dimer (43). SLN of CUR showed a great recovery in membrane lipids, as well as acetylcholinesterase (AChE) activity, in aluminium chloride-treated mice, an effect that was comparable to rivastigmine (43, 162). Recently, SLN-encapsulated QCT showed a better neuroprotective effect, indicating improved penetration *via* BBB (163). According to another study, QCT-loaded β -cyclodextrin dodecylcarbonate nanoparticles decreased inflammatory mediators in a neuronal cell line (164). Incorporating the flavone apigenin into a carbon nanopowder, solid distribution improved its low lipid and water solubility, whereas self-nanoemulsifying QCT also showed increased bioavailability, compared to free QCT. (165, 166).

Polymeric nanoparticles have been broadly tested in combination with natural products and polymers, such as poly(vinyl alcohol (PVA), polyethylene glycol (PEG) and polylactide (PLA), have been utilized to improve the bioavailability of EGCG, luteolin, silibinin and tea polyphenols. (166). Berberine-loaded nanoemulsion formulation also increased its bioavailability (167). Finally, polysaccharides, which have recently been demonstrated to form amorphous solid dispersions in nanoparticle form, significantly enhanced the solubility and bioavailability of natural compounds, including CUR, ellagic acid, naringenin, QCT and RSV(168-170).

The route of delivery is also a great effector of a compound's bioavailability. Oral delivery is beneficial because of high patient compliance. However, systemic drug

delivery relies on drugs to be targeted to brain tissue by employing cell-penetrating peptides or attaching targeting ligands to the nanocarriers' surface. The nasal pathway *via* the olfactory mucosa has also been explored for the delivery of macromolecules, small molecule drugs, enzymes and genetic material (171, 172).

The application of nanoparticles for natural compounds delivery offers further the opportunity to target specifically the desired tissues or organs. In that way, drug bioavailability is improved and toxic side effects are reduced. Targeting approaches include either attachment of a targeting ligand to the nanoparticle surface (active targeting) or application of nanoparticles without specific chemical modifications, whose physical transport and targeting relies upon their intrinsic properties, such as size, shape and surface charge (passive targeting) (166, 173). For example, one strategy for targeting the brain is to manipulate the lipophilicity of the nanoparticles, since the BBB favors crossing over of lipophilic molecules. Monoclonal antibodies conjugated onto nanoparticles are promising candidates for targeting the BBB, although there are no data regarding their use with natural compounds. (166, 174). A known amyloid-binding compound, benzothiazole aniline (BTA), has been reported to inhibit beta-amyloid protein aggregations (175). A BTA-based nanoparticle could potentially be modified to improve the delivery of neuroprotective natural compounds for the treatment of AD (166). Targeting can also be achieved using external forces, like magnetic fields. Magnetic-guided nanoparticles can be employed during magnetic resonance imaging (MRI) for targeted drug or gene delivery, tissue engineering applications and cell tracking within the brain tissue (176). Finally focused ultrasound (FU), as an approach to induce BBB permeability, has been analyzed, employing two-photon microscopy that resulted in temporary, focused alterations of BBB (177).

Application of nanoparticles for delivering natural compounds could also enable a controlled release of the drug. A variety of factors, such as the size and the type of the particle, the type and amount of the encapsulated natural compound and the microenvironment, define the amount and rate at which a compound is released from a nanoformulation (166, 178). Compound release from nanoparticles, for instance, can be triggered using environmental changes, such as ultrasound and light (179, 180). Nanoparticles, made with a new, based on the quinonemethide system, light-sensitive polymer, released an encapsulated drug after exposure to a particular light wavelength(s) (179). This approach could be beneficial for the treatment of ocular degenerative disorders due to the transparency of the optical media.

However, optimal application of neuroprotective agents still presents a pivotal challenge for treating neurodegenerative diseases. First, degenerative diseases are characterized by a neuronal compensated dysfunction for a prolonged period of time prior to cell loss that is amenable

to therapeutic intervention (181). Then, innovative strategies are needed to optimally deliver a drug to specific regions within the brain (181, 182). Finally, several issues arise from the development of these new formulations for the delivery of natural compounds. For example, higher BBB permeability conditions, which result in increased amount of the compound reaching the CNS parenchyma, may elevate the risk of neurotoxicity (94). Increased concentration, for instance, of a single antioxidant polyphenol might be harmful to human health (183). Furthermore, there are reports regarding discrepancies of a compound's effect on particular signaling pathways among different experimental settings, such as anti- or pro-oxidant function and stimulatory or inhibitory role on specific kinases activity (24, 86). This is probably due to the particular physiology of the different cells tested, as well as other parameters, such as concentration and incubation time with the compound. Further studies are also needed to investigate how targeted, but also not targeted, cells will be affected by the rise in compound availability due to nanotechnology-related strategies. Therefore, it is becoming clear that availability must be tightly controlled. In addition, the lack of knowledge concerning the interactions of nanomaterials with biological membranes, as well as the fate of the nanoparticles after they are administrated and distributed *in vivo* (184), the scarcity of adequate *in vitro* models to replicate the BBB (181) and the lack of technological platforms to screen large quantities of nanoparticles (166), are some of the issues that need to be elucidated before clinical application takes place.

The design and the development of novel compound formulation and delivery technologies based on nanomaterials in combination with a deep understanding of the targeted biological systems seems to be a favorable approach, in order for natural therapeutics agents to be used in the clinical practice against neurodegenerative diseases.

References

- Ray K and Mookherjee S: Molecular complexity of primary open angle glaucoma: current concepts. *J Genet* 88: 451-467, 2009.
- Andersen JK: Oxidative stress in neurodegeneration: cause or consequence? *Nat Med* 10 *Suppl*: S18-25, 2004.
- Zadori D, Klivenyi P, Szalardy L, Fulop F, Toldi J and Vecsei L: Mitochondrial disturbances, excitotoxicity, neuro-inflammation and kynurenines: novel therapeutic strategies for neurodegenerative disorders. *J Neurol Sci* 322: 187-191, 2012.
- Harvey AL and Cree IA: High-throughput screening of natural products for cancer therapy. *Planta Med* 76: 1080-1086, 2010.
- Bagli E, Stefaniotou M, Morbidelli L, Ziche M, Psillas K, Murphy C and Fotsis T: Luteolin inhibits vascular endothelial growth factor-induced angiogenesis; inhibition of endothelial cell survival and proliferation by targeting phosphatidylinositol 3'-kinase activity. *Cancer Res* 64: 7936-7946, 2004.
- Rahman I and Chung S: Dietary polyphenols, deacetylases and chromatin remodeling in inflammation. *J Nutrigenet Nutrigenomics* 3: 220-230, 2010.
- Balunas MJ and Kinghorn AD: Drug discovery from medicinal plants. *Life Sci* 78: 431-441, 2005.
- Harvey AL, Clark RL, Mackay SP and Johnston BF: Current strategies for drug discovery through natural products. *Expert Opin Drug Discov* 5: 559-568, 2010.
- Kimura I: Medical benefits of using natural compounds and their derivatives having multiple pharmacological actions. *Yakugaku Zasshi* 126: 133-143, 2006.
- Beking K and Vieira A: Flavonoid intake and disability-adjusted life years due to Alzheimer's and related dementias: a population-based study involving twenty-three developed countries. *Public Health Nutr* 13: 1403-1409, 2010.
- Sood PK, Nahar U and Nehru B: Curcumin attenuates aluminum-induced oxidative stress and mitochondrial dysfunction in rat brain. *Neurotox Res* 20: 351-361, 2011.
- Andreux PA, Houtkooper RH and Auwerx J: Pharmacological approaches to restore mitochondrial function. *Nat Rev Drug Discov* 12: 465-483, 2013.
- Picone P, Nuzzo D, Caruana L, Scafidi V and Di Carlo M: Mitochondrial dysfunction: different routes to Alzheimer's disease therapy. *Oxid Med Cell Longev* 2014: 780179, 2014.
- Waseem M and Parvez S: Neuroprotective activities of curcumin and quercetin with potential relevance to mitochondrial dysfunction induced by oxaliplatin. *Protoplasma* 253: 417-430, 2016.
- Kim D, Nguyen MD, Dobbin MM, Fischer A, Sananbenesi F, Rodgers JT, Delalle I, Baur JA, Sui G, Armour SM, Puigserver P, Sinclair DA and Tsai LH: SIRT1 deacetylase protects against neurodegeneration in models for Alzheimer's disease and amyotrophic lateral sclerosis. *EMBO J* 26: 3169-3179, 2007.
- Canto C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, Elliott PJ, Puigserver P and Auwerx J: AMPK regulates energy expenditure by modulating NAD⁺ metabolism and SIRT1 activity. *Nature* 458: 1056-1060, 2009.
- Scarpulla RC: Metabolic control of mitochondrial biogenesis through the PGC-1 family regulatory network. *Biochim Biophys Acta* 1813: 1269-1278, 2011.
- Fiorani M, Guidarelli A, Blasa M, Azzolini C, Candiracci M, Piatti E and Cantoni O: Mitochondria accumulate large amounts of quercetin: prevention of mitochondrial damage and release upon oxidation of the extramitochondrial fraction of the flavonoid. *J Nutr Biochem* 21: 397-404, 2010.
- Davinelli S, Sapere N, Visentin M, Zella D and Scapagnini G: Enhancement of mitochondrial biogenesis with polyphenols: combined effects of resveratrol and equol in human endothelial cells. *Immun Ageing* 10: 28, 2013.
- de Oliveira MR, Nabavi SF, Habtemariam S, Erdogan Orhan I, Daglia M and Nabavi SM: The effects of baicalein and baicalin on mitochondrial function and dynamics: A review. *Pharmacol Res* 100: 296-308, 2015.
- Cai Z, Zeng W, Tao K, Lu F, Gao G and Yang Q: Myricitrin alleviates MPP(+)-induced mitochondrial dysfunction in a DJ-1-dependent manner in SN4741 cells. *Biochem Biophys Res Commun* 458: 227-233, 2015.
- Wang B, Sun J, Ma Y, Wu G, Tian Y, Shi Y and Le G: Resveratrol preserves mitochondrial function, stimulates

- mitochondrial biogenesis, and attenuates oxidative stress in regulatory T cells of mice fed a high-fat diet. *J Food Sci* 79: H1823-1831, 2014.
- 23 Zhang Z, Zheng L, Zhao Z, Shi J, Wang X and Huang J: Grape seed proanthocyanidins inhibit H₂O₂-induced osteoblastic MC3T3-E1 cell apoptosis *via* ameliorating H₂O₂-induced mitochondrial dysfunction. *J Toxicol Sci* 39: 803-813, 2014.
- 24 de Oliveira MR, Nabavi SF, Manayi A, Daglia M, Hajheydari Z and Nabavi SM: Resveratrol and the mitochondria: From triggering the intrinsic apoptotic pathway to inducing mitochondrial biogenesis, a mechanistic view. *Biochim Biophys Acta* 1860: 727-745, 2016.
- 25 Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, Messadeq N, Milne J, Lambert P, Elliott P, Geny B, Laakso M, Puigserver P and Auwerx J: Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1 α . *Cell* 127: 1109-1122, 2006.
- 26 Zhou X, Chen M, Zeng X, Yang J, Deng H, Yi L and Mi MT: Resveratrol regulates mitochondrial reactive oxygen species homeostasis through Sirt3 signaling pathway in human vascular endothelial cells. *Cell Death Dis* 5: e1576, 2014.
- 27 Li YG, Zhu W, Tao JP, Xin P, Liu MY, Li JB and Wei M: Resveratrol protects cardiomyocytes from oxidative stress through SIRT1 and mitochondrial biogenesis signaling pathways. *Biochem Biophys Res Commun* 438: 270-276, 2013.
- 28 Ayissi VB, Ebrahimi A and Schluesener H: Epigenetic effects of natural polyphenols: a focus on SIRT1-mediated mechanisms. *Mol Nutr Food Res* 58: 22-32, 2014.
- 29 Cao K, Zheng A, Xu J, Li H, Liu J, Peng Y, Long J, Zou X, Li Y, Chen C, Liu J and Feng Z: AMPK activation prevents prenatal stress-induced cognitive impairment: modulation of mitochondrial content and oxidative stress. *Free Radic Biol Med* 75: 156-166, 2014.
- 30 Um JH, Park SJ, Kang H, Yang S, Foretz M, McBurney MW, Kim MK, Viollet B and Chung JH: AMP-activated protein kinase-deficient mice are resistant to the metabolic effects of resveratrol. *Diabetes* 59: 554-563, 2010.
- 31 Valenti D, de Bari L, de Rasmio D, Signorile A, Henrion-Caude A, Contestabile A and Vacca RA: The polyphenols resveratrol and epigallocatechin-3-gallate restore the severe impairment of mitochondria in hippocampal progenitor cells from a Down syndrome mouse model. *Biochim Biophys Acta* 2016.
- 32 Fu J, Jin J, Cichewicz RH, Hageman SA, Ellis TK, Xiang L, Peng Q, Jiang M, Arbez N, Hotaling K, Ross CA and Duan W: trans-(\pm)-epsilon-Viniferin increases mitochondrial sirtuin 3 (SIRT3), activates AMP-activated protein kinase (AMPK), and protects cells in models of Huntington Disease. *J Biol Chem* 287: 24460-24472, 2012.
- 33 Haleagrahara N, Siew CJ and Ponnusamy K: Effect of quercetin and desferrioxamine on 6-hydroxydopamine (6-OHDA) induced neurotoxicity in striatum of rats. *J Toxicol Sci* 38: 25-33, 2013.
- 34 Wu CH and Yen GC: Inhibitory effect of naturally occurring flavonoids on the formation of advanced glycation endproducts. *J Agric Food Chem* 53: 3167-3173, 2005.
- 35 Cervantes-Laurean D, Schramm DD, Jacobson EL, Halaweish I, Bruckner GG and Boissonneault GA: Inhibition of advanced glycation end product formation on collagen by rutin and its metabolites. *J Nutr Biochem* 17: 531-540, 2006.
- 36 Wang YH, Yu HT, Pu XP and Du GH: Myricitrin alleviates methylglyoxal-induced mitochondrial dysfunction and AGEs/RAGE/NF-kappaB pathway activation in SH-SY5Y cells. *J Mol Neurosci* 53: 562-570, 2014.
- 37 Shimohama S: Apoptosis in Alzheimer's disease--an update. *Apoptosis* 5: 9-16, 2000.
- 38 Agnantis NJ and Goussia AC: [Apoptosis and cancer]. *Bull Acad Natl Med* 183: 277-286; discussion 286-277, 1999.
- 39 Zhao H, Yenari MA, Cheng D, Sapolsky RM and Steinberg GK: Bcl-2 overexpression protects against neuron loss within the ischemic margin following experimental stroke and inhibits cytochrome c translocation and caspase-3 activity. *J Neurochem* 85: 1026-1036, 2003.
- 40 Donovan M and Cotter TG: Control of mitochondrial integrity by Bcl-2 family members and caspase-independent cell death. *Biochim Biophys Acta* 1644: 133-147, 2004.
- 41 Oyadomari S and Mori M: Roles of CHOP/GADD153 in endoplasmic reticulum stress. *Cell Death Differ* 11: 381-389, 2004.
- 42 Farimani MM, Sarvestani NN, Ansari N and Khodagholi F: Calycopterin promotes survival and outgrowth of neuron-like PC12 cells by attenuation of oxidative- and ER-stress-induced apoptosis along with inflammatory response. *Chem Res Toxicol* 24: 2280-2292, 2011.
- 43 Ansari N and Khodagholi F: Natural products as promising drug candidates for the treatment of Alzheimer's disease: molecular mechanism aspect. *Curr Neuropharmacol* 11: 414-429, 2013.
- 44 Defeudis FV: Bilobalide and neuroprotection. *Pharmacol Res* 46: 565-568, 2002.
- 45 Zhou LJ and Zhu XZ: Reactive oxygen species-induced apoptosis in PC12 cells and protective effect of bilobalide. *J Pharmacol Exp Ther* 293: 982-988, 2000.
- 46 Shi C, Zou J, Li G, Ge Z, Yao Z and Xu J: Bilobalide protects mitochondrial function in ovariectomized rats by up-regulation of mRNA and protein expression of cytochrome c oxidase subunit I. *J Mol Neurosci* 45: 69-75, 2011.
- 47 Zhang HY and Tang XC: Neuroprotective effects of huperzine A: new therapeutic targets for neurodegenerative disease. *Trends Pharmacol Sci* 27: 619-625, 2006.
- 48 Thoreson WB and Witkovsky P: Glutamate receptors and circuits in the vertebrate retina. *Prog Retin Eye Res* 18: 765-810, 1999.
- 49 Pin JP and Duvoisin R: The metabotropic glutamate receptors: structure and functions. *Neuropharmacology* 34: 1-26, 1995.
- 50 Crompton M, Virji S, Doyle V, Johnson N and Ward JM: The mitochondrial permeability transition pore. *Biochem Soc Symp* 66: 167-179, 1999.
- 51 Kroemer G, Galluzzi L and Brenner C: Mitochondrial membrane permeabilization in cell death. *Physiol Rev* 87: 99-163, 2007.
- 52 Crompton M: The mitochondrial permeability transition pore and its role in cell death. *Biochem J* 341(Pt 2): 233-249, 1999.
- 53 Zeevalk GD and Nicklas WJ: Evidence that the loss of the voltage-dependent Mg²⁺ block at the N-methyl-D-aspartate receptor underlies receptor activation during inhibition of neuronal metabolism. *J Neurochem* 59: 1211-1220, 1992.
- 54 Mosinger JL, Price MT, Bai HY, Xiao H, Wozniak DF and Olney JW: Blockade of both NMDA and non-NMDA receptors is required for optimal protection against ischemic neuronal degeneration in the *in vivo* adult mammalian retina. *Exp Neurol* 113: 10-17, 1991.

- 55 Ahmed AH, Hamada M, Shinada T, Ohfuné Y, Weerasinghe L, Garner PP and Oswald RE: The structure of (–)-kaiotocephalin bound to the ligand binding domain of the (S)-alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA)/glutamate receptor, GluA2. *J Biol Chem* 287: 41007-41013, 2012.
- 56 Shimmyo Y, Kihara T, Akaike A, Niidome T and Sugimoto H: Three distinct neuroprotective functions of myricetin against glutamate-induced neuronal cell death: involvement of direct inhibition of caspase-3. *J Neurosci Res* 86: 1836-1845, 2008.
- 57 Lin TY, Huang WJ, Wu CC, Lu CW and Wang SJ: Acacetin inhibits glutamate release and prevents kainic acid-induced neurotoxicity in rats. *PLoS One* 9: e88644, 2014.
- 58 Amor S, Puentes F, Baker D and van der Valk P: Inflammation in neurodegenerative diseases. *Immunology* 129: 154-169, 2010.
- 59 Dheen ST, Kaur C and Ling EA: Microglial activation and its implications in the brain diseases. *Curr Med Chem* 14: 1189-1197, 2007.
- 60 Brown GC: Mechanisms of inflammatory neurodegeneration: iNOS and NADPH oxidase. *Biochem Soc Trans* 35: 1119-1121, 2007.
- 61 Eikelenboom P, Bate C, Van Gool WA, Hoozemans JJ, Rozemuller JM, Veerhuis R and Williams A: Neuroinflammation in Alzheimer's disease and prion disease. *Glia* 40: 232-239, 2002.
- 62 Rothwarf DM and Karin M: The NF-kappa B activation pathway: a paradigm in information transfer from membrane to nucleus. *Sci STKE* 1999: RE1, 1999.
- 63 Camandola S and Mattson MP: NF-kappa B as a therapeutic target in neurodegenerative diseases. *Expert Opin Ther Targets* 11: 123-132, 2007.
- 64 Qin ZH, Tao LY and Chen X: Dual roles of NF-kappaB in cell survival and implications of NF-kappaB inhibitors in neuroprotective therapy. *Acta Pharmacol Sin* 28: 1859-1872, 2007.
- 65 Sarnico I, Lanzillotta A, Benarese M, Alghisi M, Baiguera C, Battistin L, Spano P and Pizzi M: NF-kappaB dimers in the regulation of neuronal survival. *Int Rev Neurobiol* 85: 351-362, 2009.
- 66 Pizzi M, Goffi F, Boroni F, Benarese M, Perkins SE, Liou HC and Spano P: Opposing roles for NF-kappa B/Rel factors p65 and c-Rel in the modulation of neuron survival elicited by glutamate and interleukin-1beta. *J Biol Chem* 277: 20717-20723, 2002.
- 67 Gilgun-Sherki Y, Melamed E and Offen D: Anti-inflammatory drugs in the treatment of neurodegenerative diseases: current state. *Curr Pharm Des* 12: 3509-3519, 2006.
- 68 Kim HP, Son KH, Chang HW and Kang SS: Anti-inflammatory plant flavonoids and cellular action mechanisms. *J Pharmacol Sci* 96: 229-245, 2004.
- 69 Kotanidou A, Xagorari A, Bagli E, Kitsanta P, Fotsis T, Papapetropoulos A and Roussos C: Luteolin reduces lipopolysaccharide-induced lethal toxicity and expression of proinflammatory molecules in mice. *Am J Respir Crit Care Med* 165: 818-823, 2002.
- 70 Choi DK, Koppula S and Suk K: Inhibitors of microglial neurotoxicity: focus on natural products. *Molecules* 16: 1021-1043, 2011.
- 71 Chinta SJ, Ganesan A, Reis-Rodrigues P, Lithgow GJ and Andersen JK: Anti-inflammatory role of the isoflavone diadzein in lipopolysaccharide-stimulated microglia: implications for Parkinson's disease. *Neurotox Res* 23: 145-153, 2013.
- 72 Kim JY, Kina T, Iwanaga Y, Noguchi H, Matsumura K and Hyon SH: Tea polyphenol inhibits allostimulation in mixed lymphocyte culture. *Cell Transplant* 16: 75-83, 2007.
- 73 Kang TH, Lee JH, Song CK, Han HD, Shin BC, Pai SI, Hung CF, Trimble C, Lim JS, Kim TW and Wu TC: Epigallocatechin-3-gallate enhances CD8+ T cell-mediated antitumor immunity induced by DNA vaccination. *Cancer Res* 67: 802-811, 2007.
- 74 Min K, Yoon WK, Kim SK and Kim BH: Immunosuppressive effect of silibinin in experimental autoimmune encephalomyelitis. *Arch Pharm Res* 30: 1265-1272, 2007.
- 75 Xu Z, Chen S, Li X, Luo G, Li L and Le W: Neuroprotective effects of (–)-epigallocatechin-3-gallate in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurochem Res* 31: 1263-1269, 2006.
- 76 Liang YC, Huang YT, Tsai SH, Lin-Shiau SY, Chen CF and Lin JK: Suppression of inducible cyclooxygenase and inducible nitric oxide synthase by apigenin and related flavonoids in mouse macrophages. *Carcinogenesis* 20: 1945-1952, 1999.
- 77 Wang HM, Zhao YX, Zhang S, Liu GD, Kang WY, Tang HD, Ding JQ and Chen SD: PPARgamma agonist curcumin reduces the amyloid-beta-stimulated inflammatory responses in primary astrocytes. *J Alzheimers Dis* 20: 1189-1199, 2010.
- 78 Sikora E, Scapagnini G and Barbagallo M: Curcumin, inflammation, ageing and age-related diseases. *Immun Ageing* 7: 1, 2010.
- 79 Geng Y, Zhang B and Lotz M: Protein tyrosine kinase activation is required for lipopolysaccharide induction of cytokines in human blood monocytes. *J Immunol* 151: 6692-6700, 1993.
- 80 Sharma V, Mishra M, Ghosh S, Tewari R, Basu A, Seth P and Sen E: Modulation of interleukin-1beta mediated inflammatory response in human astrocytes by flavonoids: implications in neuroprotection. *Brain Res Bull* 73: 55-63, 2007.
- 81 Kempuraj D, Madhappan B, Christodoulou S, Boucher W, Cao J, Papadopoulos N, Cetrulo CL and Theoharides TC: Flavonols inhibit proinflammatory mediator release, intracellular calcium ion levels and protein kinase C theta phosphorylation in human mast cells. *Br J Pharmacol* 145: 934-944, 2005.
- 82 Wadsworth TL, McDonald TL and Koop DR: Effects of Ginkgo biloba extract (EGb 761) and quercetin on lipopolysaccharide-induced signaling pathways involved in the release of tumor necrosis factor-alpha. *Biochem Pharmacol* 62: 963-974, 2001.
- 83 Albarracin SL, Stab B, Casas Z, Sutachan JJ, Samudio I, Gonzalez J, Gonzalo L, Capani F, Morales L and Barreto GE: Effects of natural antioxidants in neurodegenerative disease. *Nutr Neurosci* 15: 1-9, 2012.
- 84 Hwang O: Role of oxidative stress in Parkinson's disease. *Exp Neurobiol* 22: 11-17, 2013.
- 85 Halliwell B: Oxidative stress and neurodegeneration: where are we now? *J Neurochem* 97: 1634-1658, 2006.
- 86 Aquilano K, Baldelli S, Rotilio G and Ciriolo MR: Role of nitric oxide synthases in Parkinson's disease: a review on the antioxidant and anti-inflammatory activity of polyphenols. *Neurochem Res* 33: 2416-2426, 2008.
- 87 Yuste JE, Tarragon E, Campuzano CM and Ros-Bernal F: Implications of glial nitric oxide in neurodegenerative diseases. *Front Cell Neurosci* 9: 322, 2015.
- 88 Mosley RL, Benner EJ, Kadiu I, Thomas M, Boska MD, Hasan K, Laurie C and Gendelman HE: Neuroinflammation, Oxidative Stress and the Pathogenesis of Parkinson's Disease. *Clin Neurosci Res* 6: 261-281, 2006.

- 89 Park JH, Choi JW, Ju EJ, Pae AN and Park KD: Antioxidant and Anti-Inflammatory Activities of a Natural Compound, Shizukahenriol, through Nrf2 Activation. *Molecules* 20: 15989-16003, 2015.
- 90 de Vries HE, Witte M, Hondius D, Rozemuller AJ, Drukarch B, Hoozemans J and van Horsen J: Nrf2-induced antioxidant protection: a promising target to counteract ROS-mediated damage in neurodegenerative disease? *Free Radic Biol Med* 45: 1375-1383, 2008.
- 91 Sporn MB and Liby KT: NRF2 and cancer: the good, the bad and the importance of context. *Nat Rev Cancer* 12: 564-571, 2012.
- 92 Koh K, Kim J, Jang YJ, Yoon K, Cha Y, Lee HJ and Kim J: Transcription factor Nrf2 suppresses LPS-induced hyperactivation of BV-2 microglial cells. *J Neuroimmunol* 233: 160-167, 2011.
- 93 Lee IS, Lim J, Gal J, Kang JC, Kim HJ, Kang BY and Choi HJ: Anti-inflammatory activity of xanthohumol involves heme oxygenase-1 induction via NRF2-ARE signaling in microglial BV2 cells. *Neurochem Int* 58: 153-160, 2011.
- 94 Ossola B, Kaariainen TM and Mannisto PT: The multiple faces of quercetin in neuroprotection. *Expert Opin Drug Saf* 8: 397-409, 2009.
- 95 Fraunberger EA, Scola G, Laliberte VL, Duong A and Andrezza AC: Redox Modulations, Antioxidants, and Neuropsychiatric Disorders. *Oxid Med Cell Longev* 2016: 4729192, 2016.
- 96 Bieschke J: Natural compounds may open new routes to treatment of amyloid diseases. *Neurotherapeutics* 10: 429-439, 2013.
- 97 Dobson CM: Protein folding and misfolding. *Nature* 426: 884-890, 2003.
- 98 Uversky VN: Intrinsic disorder in proteins associated with neurodegenerative diseases. *Front Biosci (Landmark Ed)* 14: 5188-5238, 2009.
- 99 Powers ET and Powers DL: The kinetics of nucleated polymerizations at high concentrations: amyloid fibril formation near and above the "supercritical concentration". *Biophys J* 91: 122-132, 2006.
- 100 Taylor JP, Hardy J and Fischbeck KH: Toxic proteins in neurodegenerative disease. *Science* 296: 1991-1995, 2002.
- 101 Goedert M, Spillantini MG, Jakes R, Rutherford D and Crowther RA: Multiple isoforms of human microtubule-associated protein tau: sequences and localization in neurofibrillary tangles of Alzheimer's disease. *Neuron* 3: 519-526, 1989.
- 102 Vulliet R, Halloran SM, Braun RK, Smith AJ and Lee G: Proline-directed phosphorylation of human Tau protein. *J Biol Chem* 267: 22570-22574, 1992.
- 103 Olanow CW and Tatton WG: Etiology and pathogenesis of Parkinson's disease. *Annu Rev Neurosci* 22: 123-144, 1999.
- 104 Rezai-Zadeh K, Shytle D, Sun N, Mori T, Hou H, Jeanniton D, Ehrhart J, Townsend K, Zeng J, Morgan D, Hardy J, Town T and Tan J: Green tea epigallocatechin-3-gallate (EGCG) modulates amyloid precursor protein cleavage and reduces cerebral amyloidosis in Alzheimer transgenic mice. *J Neurosci* 25: 8807-8814, 2005.
- 105 Rezai-Zadeh K, Arendash GW, Hou H, Fernandez F, Jensen M, Runfeldt M, Shytle RD and Tan J: Green tea epigallocatechin-3-gallate (EGCG) reduces beta-amyloid mediated cognitive impairment and modulates tau pathology in Alzheimer transgenic mice. *Brain Res* 1214: 177-187, 2008.
- 106 Wang J, Ho L, Zhao W, Ono K, Rosensweig C, Chen L, Humala N, Teplow DB and Pasinetti GM: Grape-derived polyphenolics prevent Abeta oligomerization and attenuate cognitive deterioration in a mouse model of Alzheimer's disease. *J Neurosci* 28: 6388-6392, 2008.
- 107 Mandel SA, Amit T, Kalfon L, Reznichenko L, Weinreb O and Youdim MB: Cell signaling pathways and iron chelation in the neurorestorative activity of green tea polyphenols: special reference to epigallocatechin gallate (EGCG). *J Alzheimers Dis* 15: 211-222, 2008.
- 108 Fernandez JW, Rezai-Zadeh K, Obregon D and Tan J: EGCG functions through estrogen receptor-mediated activation of ADAM10 in the promotion of non-amyloidogenic processing of APP. *FEBS Lett* 584: 4259-4267, 2010.
- 109 Lee JW, Lee YK, Ban JO, Ha TY, Yun YP, Han SB, Oh KW and Hong JT: Green tea (-)-epigallocatechin-3-gallate inhibits beta-amyloid-induced cognitive dysfunction through modification of secretase activity via inhibition of ERK and NF-kappaB pathways in mice. *J Nutr* 139: 1987-1993, 2009.
- 110 Jeon SY, Bae K, Seong YH and Song KS: Green tea catechins as a BACE1 (beta-secretase) inhibitor. *Bioorg Med Chem Lett* 13: 3905-3908, 2003.
- 111 Ehrnhoefer DE, Bieschke J, Boeddrich A, Herbst M, Masino L, Lurz R, Engemann S, Pastore A and Wanker EE: EGCG redirects amyloidogenic polypeptides into unstructured, off-pathway oligomers. *Nat Struct Mol Biol* 15: 558-566, 2008.
- 112 Rigacci S, Guidotti V, Bucciantini M, Nichino D, Relini A, Berti A and Stefani M: Abeta(1-42) aggregates into non-toxic amyloid assemblies in the presence of the natural polyphenol oleuropein aglycon. *Curr Alzheimer Res* 8: 841-852, 2011.
- 113 Diomede L, Rigacci S, Romeo M, Stefani M and Salmona M: Oleuropein aglycone protects transgenic *C. elegans* strains expressing Abeta42 by reducing plaque load and motor deficit. *PLoS One* 8: e58893, 2013.
- 114 Luccarini I, Ed Dami T, Grossi C, Rigacci S, Stefani M and Casamenti F: Oleuropein aglycone counteracts Abeta42 toxicity in the rat brain. *Neurosci Lett* 558: 67-72, 2014.
- 115 Grossi C, Rigacci S, Ambrosini S, Ed Dami T, Luccarini I, Traini C, Failli P, Berti A, Casamenti F and Stefani M: The polyphenol oleuropein aglycone protects TgCRND8 mice against Ass plaque pathology. *PLoS One* 8: e71702, 2013.
- 116 Daccache A, Lion C, Sibille N, Gerard M, Slomianny C, Lippens G and Cotelle P: Oleuropein and derivatives from olives as Tau aggregation inhibitors. *Neurochem Int* 58: 700-707, 2011.
- 117 Rigacci S, Guidotti V, Bucciantini M, Parri M, Nediani C, Cerbai E, Stefani M and Berti A: Oleuropein aglycon prevents cytotoxic amyloid aggregation of human amylin. *J Nutr Biochem* 21: 726-735, 2010.
- 118 Lu C, Guo Y, Yan J, Luo Z, Luo HB, Yan M, Huang L and Li X: Design, synthesis, and evaluation of multitarget-directed resveratrol derivatives for the treatment of Alzheimer's disease. *J Med Chem* 56: 5843-5859, 2013.
- 119 Levy YS, Gilgun-Sherki Y, Melamed E and Offen D: Therapeutic potential of neurotrophic factors in neurodegenerative diseases. *BioDrugs* 19: 97-127, 2005.
- 120 Moosavi F, Hosseini R, Saso L and Firuzi O: Modulation of neurotrophic signaling pathways by polyphenols. *Drug Des Devel Ther* 10: 23-42, 2016.
- 121 Huang EJ and Reichardt LF: Trk receptors: roles in neuronal signal transduction. *Annu Rev Biochem* 72: 609-642, 2003.
- 122 Kaplan DR and Miller FD: Neurotrophin signal transduction in the nervous system. *Curr Opin Neurobiol* 10: 381-391, 2000.

- 123 Li Y, Chen X, Satake M, Oshima Y and Ohizumi Y: Acetylated flavonoid glycosides potentiating NGF action from *Scoparia dulcis*. *J Nat Prod* 67: 725-727, 2004.
- 124 Zhang F, Wang YY, Liu H, Lu YF, Wu Q, Liu J and Shi JS: Resveratrol Produces Neurotrophic Effects on Cultured Dopaminergic Neurons through Prompting Astroglial BDNF and GDNF Release. *Evid Based Complement Alternat Med* 2012: 937605, 2012.
- 125 Gupta VK, You Y, Li JC, Klistorner A and Graham SL: Protective effects of 7,8-dihydroxyflavone on retinal ganglion and RGC-5 cells against excitotoxic and oxidative stress. *J Mol Neurosci* 49: 96-104, 2013.
- 126 Al-Gayyar MM, Matragoon S, Pillai BA, Ali TK, Abdelsaid MA and El-Remessy AB: Epicatechin blocks pro-nerve growth factor (proNGF)-mediated retinal neurodegeneration via inhibition of p75 neurotrophin receptor expression in a rat model of diabetes [corrected]. *Diabetologia* 54: 669-680, 2011.
- 127 Pan Q, Ip FC, Ip NY, Zhu HX and Min ZD: Activity of macrocyclic jatrophone diterpenes from *Euphorbia kansui* in a TrkA fibroblast survival assay. *J Nat Prod* 67: 1548-1551, 2004.
- 128 Hur JY, Lee P, Kim H, Kang I, Lee KR and Kim SY: (-)-3,5-Dicaffeoyl-muco-quinic acid isolated from *Aster scaber* contributes to the differentiation of PC12 cells: through tyrosine kinase cascade signaling. *Biochem Biophys Res Commun* 313: 948-953, 2004.
- 129 Hines J, Groll M, Fahnestock M and Crews CM: Proteasome inhibition by fellutamide B induces nerve growth factor synthesis. *Chem Biol* 15: 501-512, 2008.
- 130 Joyner PM and Cichewicz RH: Bringing natural products into the fold - exploring the therapeutic lead potential of secondary metabolites for the treatment of protein-misfolding-related neurodegenerative diseases. *Nat Prod Rep* 28: 26-47, 2011.
- 131 Jeon SJ, Bak H, Seo J, Han SM, Lee SH, Han SH, Kwon KJ, Ryu JH, Cheong JH, Ko KH, Yang SI, Choi JW, Park SH and Shin CY: Oroxylin A Induces BDNF Expression on Cortical Neurons through Adenosine A2A Receptor Stimulation: A Possible Role in Neuroprotection. *Biomol Ther (Seoul)* 20: 27-35, 2012.
- 132 Cadigan KM: TCFs and Wnt/beta-catenin signaling: more than one way to throw the switch. *Curr Top Dev Biol* 98: 1-34, 2012.
- 133 Huang HC, Xu K and Jiang ZF: Curcumin-mediated neuroprotection against amyloid-beta-induced mitochondrial dysfunction involves the inhibition of GSK-3beta. *J Alzheimers Dis* 32: 981-996, 2012.
- 134 Zhang X, Yin WK, Shi XD and Li Y: Curcumin activates Wnt/beta-catenin signaling pathway through inhibiting the activity of GSK-3beta in APPswe transfected SY5Y cells. *Eur J Pharm Sci* 42: 540-546, 2011.
- 135 Xiong Z, Hongmei Z, Lu S and Yu L: Curcumin mediates presenilin-1 activity to reduce beta-amyloid production in a model of Alzheimer's Disease. *Pharmacol Rep* 63: 1101-1108, 2011.
- 136 Sagara Y, Vanhnasy J and Maher P: Induction of PC12 cell differentiation by flavonoids is dependent upon extracellular signal-regulated kinase activation. *J Neurochem* 90: 1144-1155, 2004.
- 137 Lin CW, Wu MJ, Liu IY, Su JD and Yen JH: Neurotrophic and cytoprotective action of luteolin in PC12 cells through ERK-dependent induction of Nrf2-driven HO-1 expression. *J Agric Food Chem* 58: 4477-4486, 2010.
- 138 Hoppe JB, Coradini K, Frozza RL, Oliveira CM, Meneghetti AB, Bernardi A, Pires ES, Beck RC and Salbego CG: Free and nanoencapsulated curcumin suppress beta-amyloid-induced cognitive impairments in rats: involvement of BDNF and Akt/GSK-3beta signaling pathway. *Neurobiol Learn Mem* 106: 134-144, 2013.
- 139 Zhao J, Cheng YY, Fan W, Yang CB, Ye SF, Cui W, Wei W, Lao LX, Cai J, Han YF and Rong JH: Botanical drug puerarin coordinates with nerve growth factor in the regulation of neuronal survival and neuritogenesis via activating ERK1/2 and PI3K/Akt signaling pathways in the neurite extension process. *CNS Neurosci Ther* 21: 61-70, 2015.
- 140 Lonze BE and Ginty DD: Function and regulation of CREB family transcription factors in the nervous system. *Neuron* 35: 605-623, 2002.
- 141 Marino G, Madoe F and Kroemer G: Autophagy for tissue homeostasis and neuroprotection. *Curr Opin Cell Biol* 23: 198-206, 2011.
- 142 Costa LG, Garrick JM, Roque PJ and Pellacani C: Mechanisms of Neuroprotection by Quercetin: Counteracting Oxidative Stress and More. *Oxid Med Cell Longev* 2016: 2986796, 2016.
- 143 Gabryel B, Kost A and Kasprowska D: Neuronal autophagy in cerebral ischemia - a potential target for neuroprotective strategies? *Pharmacol Rep* 64: 1-15, 2012.
- 144 Wu Y, Li X, Zhu JX, Xie W, Le W, Fan Z, Jankovic J and Pan T: Resveratrol-activated AMPK/SIRT1/autophagy in cellular models of Parkinson's disease. *Neurosignals* 19: 163-174, 2011.
- 145 Qu L, Liang X, Gu B and Liu W: Quercetin alleviates high glucose-induced Schwann cell damage by autophagy. *Neural Regen Res* 9: 1195-1203, 2014.
- 146 Marques SC, Oliveira CR, Pereira CM and Outeiro TF: Epigenetics in neurodegeneration: a new layer of complexity. *Prog Neuropsychopharmacol Biol Psychiatry* 35: 348-355, 2011.
- 147 Nicolai V, Lucarelli M and Fuso A: Environment, epigenetics and neurodegeneration: Focus on nutrition in Alzheimer's disease. *Exp Gerontol* 68: 8-12, 2015.
- 148 Mastroeni D, Grover A, Delvaux E, Whiteside C, Coleman PD and Rogers J: Epigenetic mechanisms in Alzheimer's disease. *Neurobiol Aging* 32: 1161-1180, 2011.
- 149 Rees TM and Brimijoin S: The role of acetylcholinesterase in the pathogenesis of Alzheimer's disease. *Drugs Today (Barc)* 39: 75-83, 2003.
- 150 Almasieh M, Zhou Y, Kelly ME, Casanova C and Di Polo A: Structural and functional neuroprotection in glaucoma: role of galantamine-mediated activation of muscarinic acetylcholine receptors. *Cell Death Dis* 1: e27, 2010.
- 151 Carradori S, D'Ascenzio M, Chimenti P, Secci D and Bolasco A: Selective MAO-B inhibitors: a lesson from natural products. *Mol Divers* 18: 219-243, 2014.
- 152 Cai Z: Monoamine oxidase inhibitors: promising therapeutic agents for Alzheimer's disease (Review). *Mol Med Rep* 9: 1533-1541, 2014.
- 153 Shoji Y and Nakashima H: Nutraceuticals and delivery systems. *J Drug Target* 12: 385-391, 2004.
- 154 Coimbra M, Isacchi B, van Bloois L, Torano JS, Ket A, Wu X, Broere F, Metselaar JM, Rijcken CJ, Storm G, Bilia R and Schiffflers RM: Improving solubility and chemical stability of natural compounds for medicinal use by incorporation into liposomes. *Int J Pharm* 416: 433-442, 2011.
- 155 Hodgson AB, Randell RK and Jeukendrup AE: The effect of green tea extract on fat oxidation at rest and during exercise: evidence of efficacy and proposed mechanisms. *Adv Nutr* 4: 129-140, 2013.

- 156 Neves AR, Lucio M, Lima JL and Reis S: Resveratrol in medicinal chemistry: a critical review of its pharmacokinetics, drug-delivery, and membrane interactions. *Curr Med Chem* 19: 1663-1681, 2012.
- 157 Sharma RA, Steward WP and Gescher AJ: Pharmacokinetics and pharmacodynamics of curcumin. *Adv Exp Med Biol* 595: 453-470, 2007.
- 158 Van der Schyf CJ, Geldenhuys WJ and Youdim MB: Multifunctional drugs with different CNS targets for neuropsychiatric disorders. *J Neurochem* 99: 1033-1048, 2006.
- 159 Krol S: Challenges in drug delivery to the brain: nature is against us. *J Control Release* 164: 145-155, 2012.
- 160 Gao H, Pang Z and Jiang X: Targeted delivery of nano-therapeutics for major disorders of the central nervous system. *Pharm Res* 30: 2485-2498, 2013.
- 161 Smith A, Giunta B, Bickford PC, Fountain M, Tan J and Shytle RD: Nanolipidic particles improve the bioavailability and alpha-secretase inducing ability of epigallocatechin-3-gallate (EGCG) for the treatment of Alzheimer's disease. *Int J Pharm* 389: 207-212, 2010.
- 162 Alam S, Panda JJ and Chauhan VS: Novel dipeptide nanoparticles for effective curcumin delivery. *Int J Nanomedicine* 7: 4207-4222, 2012.
- 163 Dhawan S, Kapil R and Singh B: Formulation development and systematic optimization of solid lipid nanoparticles of quercetin for improved brain delivery. *J Pharm Pharmacol* 63: 342-351, 2011.
- 164 Testa G, Gamba P, Badilli U, Gargiulo S, Maina M, Guina T, Calfapietra S, Biasi F, Cavalli R, Poli G and Leonarduzzi G: Loading into nanoparticles improves quercetin's efficacy in preventing neuroinflammation induced by oxysterols. *PLoS One* 9: e96795, 2014.
- 165 Tran TH, Guo Y, Song D, Bruno RS and Lu X: Quercetin-containing self-nanoemulsifying drug delivery system for improving oral bioavailability. *J Pharm Sci* 103: 840-852, 2014.
- 166 Watkins R, Wu L, Zhang C, Davis RM and Xu B: Natural product-based nanomedicine: recent advances and issues. *Int J Nanomedicine* 10: 6055-6074, 2015.
- 167 Pund S, Borade G and Rasve G: Improvement of anti-inflammatory and anti-angiogenic activity of berberine by novel rapid dissolving nanoemulsifying technique. *Phytomedicine* 21: 307-314, 2014.
- 168 Li B, Harich K, Wegiel L, Taylor LS and Edgar KJ: Stability and solubility enhancement of ellagic acid in cellulose ester solid dispersions. *Carbohydr Polym* 92: 1443-1450, 2013.
- 169 Li B, Konecke S, Harich K, Wegiel L, Taylor LS and Edgar KJ: Solid dispersion of quercetin in cellulose derivative matrices influences both solubility and stability. *Carbohydr Polym* 92: 2033-2040, 2013.
- 170 Li B, Konecke S, Wegiel LA, Taylor LS and Edgar KJ: Both solubility and chemical stability of curcumin are enhanced by solid dispersion in cellulose derivative matrices. *Carbohydr Polym* 98: 1108-1116, 2013.
- 171 Vaka SR, Sammeta SM, Day LB and Murthy SN: Delivery of nerve growth factor to brain *via* intranasal administration and enhancement of brain uptake. *J Pharm Sci* 98: 3640-3646, 2009.
- 172 van Woensel M, Wauthoz N, Rosiere R, Amighi K, Mathieu V, Lefranc F, van Gool SW and de Vleeschouwer S: Formulations for Intranasal Delivery of Pharmacological Agents to Combat Brain Disease: A New Opportunity to Tackle GBM? *Cancers (Basel)* 5: 1020-1048, 2013.
- 173 Panyam J and Labhasetwar V: Biodegradable nanoparticles for drug and gene delivery to cells and tissue. *Adv Drug Deliv Rev* 55: 329-347, 2003.
- 174 Loureiro JA, Gomes B, Coelho MA, do Carmo Pereira M and Rocha S: Targeting nanoparticles across the blood-brain barrier with monoclonal antibodies. *Nanomedicine (Lond)* 9: 709-722, 2014.
- 175 Megill A, Lee T, DiBattista AM, Song JM, Spitzer MH, Rubinshtein M, Habib LK, Capule CC, Mayer M, Turner RS, Kirkwood A, Yang J, Pak DT, Lee HK and Hoe HS: A tetra(ethylene glycol) derivative of benzothiazole aniline enhances Ras-mediated spinogenesis. *J Neurosci* 33: 9306-9318, 2013.
- 176 Agyare EK, Jaruszewski KM, Curran GL, Rosenberg JT, Grant SC, Lowe VJ, Ramakrishnan S, Paravastu AK, Poduslo JF and Kandimalla KK: Engineering theranostic nanovehicles capable of targeting cerebrovascular amyloid deposits. *J Control Release* 185: 121-129, 2014.
- 177 Burgess A, Nhan T, Moffatt C, Klivanov AL and Hynynen K: Analysis of focused ultrasound-induced blood-brain barrier permeability in a mouse model of Alzheimer's disease using two-photon microscopy. *J Control Release* 192: 243-248, 2014.
- 178 Yallapu MM, Jaggi M and Chauhan SC: Curcumin nanomedicine: a road to cancer therapeutics. *Curr Pharm Des* 19: 1994-2010, 2013.
- 179 Fomina N, McFearin C, Sermakdi M, Edigin O and Almutairi A: UV and near-IR triggered release from polymeric nanoparticles. *J Am Chem Soc* 132: 9540-9542, 2010.
- 180 Di J, Price J, Gu X, Jiang X, Jing Y and Gu Z: Ultrasound-triggered regulation of blood glucose levels using injectable nano-network. *Adv Healthc Mater* 3: 811-816, 2014.
- 181 Choonara YE, Kumar P, Modi G and Pillay V: Improving drug delivery technology for treating neurodegenerative diseases. *Expert Opin Drug Deliv*: 1-15, 2016.
- 182 Tofaris GK and Schapira AH: Neurodegenerative diseases in the era of targeted therapeutics: how to handle a tangled issue. *Mol Cell Neurosci* 66: 1-2, 2015.
- 183 Lee KW and Lee HJ: The roles of polyphenols in cancer chemoprevention. *Biofactors* 26: 105-121, 2006.
- 184 Kamaly N, Xiao Z, Valencia PM, Radovic-Moreno AF and Farokhzad OC: Targeted polymeric therapeutic nanoparticles: design, development and clinical translation. *Chem Soc Rev* 41: 2971-3010, 2012.

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